

# Anemonin improves intestinal barrier restoration and influences TGF-βI and EGFR signaling pathways in LPS-challenged piglets

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#### **Abstract**

The present study was aimed at investigating whether dietary anemonin could alleviate LPS-induced intestinal injury and improve intestinal barrier restoration in a piglet model. Eighteen 35-d-old pigs were randomly assigned to three treatment groups (control, LPS and LPS+anemonin). The control and LPS groups were fed a basal diet, and the LPS+anemonin group received the basal diet + 100 mg anemonin/kg diet. After 21 d of feeding, the LPS- and anemonin-treated piglets received i.p. administration of LPS; the control group received saline. At 4 h post-injection, jejunum samples were collected. The results showed that supplemental anemonin increased villus height and transepithelial electrical resistance, and decreased crypt depth and paracellular flux of dextran (4 kDa) compared with the LPS group. Moreover, anemonin increased tight junction claudin-1, occludin and ZO-1 expression in the jejunal mucosa, compared with LPS group. Anemonin also decreased TNF- $\alpha$ , IL-6, IL-8 and IL-1 $\beta$  mRNA expression. Supplementation with anemonin also increased TGF- $\beta$ 1 mRNA and protein expression, Smad4 and Smad7 mRNA expressions, and epidermal growth factor and epidermal growth factor receptor (EGFR) mRNA expression in the jejunal mucosa. These findings suggest that dietary anemonin attenuates LPS-induced intestinal injury by improving mucosa restoration, alleviating intestinal inflammation and influencing TGF- $\beta$ 1 canonical Smads and EGFR signaling pathways.

#### **Keywords**

Anemonin, intestinal injury, TGF- $\beta$ I, EGFR, piglets

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Introduction		. <sup>11</sup> 13 TGF-β1
Pulsatilla chinensis	$\mathbf{C}$	(EGFR)
C	. <sup>1,2</sup> T -	. <sup>14</sup> 16 TGF-β
	, -	
· · · · · · · · · · · · · · · · · · ·	_	14,17
. <sup>3,4</sup> A		T TGF-β S
.4 6 R	-	Animal Science College, Zhejiang University, The Key Laboratory of Molecular Animal Nutrition, Ministry of Education, Hangzhou, 310058, China
A , ,	. <sup>10</sup> H ,	Corresponding authors: Cai Hong Hu, Animal Science College, Zhejiang University, Yuhangtang Rd No. 866, Hangzhou 310058, P.R. China. Email: chhu@zju.edu.cn. Li Wang, institute of animal Science, Guangdong academy of agricultural sciences, Jinyin Rd No. 29, Guangzhou, 510640, P.R. China. Email: wanglineau@163.com.

	.18 T	TGF-β1	-
(EGF)	. <sup>16</sup> E_ EGFR		
15,19,20	Э Н ,	. ,	
TGF-β1	EGF,	F	-
W LPS-			
· . I	TGF-β1	EGFR , Escherichia coll	i LPS
T			. •
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## Materials and methods

### Animals and treatment

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E (	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	PS. ).
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. P (1.8 × 1.1 <sup>2</sup> ) . T 27°C. E	. T . D	25
N R	C (2012) (T 1). A 2 100 / T , (LPS LPS + E. coli LPS (E. coli	
055: B5, S 100 μ /	C , S . L , MO, USA) (BM) 0.9% N C	_
. T	LPS , L <sup>21</sup>	

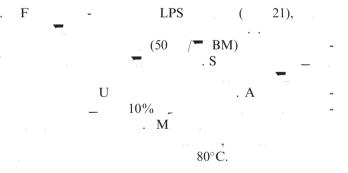
**Table 1.** Ingredients and chemical composition of the weaned diet on an as-fed basis.

Ingredients (g/kg)	Basal diet
Maize	573.5
Soybean meal	280
Fish meal	55
Dried whey	45
Limestone meal	5
Dicalcium phosphate	11.5
Sodium chloride	3
L-Lysine HCI	1.5
DL-Methionine	0.5
Vitamin-mineral premix <sup>a</sup>	10
Soybean oil	15
Composition (g/kg)	
Digestible energy (MJ/kg) <sup>b</sup>	14.40
Crude protein	229.6
Lysine	14.1
Methionine	4.6
Calcium	8.5
Total phosphorus	7.4

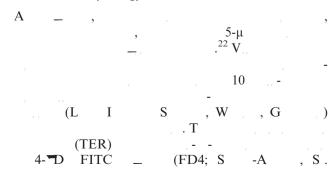
 $^{\rm a}$ Provided per kg of diet: vitamin A, 6000 IU; vitamin E, 50 IU; vitamin D\_3, 600 IU; vitaminK\_3, 1.5 mg; biotin, 0.10 mg; riboflavin, 8.0 mg; thiamine, 2.0 mg; niacin, 30 mg; pantothenic acid, 20 mg; pyridoxine, 3.0 mg; folic acid, 0.6 mg; vitamin B<sub>12</sub>, 0.04 mg; choline, 800 mg; Cu (CuSO<sub>4</sub>·5H<sub>2</sub>O), 16 mg; Fe (FeSO<sub>4</sub>), 125 mg; Zn (ZnSO<sub>4</sub>), 100 mg; Mn (MnSO<sub>4</sub>·H<sub>2</sub>O), 15 mg; Se (Na<sub>2</sub>SeO<sub>3</sub>), 0.3 mg; I (KI), 0.2 mg.

<sup>b</sup>Digestible energy was calculated from data provide by Feed Database in China (2011).

## Sample collection



# Intestinal morphology and barrier function



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L , MO, USA) in vitro U	Table 2. Fo	rward and reverse primers used for real-time PCR.
$M = \frac{1}{23} B$	Target	Primer sequence
· · · · · · · · · · · · · · · · · · ·	TNF-α	F:5'CATCGCCGTCTCCTACCA3'
$(95\% O_2/5\% CO_2) R$ , E M U		R:5'CCCAGATTCAGCAAAGTCCA3'
- · · · · · · · · · · · · · · · · · · ·	IL-6	F:5'CCTGTCCACTGGGCACATAAC3'
MC6; P I ). T -		R:5'CAAGAAACACCTGGCTCTGAAAC3'
(P	IL-8	F:5/TGGCAGTTTTCCTGCTTTCT 3/
I , S D , CA, USA)		R:5'CAGTGGGGTCCACTCTCAAT 3'
. A 30-	IL-Iβ	F:5'CAAAGGCCGCCAAGATATAA 3'
U , TER $(\Omega \cdot ^{2})$		R:5'GAAATTCAGGCAGCAACAT 3'
15- 2-	IFN-γ	F:5'GAGCCAAATTGTCTCCTTCTAC3'
TER . FD4 (S -		R:5'CGAAGTCATTCAGTTTCCCAG3'
A ) -	Τβ <b>R</b> I	F:5'CTGTGTCTGTCCACCATTCATTTG3'
0.375 / M		R:5'CAACTTTGCTATGTCTGTCTCCCC3'
FD4 ( $\mu / 2/1$ )	TβRII	F:5'CATCTCCTGCTAATGTTGTTGCC3'
30- 120 . T FD4		R:5′CGGTTCTAAATCCTGGGACACG3′
(FL_800; B -T <b>™</b> I I .,	Smad2	F:5'GAAGAGAAGTGGTGAGAAAGCAG3'
W , VT, USA). T _ 2-		R:5'AATACTGGAGGCAAAACTGGTGTC3'
w , v1, OSM). 1 = 2-	Smad3	F:5'TGGAGGAGGTGGAGAAATCAGAAC3'
		R:5'CACACTCGCTTGCTCACTGTAATC3'
Duetain automorphis and mic hi Mastern hist	Smad4	F:5'CCTGAGTATTGGTGTTCCATTGC3'
Protein expression analysis by Western blot		R:5/TGATGCTCTGCCTTGGGTAATC3/
T W	Smad7	F:5'TACTGGGAGGAGAGACGAGAGTG3'
$_{2}$ H $_{2}$ $^{22}$ B $_{3}$ , $_{4}$		R:5'TGGCTGACTTGATGAAGATGGG3'
41 Jay -	EGF	F:5'TGCCATAAGGGTGTCAGGTATTT3'
$(M_{\perp}, B_{\perp}, MA, MA)$		R:5/TGCTTTGCTCTTGCCCTCTAC3/
USA). T A 4°C 10 A	EGFR	F: 5'GGCCTCCATGCTTTTGAGAA 3'
2 T A [		R:5'GACGCTATGTCCAGGCCAA3'
-1, -1 (ZO-1), TGF-β1, β-	GAPDH	F:5'ATGGTGAAGGTCGGAGTGAAC3'
S C T I (S		R:5'CTCGCTCCTGGAAGATGGT3'
C , CA, USA). T A HRP		
- A (C S T		, . T
D , MA, USA). W		RNA .
· · . · . · . · . · . · . · . · . ·	N D	(ND-2000; N D
(A , A , H , IL, USA), -	Τ ,	, W. , DE, USA). R
C S 3400 (C - S)	/T I/ D I	P S RT
$I \qquad , \qquad S \qquad , \qquad C \qquad )$	(T K R I	$B \longrightarrow \{A, D, A, C, B, B,$
Q , Ο , β-Α	Q	PCR
. T		P - PCR (A
_ /β-		, F C , CA, USA) SYBR
T	G M	= (P , M , WI, USA),
,		, T -
		T 2. G
mRNA expression analysis by RT-PCR		- AAC
, ,		. T 2 ΔΔC
T RNA TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-8, IFN- $\gamma$ ,		<u> </u>
TGF- $\beta$ 1 TGF- $\beta$ , S (S 2, S 3, S 4	т	$(\Lambda)$ $C$
S (S 2, S 3, S 4 S 7), EGF EGFR RT-	T	$(\Delta)$ C $(\Delta C)$ GAPDH $(\Delta C)$ .
PCR, L .21 B , RNA	S	$\Delta \Delta \Delta CT$
TRI	~ .	.,
(I , C , CA, USA),		ACT . T

$$^{2}$$
 . T  $^{\Delta\Delta C}$  GAPDH  $_{-}$ 

Statistical analysis

# **Results**

## Intestinal morphology and barrier function

# Tight junction protein expression

**Table 3.** Effects of dietary anemonin on jejunum morphology and barrier function of piglets.

Item	Control	LPS	LPS + anemonin	SEM <sup>3</sup>
Villus height (μm)	381ª	343.2 <sup>b</sup>	368.5 <sup>a</sup>	5.15
Crypt depth (μm)	111.33	121.57	117.92	3.22
Villus height:crypt depth	3.42 <sup>a</sup>	2.84 <sup>c</sup>	3.31 <sup>b</sup>	0.06
TER $(\Omega \cdot cm^2)$	63.12 <sup>a</sup>	46.35 <sup>b</sup>	57.97 <sup>a</sup>	2.98
$FD_4$ flux $(\mu g/cm^{-2}/h^{-1})$	1.21°	2.59 <sup>a</sup>	1.54 <sup>b</sup>	0.10

FD4: fluorescein isothiocyanate dextran (4 kDa).

$$(P < 0.05)$$
 \_ LPS .

## Pro-inflammatory cytokine mRNA

T RNA (TNF-α, IL-6, IFN-γ, IL-1β, TGF-β1) T 4. I LPS (
$$P < 0.05$$
) RNA  $\_$  TNF-α, IL-1β, IL-6, IFN-γ IL-8 , LPS  $\_$  TGF-β1 RNA ( $P > 0.05$ ). D  $\_$  ( $P < 0.05$ ) TNF-α, IL-1β, IL-6 IL-8 , RNA TGF-β1 LPS .

## TGF- $\beta$ 1 expression

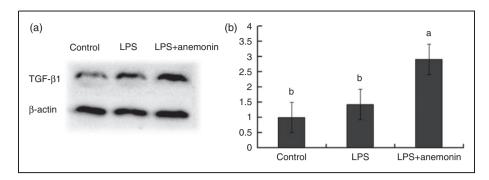
## mRNA expression of Smads

# mRNA expression of EGFR signal pathways

T 6 RNA \_ EGF EGFR . I 
$$P < 0.05) RNA = EGFR RNA$$
 EGFR RNA EGF EGFR RNA 
$$P < 0.05) RNA = EGFR RNA$$

 $<sup>^{</sup>a,b,c}$ Means within a row with different letters differ significantly (P < 0.05). Data are means of six pigs.

LPS . H , 100 - (P < 0.05) EGF EGFR RNA \_ LPS .



**Figure 2.** Effects of dietary anemonin on the protein expression of TGF- $\beta$ I in the jejunal mucosa of piglets. (a) Representative blots of TGF- $\beta$ I expression and  $\beta$ -actin. (b) Relative TGF- $\beta$ I protein expression in jejunal mucosa of piglets. <sup>a,b</sup>Mean values with different letters were significantly different (*P* < 0.05). Values are means and SD, represented by vertical bars. The control sample was used as the reference sample. The protein expression of all samples was expressed as fold changes, calculated relative to the control group. Control (non-challenged control), piglets receiving a control diet and injected with 0.9% sterile saline; LPS (LPS-challenged control), piglets receiving the same control diet and injected with *E. coli* LPS; LPS + anemonin (LPS challenged + 100 mg/kg anemonin), piglets receiving a 100 mg/kg anemonin diet and injected with LPS.

**Table 5.** Effects of dietary anemonin mRNA expressions of Smad signals on the jejunal mucosa of piglets.<sup>a</sup>

Item	Control	LPS	LPS + anemonin	SEM
ΤβRΙ	1.00	1.22	1.13	0.19
TβRII	1.00 <sup>c</sup>	1.42 <sup>c</sup>	2.81 <sup>b</sup>	0.28
Smad2	1.00	1.14	1.52	0.24
Smad3	1.00	1.75	1.23	0.28
Smad4	1.00 <sup>c</sup>	1.57 <sup>c</sup>	3.74 <sup>b</sup>	0.37
Smad7	1.00 <sup>c</sup>	1.36 <sup>c</sup>	4.33 <sup>b</sup>	0.38

<sup>&</sup>lt;sup>a</sup>The  $2^{-\Delta\Delta Ct}$  method was used to analyze the relative expression (fold changes), calculated relative to the values in samples from the control pigs. Data are means of six pigs.

**Table 6.** Effects of dietary anemonin on EGF and EGFR mRNA expression in the jejunal mucosa of piglets.<sup>a</sup>

Item	Control	LPS	LPS + anemonin	SEM <sup>2</sup>
EGF	1.00°	1.25°	2.69 <sup>b</sup>	0.29
EGFR	1.00 <sup>b</sup>	0.57 <sup>c</sup>	1.36 <sup>b</sup>	0.13

<sup>&</sup>lt;sup>a</sup>The  $2^{-\Delta\Delta Ct}$  method was used to analyze the relative expression (fold changes), calculated relative to the values in samples from the control pigs. Data are means of six pigs.

X

M

PRV

Pulsatilla

**LPS** 

 $<sup>^{</sup>b,c}$ Within a row with different letters differ significantly (P < 0.05)

 $<sup>^{</sup>b,c}$ Means within a row with different letters differ significantly (P < 0.05).

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35,36   1   (TNF-α, IL-1β, IL-6, IFN-γ   IL-8)     LPS.   C   (TNF-α, IL-1β, IL-8).   C   (TNF-α, IL-1β,	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$
Pulsatilla _ LPS-  (TNF-α, IL-1β, IL-6).8 T , -  , -	LPS-  LPS-  TGF-β1 S  ,
en e	. 13,15 I
TGF-β  TGF-β-	.15,40,41 A EGF/EGFR .42 44 EGF
. 14,16 I TGF-β1 _ , LPS	, 45 I
S , L .7 P. chinensis  TGF- $\beta$ 1	EGFR , EGFR LPS . T
. T , - TGF-β1. T TGF-β	M , EGF RNA LPS S . , L
TβR . W $TGF-β$	_ TGF-β1 EGFR. T , -
TGF-β II (TβRII), TGF-β I (ΤβRI). ΤβRII - ΤβRI, ΤβRI	I 100- /-
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<sup>38</sup> T TGF-β1 -	- - 1

LPS ); (3)	15. N RR, W BB W BW. R
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