

SCIENTIFIC REPORTS

OPEN

B cells Using Calcium Signaling for Specific and Rapid Detection of O157:H7

Received: 26 September 2014

Accepted: 20 April 2015

Published: 02 June 2015

Ling Wang^{1,2}, Ronghui Wang², Byung-Whi Kong³, Sha Jin⁴, Kaiming Ye⁴, Weihuan Fang⁵ & Yanbin Li^{1,2}

A rapid and sensitive detection technology is highly desirable for specific detection of O157:H7, one of the leading bacterial pathogens causing foodborne illness. In this study, we reported the rapid detection of O157:H7 by using calcium signaling of the B cell upon cellular membrane anchors anti-O157:H7 IgM. The binding of O157:H7 to the IgM on B cell surface activates the B cell receptor (BCR)-induced Ca^{2+} signaling pathway and results in the release of Ca^{2+} within seconds. The elevated intracellular Ca^{2+} triggers Fura-2, a fluorescent Ca^{2+} indicator, for reporting the presence of pathogens. The Fura-2 is transferred to B cells before detection. The study demonstrated that the developed B cell based biosensor was able to specifically detect O157:H7 at the low concentration within 10 min in pure culture samples. Finally, the B cell based biosensor was used for the detection of O157:H7 in ground beef samples. With its short detection time and high sensitivity at the low concentration of the target bacteria, this B cell biosensor shows promise in future application of the high throughput and rapid food detection, biosafety and environmental monitoring.

78 48 1 3. I 3,000 , Cam l bac e jej ni, E. c li
U S 4. I , E. c li O157:H7
Li e ia m n c gene , Salm nella en e ica, E. c li O157:H7
(-O157 STEC), Vib i .
5. E. c li O157:H7
(38%)⁶.
C - (CBB)
CBB
B , B , T , 16 24. A , B
BCR (B C R)
(P -2E9)
Bacill 13,25,26. R e al.¹¹ Li e ia, 25. M B O

¹College of Biosystems Engineering and Food Science, Zhejiang University, Hangzhou 310058, China. ²Department of Biological and Agricultural Engineering, University of Arkansas, Fayetteville, AR 72701, USA. ³Department of Poultry Science, University of Arkansas, Fayetteville, AR 72701, USA. ⁴Department of Biomedical Engineering, Binghamton University-SUNY, Binghamton, NY 13902, USA. ⁵College of Animal Sciences, Zhejiang University, Hangzhou 310058, China. Correspondence and requests for materials should be addressed to Y.L. (email: yanbinli@zju.edu.cn)

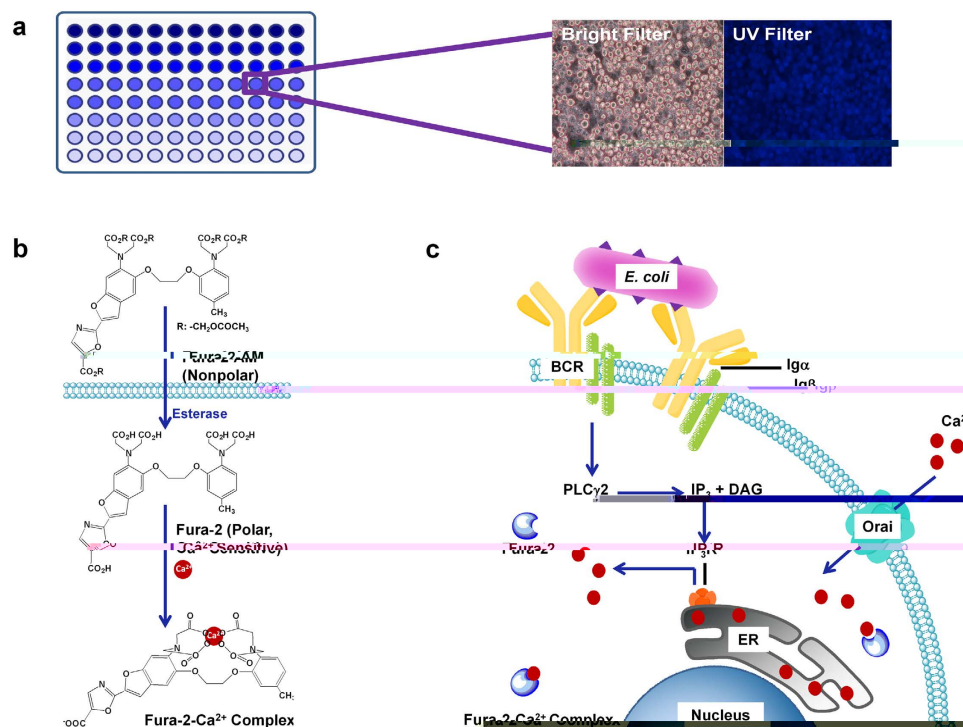


Figure 1. P

Figure 1. P (a), S (b), O (c).

AM C²⁺ F -2- (C²⁺ B - B (BCR) C²⁺ F -2 340/380

5 11. B C²⁺ A F -2 B I C²⁺ B C²⁺ F -2 340/380

E. coli O157:H7 (500 CFU/ <1)^{27,28} BCR²⁹ 30,31 C²⁺ 32,33 I 34 36

F -2 B I C²⁺ F -2 340/380

E. coli O157:H7 BCR²⁹ C²⁺ 31 I C²⁺ C²⁺ I C²⁺ 30

E. coli O157:H7

Results

Principles of the B cell biosensor. C^6 , C^{2+} , B, F, -2, C^{2+} , $E. coli$ O157:H7, A, F, -1, F, -2, B, 96-, -, (AM)

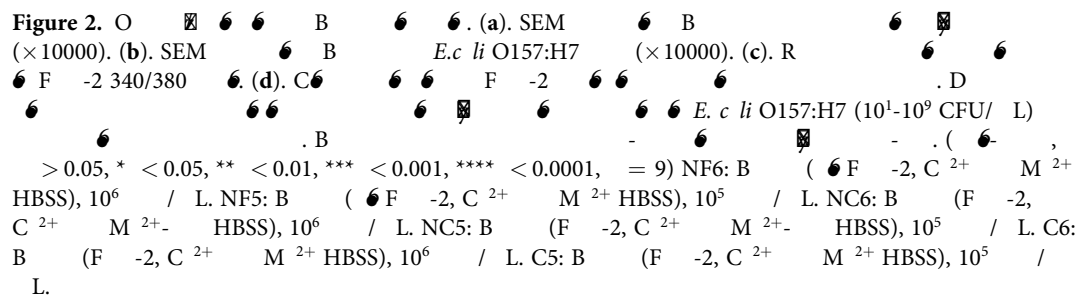
AM C²⁺ F -2- (C²⁺), BCR- C²⁺ (F . 1). *E. c li* O157: H7. BCR- C²⁺ BCR²⁵ (I) BCR F . 1 , B I α I β. C BCR (IP₃R), 40. BCR- PLC-γ2 4,5- (PIP₂) IP₃ (IP₃), 1,4,5- (IP₃) (ER) (ER) O , (CRAC), CRAC 29,42,43. IP₃ C²⁺ ER C²⁺ C²⁺ C²⁺ 44 46.

Optimization of the B cell biosensor.

B C²⁺ (MA) LPS *E. c li* O157:H7⁴⁷. U ELISA (E I) ELISA B (S) ELISA (F . S1) 12 B . I B (*E. c li* O157:H7 B 5 12) B (SEM). F -2 (FR) 1 . A 10³CFU/ L *E. c li* O157:H7, 0 . A F . 2 , (HBSS) (*E. c li* O157:H7, . I (C_{min}) C²⁺- (C_{ma}) C_{min} C_{ma} , FR C_{n l} (F . 2) 0 60 32. T₆ C²⁺ C²⁺ C²⁺ *E. c li* O157:H7 (10¹-10⁹ CFU/ L). P F . 2 . I NF (F -2), B F -2, NF5 (F -2, 10⁵ / L) (< 0.0001). I NC (C²⁺), B (> 0.05) F -2 C²⁺ M²⁺ C6 (C²⁺, 10⁶ / L) S F -2 10⁶ / L B , 10⁶ / L B F -2 NF6 NC6 (C²⁺, 10⁶ / L) (= 0.0324) C6 (C²⁺, 10⁶ / L) (< 0.0001). A C²⁺ M²⁺ (= 0.0006) C6, C²⁺ M²⁺

Detection of O157:H7 in pure culture.

E. c li O157:H7 10¹ 10⁵ CFU/ L B HBSS *E. c li* O157:H7. C *E. c li* O157:H7 10²-10⁵ CFU/ L. A F . 3 10¹ 10³ CFU/ L. A = 0.0565 + 0.6753 (R² = 0.96). B (10¹-10⁷ CFU/ L), (F . 3). A LPS LPS = 0.03183 + 0.6532 (R² = 0.83).



S

B

E. coli O157:H7

(O157 -O157)

E

E. coli

2

EHEC (-O157),

(F .3).

7

6

-

S

T

S5.

Li e ia m n c gene, S. T

V. aahaem l ic

-

B

(MA)

LPS

E. coli O157

N Salm nella

I M

(C

B

(L , NH, USA)

BE0087)

MARC 29F8. M

29F8

12- 16- D

LPS⁴⁷. *L. m n c gene*

V. aahaem l ic

G

-

O-

A

S. T

N Salm nella

S. g de be g

-

I (

I:

). S. T

D

(F .3).

F -2

E. coli O157:H7 ATCC 43888

30

ROC

ROC (AUR)

0.7319, 0.7690, 0.8484, 0.7817

0.7885,

E. coli O157:H7

10¹, 10², 10³, 10⁴

10⁵ CFU/ L. A

AUR

E. coli O157:H7

0.7

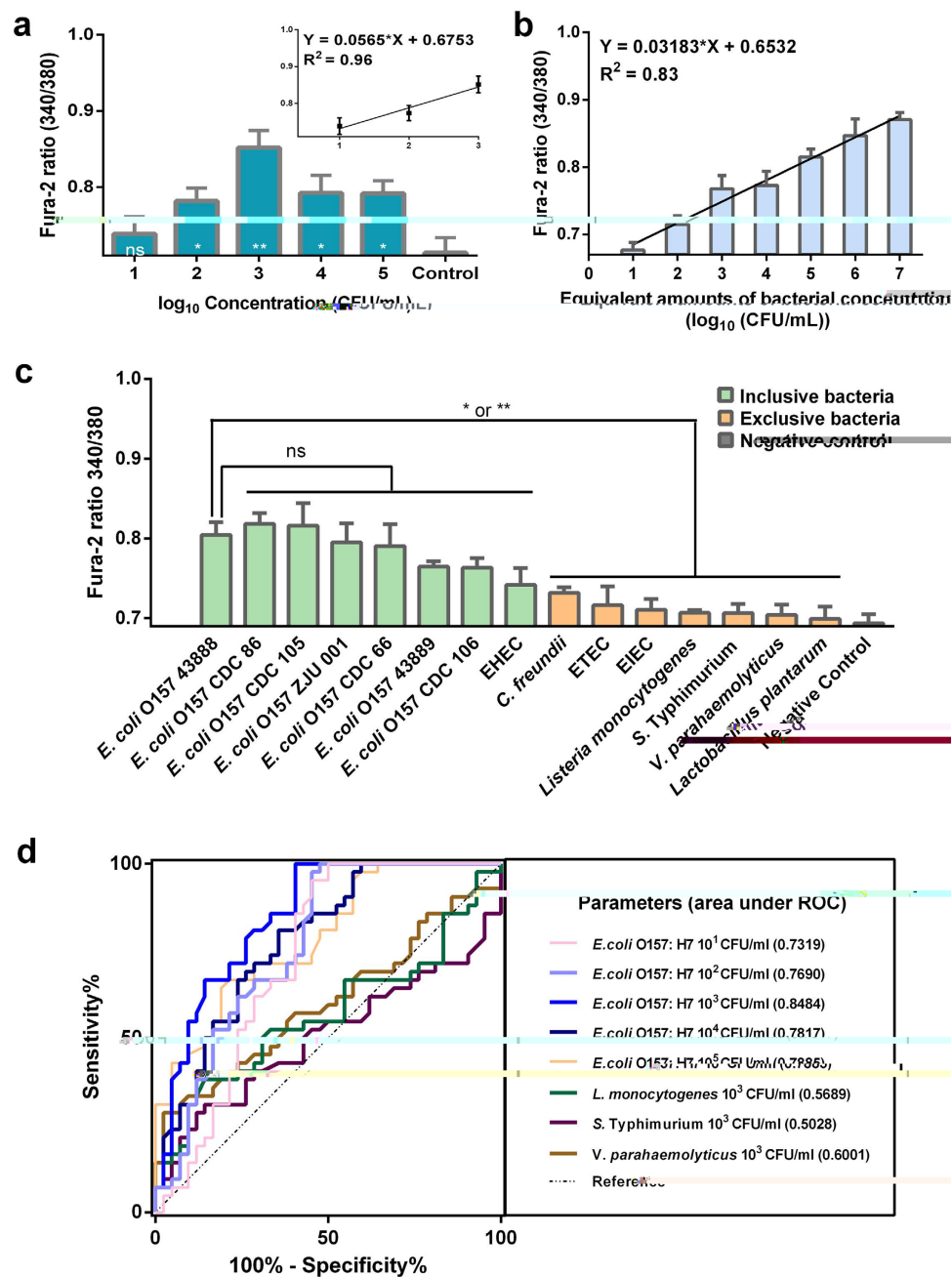


Figure 3. D *E. coli* O157:H7. (a). P, F, -2, I, L, 10¹, 10³ CFU/ L. A, ±, . . ., > 0.05, * < 0.05, ** < 0.01, = 4). (b). P, F, -2, I, L, 10¹, 10⁷ CFU/ L. A, ±, . . ., > 0.05, * < 0.05, ** < 0.01, = 3). (c). I, B, 10³ CFU/ L. A, ±, . . ., > 0.05, * < 0.05, ** < 0.01, = 3). (d). ROC, 30.

0.7319 0.8484, B 0.7817 0.7885 *E. coli* O157:H7
AUR 10⁴ CFU/ L 10⁵ CFU/ L, 10³ CFU/ L *L. monocytogenes*, *S. Typhimurium*
V. alba hemolytic, AUR 0.5689, 0.5028 0.6001,

Detection of

I *E. coli* O157:H7, B *E. coli* O157:H7 (10¹-10³ CFU/L). L). *E. coli* O157:H7 10⁴ CFU/L 10⁵ CFU/L, *E. coli* BCR- C²⁺ T. B, LPS *E. coli* O157:H7 B LPS (10¹-10⁷ CFU/L). I T I *E. coli* O157:H7, B O- LPS G - MA B EHEC (O-157) G - L. m n c gene Lac bacill lan a m, I F . 3, (S) T S5), *E. coli* O157:H7 B ROC F . 3, B 10 *E. coli* O157:H7 10¹ CFU/L, *E. coli* O157:H7 10²-10⁵ CFU/L. F CFU/L. A B 10 . I 5.9 × 10² (SPR)⁵², (ELISA)⁵³, (QCM)⁵⁵. *E. coli* O157:H7 7 CFU (SPCE) 70 56, 10 CFU/L (LRSP-FS) 40 57, 67 CFU/L (MNP) 8 58, 500 CFU/L CANAR (5 11. I 30 μL 18 *E. coli* O157:H7. I B L. 8.6 × 10² CFU/L (26 CFU 30 μL) *E. coli* O157:H7 B C²⁺ F -2 *E. coli* O157:H7 B C²⁺ F -2 T B MARC 29 8 C²⁺ F -2 *E. coli* O157:H7 C²⁺ BCR C²⁺ F -2 B 10² CFU/L B *E. coli* O157:H7 10

Methods

Reagents. D (DMEM), D (FBS), H (HBSS), C²⁺ M²⁺ HBSS, MEM (MEM NAA), 0.4% T B S, F -2/AM 0.5 M EDTA BD (S, MD, USA). G (CA, USA). A I M-HRP (), L (LPS) (G - K) S B (S, C), A B (S, L, MO, USA). A -8 (B, MA, USA).

B cell lines and culture conditions. B MARC 29F8 ATCC (A T C C, M A; ATCC CRL-2508). B DMEM 4 M L- 1.5 /L , 4.5 /L , 1 % MEM NAA 10% - FBS. 37 C 5 10) 7% CO₂ L - - MARC 29F8 (72 . MARC 29F8 1:10 10% - FBS T-25 T-75 (F, O, USA) (72) C (B-R, H, CA, USA) TC10 A C .

Bacterial strains and culture conditions. B A T C (ATCC), C C I C (CICC), C N C M C (CMCC), C U (JU). D C P (P CDC) S T S1 T S2. *Lac bacill lan a m* MRS BHI 37 C, - *Li e ia m n c gene* 48 24 , S A (TSA). S I .

Scanning electron microscopy (SEM) imaging SEM I () H TM1000 (SEM) (H, J).

Preparation of a B cell biosensor. B C²⁺ - F -2, C²⁺ - MARC 29F8 HBSS DMEM (PR) 37 C 5 . B 4.5 F -2/AM 10⁶ 30 37 C 30 L HBSS . A F -2/AM, C²⁺ M²⁺ HBSS 30 L HBSS

Detection of O157:H7 in pure culture. B 30 μ F -2 MARC 29F8 96- 30 μ (340 T 380) F -2 MARC 29F8 0.1% T -100 4.5 M (F340 (EDTA), F 380- 380- (B T , F380) 510 S H1 H M -M M) D 120 . D 340/380 C²⁺30. *E. c li* O157:H7 ATCC 43888 HBSS 6.2×10¹ 6.2×10⁹ 5.9×10¹ 5.9×10⁵ LPS *E. c li* O157:H7 ATCC 43888 B . E T 8 7 S I . 10³ CFU/ L (S T S1 T S2). *E. c li* O157:H7 ATCC 43888 HBSS F -2 B

Detection of O157:H7 in ground beef.

I., Q., C. (S. 400 (S. 225, N. 1, UK) 10³, 10⁴, 10⁵ CFU/ L. *E. coli* O157:H7 HBSS 10², 10³, 10⁴ CFU/ L. *E. coli* O157:H7 HBSS 10², 10³, 10⁴ CFU/ L.

Statistical analysis.

Optimization of the B cell biosensor

References

1. S , E. & M , B. E. F , D A N (F66 N) 2012:
- U S . Clin. Infec . Di . 54 S 5, S381 384 (2012).
2. S , E., G , P. M., A , F. J., T , . & H , . M. F U S
- . Emeg. Infec . Di . 17, 16 22 (2011).
3. S , E. e al. F U S . Emeg. Infec . Di . 17, 7 15 (2011).
4. S , L. E U S . J. F d P . 75, 123 131 (2012).
5. A -C , T. e al. D DNA Talan a 101, 405 412 (2012).
6. S M. C e al. I , 10 U.S. S , 2006 2013. M b. M al. Wee l e . 63, 328 332 (2014).
7. A , P, S , A., D , N. & C A. B . Bi en . Bi elec n. 28, 1 12 (2011).
8. , D. M., , A. J. & T , A. P. , . Bi en 4, 299 311 (1989).
9. N , T. e al. G fabA' fab : Anal. Chem. 78, 4952 4956 (2006).
10. F , T. e al. L Bi elec n. 21, 2263 2269 (2006).
11. , T. H. e al. A B Science 301, 213 215 (2003).
12. M , M., , B , L. G. & A , D Anal. Chem. 76, 4156 4161 (2004).
13. M , e al. C Lab. In e . 85, 1429 1439 (2005).
14. L , Q. e al. D Bi en . Bi elec n. 22, 3224 3229 (2007).
15. B , P, L D., , J. P., , J. L. & B , A. A Lab. In e . 88, 196 206 (2008).
16. B , P. & B , A. M Tend Bi echm l. 27, 179 188 (2009).
17. C , L.-E., C , S., , & , M. P Elec chim. Ac a 53, 6690 6697 (2008).
18. E , A. , M , T. H. & P , P. S. B Ad . A l. Mic bi l. 64, 137 166 (2008).
19. H , T. C : C Anal. Chim. Ac a 568, 211 216 (2006).
20. , S. E. C - F n. D g De . Di c e 2, 225 239 (2005).
21. , G. T. E P. IEEE 91, 915 929 (2003).
22. , S. M. e al. A Li e ia m n c gene : HT-29 In . J. F d Mic bi l. 68, 33 44 (2001).
23. C , T, N , .M. , B , C., T , J. & H , D. D Bi en . Bi elec n. 23, 1024 1031 (2008).
24. L , B., L, P. & C , B. T. A Sen . Ac a . B-Chem. 114, 559 564 (2006).
25. S , A. e al. L BC Imm ni 17, 451 462 (2002).
26. B , A. & , D. A Li e ia Le . A l. Mic bi l. 26, 305 310 (1998).
27. , T. CANA , Bi . Mic . and Nan em C nfe ence: BMN' 06, S F N USA: IEEE. (2006, J . 15-18)
28. P , M. S. e al. Linc ln Lab. J. 17, 63 84 (2007).
29. , L. B. & F , B. D. B- Imm n l. e . 231, 265 277 (2009).
30. G , G., P , M. & T , A J. Cell Bi l. Chem. 260, 3440 3450 (1985).
31. N , E. -2 C Ne ha mac l g 34, 1423 1442 (1995).
32. M , A., M , D., M , J. & P , T. F -2 C + J. Cell Bi l. 105, 2145 2155 (1987).

33. H , A. H , C. H , & L , D. G. *Calcium Signaling Pathways*, Vol. 312 (L , D.G.) C . 2, 37 45 (H P , 2006).
34. P , A. B. & P J. S - . *Phil. e.* **85**, 757 810 (2005).
35. , U. B. & S . *Phil. e.* **82**, 769 824 (2002).
36. S , D., L. , & E , T. P P2 . *Fun. Cell. Neurosci.* **8**, 1 18 (2014).
37. H , F., I . & S , B. C ²⁺ C ²⁺ . *Biophys. J.* **70**, 1069 1081 (1996).
38. N , J., M , M., E -D , G. C. & , H. S - , NMDA C ²⁺ . *Neuron* **46**, 609 622 (2005).
39. G , C. & , A. I . *Neuron* **73**, 862 885 (2012).
40. H , J., T., L , L., S , H. & , M. M B- . *EMBO J.* **21**, 83 92 (2002).
41. , . *al. BAN* BC - . *EMBO J.* **21**, 83 92 (2002).
42. P , M. *al. O* 1 C AC . *Neuron* **443**, 230 233 (2006).
43. F , S. & P , M. C STIM1 . *Nat. Sci. M. l. Bi l.* **20**, 918 919 (2013).
44. O - , M. & , A. C . *Cell. Immunol.* **20**, 250 258 (2008).
45. F , S. C . *Nat. Rev. Immunol.* **7**, 690 702 (2007).
46. , S. G. & B , S . C . *J. Biol. Chem.* **272**, 15045 15048 (1997).
47. , B., H , J. E., L , E. T. & , J. P . *Eur. J. Clin. Microbiol.* **35**, 679 684 (1997).
48. L , O., S , A. & O . *Bacteriol. Rev.* **30**, 192 255 (1966).
49. , C. A. *al. I* CD20 B C . *J. Biol. Chem.* **283**, 16971 16984 (2008).
50. A , A. O. *al. C* -2. *Science* **285**, 736 739 (1999).
51. F , S. *al. B* . *Science* **312**, 738 741 (2006).
52. O , B- , B , L , & C , J.-D . *Eur. J. Clin. Microbiol.* **12**, 780 786 (2002).
53. , N., H , M. & S , H.-C. N (ELISA) T E. c li . *Anal. Chim. Acta* **590**, 224 231 (2007).
54. G , T., U , J., T., S.-I. & B , A. . F -A -F . *Sensory* **6**, 796 807 (2006).
55. S , -L. & L., A - . *H7. Biophys. J.* **19**, 563 574 (2004).
56. S , E. B. & A , E. C . *Biophys. J.* **26**, 2208 2214 (2011).
57. H , C.-J., D , J., S , A. & , L . E c li O157: H7. *Anal. Chem.* **83**, 674 677 (2011).
58. L , *al. N* . *IEEE Trans. Nanotechnol.* **11**, 676 681 (2012).

Acknowledgements

I (ABI) C M S T L D R K SEM A B (P N. 2013BAD19B02).

Author Contributions

Other Contributions


Additional Information

Supplementary information [6](#) :// . . [6](#) /

Competing financial interests: ☐ ☐ ☐

How to cite this article: , L. e al. B U C S S R
D *E che ichia c* D O157:H7. *Sci. Re.* 5, 10598; : 10.1038/10598 (2015).



 CC BY 4.0 I L C -