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B cells Using Calcium Signaling for Specific and Rapid Detection of O157:H7

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A rapid and sensitive detection technology is highly desirable for specific detection of O157:H7, one of the leading bacterial pathogens causing foodborne illness. In this study, we reported the rapid detection of O157:H7 by using calcium signaling of the B cell upon cellular membrane anchors anti-O157:H7 IgM. The binding of O157:H7 to the IgM on B cell surface activates the B cell receptor (BCR)-induced Ca²⁺ signaling pathway and results in the release of Ca²⁺ within seconds. The elevated intracellular Ca²⁺ triggers Fura-2, a fluorescent Ca²⁺ indicator, for reporting the presence of pathogens. The Fura-2 is transferred to B cells before detection. The study demonstrated that the developed B cell based biosensor was able to specifically detect O157:H7 at the low concentration within 10 min in pure culture samples. Finally, the B cell based biosensor was used for the detection of O157:H7 in ground beef samples. With its short detection time and high sensitivity at the low concentration of the target bacteria, this B cell biosensor shows promise in future application of the high throughput and rapid food detection, biosafety and environmental monitoring.

78 U S 48 1 3. I 4. I 3,000 , Cam . l o b a c e j e j n i , E . c o l i O157:H7 (-O157 STEC), V i b i o . 5. E . c o l i O157:H7 (38%)⁶. C - (CBB) 8,9, 10 11 15. . B CBB , B , , T , 16 24. A , B - BCR (B C R) T 25. M - (P -2E9) B a c i l l . 13,25,26. R e . a l .¹¹ L i . e i a , B O

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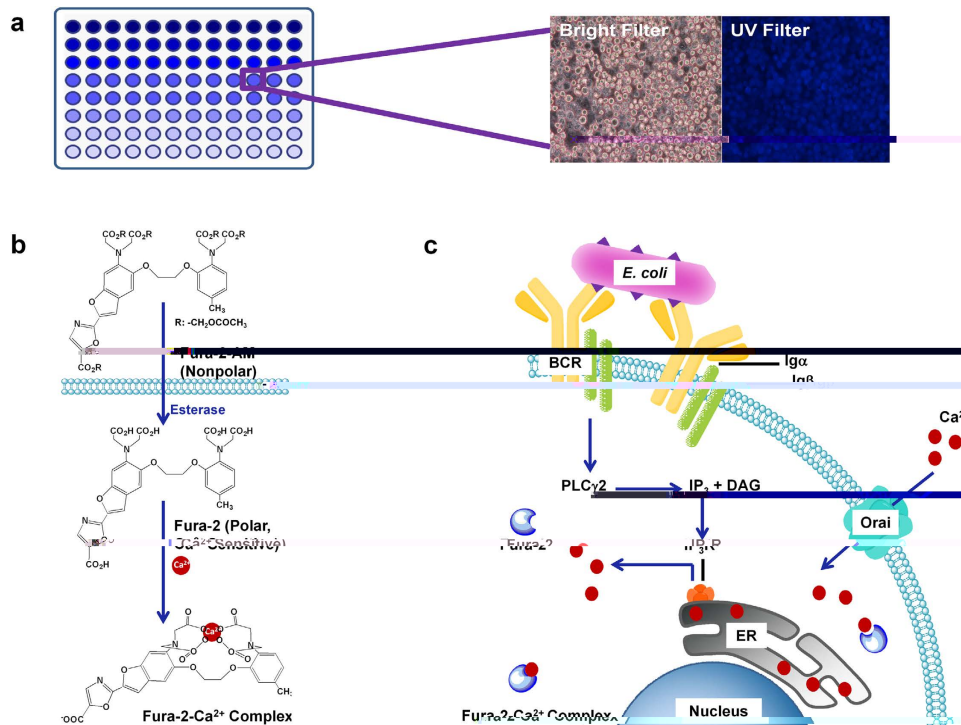
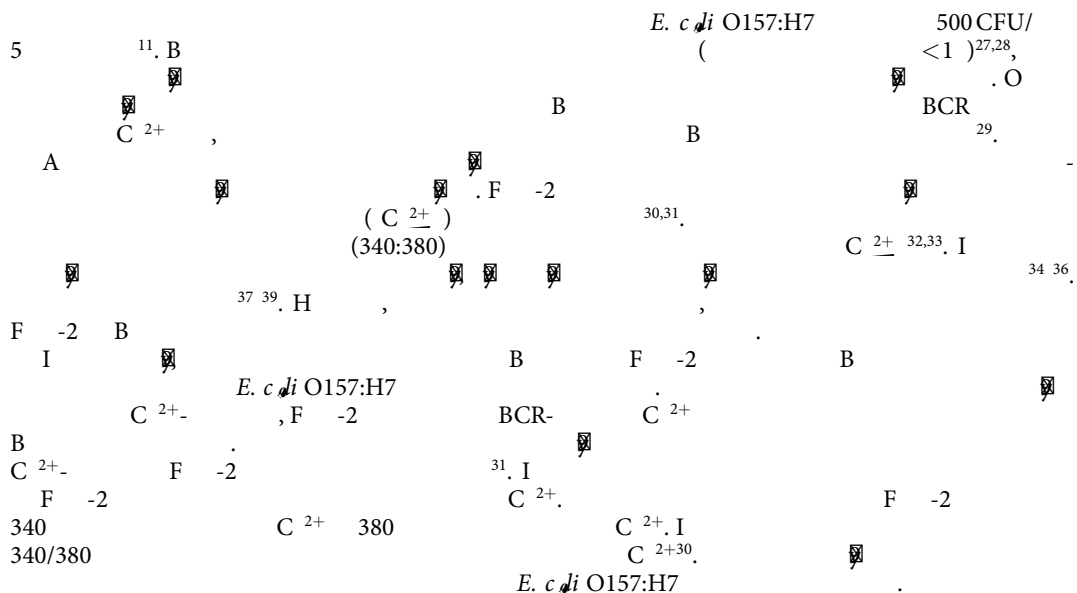


Figure 1. (a) Schematic of the biosensor array. (b) Chemical synthesis of Fura-2 from Fura-2-AM (Nonpolar) via an Esterase step to Fura-2 (Polar), which then forms a Fura-2-Ca²⁺ complex. (c) Schematic of the B cell biosensor mechanism showing *E. coli* interaction with BCR, signaling through PLCγ2 to produce IP₃ and DAG, leading to Ca²⁺ release from the ER and nuclear translocation of the Fura-2-Ca²⁺ complex.



Results

Principles of the B cell biosensor.

E. coli O157:H7 (340:380) binds to B cells (A, B, C²⁺), leading to Ca²⁺ release from the ER (F-2) and nuclear translocation of the Fura-2-Ca²⁺ complex (AM). The resulting fluorescence is detected by the biosensor array.

AM C²⁺ F⁻² (C²⁺),
 BCR- C²⁺ (F⁻¹).
 BCR-²⁹ A BCR²⁵ (I) I α I β. C BCR
 40. BCR- PLC-γ2
 1,4,5- (IP₃) 4,5- (PIP₂)
 (IP₃R), (ER) IP₃
 41. ER C²⁺ (CRAC), CRAC
 C²⁺ ER C²⁺ 29,42,43. IP₃
 44 46.

Optimization of the B cell biosensor.

B C²⁺ B MARC 29F8
 (MA) LPS *E. coli* O157:H7⁴⁷. U
 ELISA (E I) ELISA B
 (S) F (S1) 12 B I B (SEM).
 (*E. coli* O157:H7 B 5 12) B B
 R F⁻² (FR) F⁻² 1 A 10³CFU/
 L *E. coli* O157:H7 0 A F⁻² (HBSS
 FR), *E. coli* O157:H7, I
 FR C²⁺ (C_{min}) C²⁺ (C_{ma}) R
 F⁻² FR C²⁺ (F⁻²)
 0 60 F⁻²
 T C²⁺ C²⁺
 B *E. coli* O157:H7 (10¹-10⁹ CFU/ L). P
 F⁻² I NF (F⁻², B F⁻²,
 NF6 (F⁻², 10⁶ / L)
 NF5 (F⁻², 10⁵ / L) (< 0.0001). I NC (C²⁺), B (> 0.05)
 F⁻² C²⁺ M²⁺ C (C²⁺), B
 NF5 (F⁻², 10⁵ / L) NF6. I C6 (C²⁺, 10⁶ / L)
 10⁶ / L B C5 (C²⁺, 10⁵ / L) (= 0.0025). S F⁻²
 L) (< 0.0001). A NC6 (C²⁺, 10⁶ / L) (= 0.0324) C6 (C²⁺, 10⁶ /
 C²⁺ M²⁺ (= 0.0006) NC6
 M²⁺ C²⁺ C²⁺

Detection of O157:H7 in pure culture.

E. coli O157:H7 10¹ 10⁵ CFU/ L B
 HBSS *E. coli* O157:H7. C
 F⁻³ *E. coli* O157:H7 10²-10⁵ CFU/ L. A
 10¹ 10³ CFU/ L. A
 = 0.0565 + 0.6753 (R² = 0.96).
 B (10¹-10⁷ CFU/ L), LPS
 (F⁻³). A LPS = 0.03183 + 0.6532 (R² = 0.83).

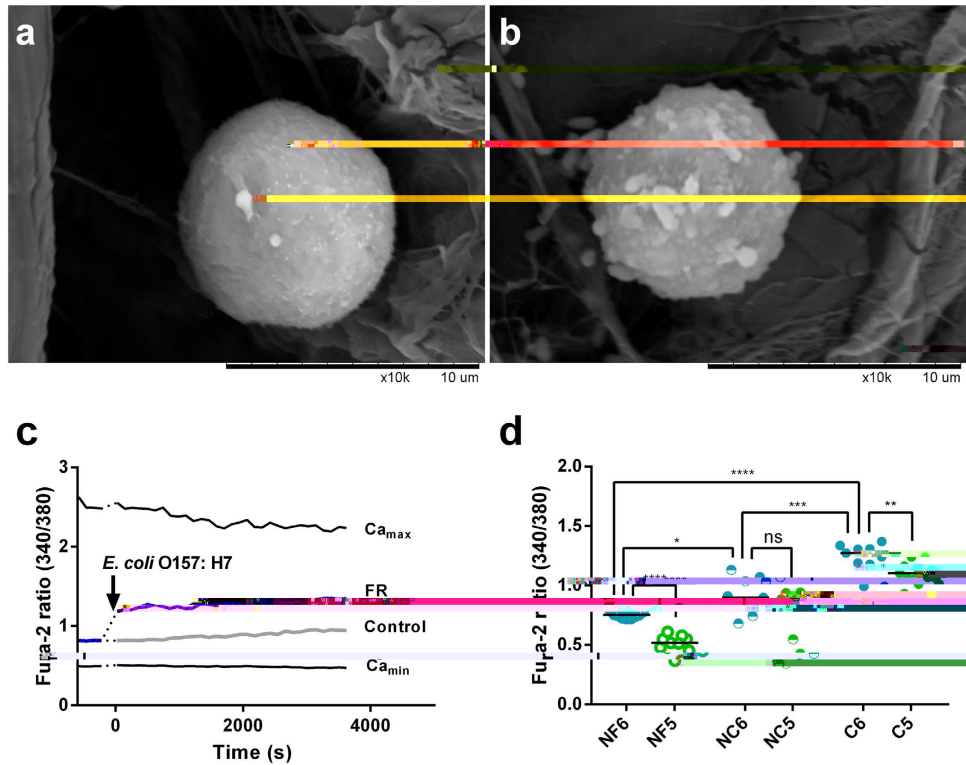


Figure 2. (a). SEM image of *E. coli* O157:H7 (×10000). (b). SEM image of *E. coli* O157:H7 (×10000). (c). Fluorescence traces showing Fura-2 ratio (340/380) over time (s) for *E. coli* O157:H7 (FR), Control, Ca_{max} , and Ca_{min} . (d). Scatter plot of Fura-2 ratio (340/380) for different conditions: NF6, NF5, NC6, NC5, C6, and C5. Statistical significance is indicated by asterisks (*, **, ***, ****) and NS (not significant).

E. coli O157:H7 (10¹-10⁹ CFU/ L) (F⁻², C²⁺ M²⁺ HBSS), 10⁶ / L. NF5: B (F⁻², C²⁺ M²⁺ HBSS), 10⁵ / L. NC6: B (F⁻², C²⁺ M²⁺ HBSS), 10⁶ / L. NC5: B (F⁻², C²⁺ M²⁺ HBSS), 10⁵ / L. C6: B (F⁻², C²⁺ M²⁺ HBSS), 10⁶ / L. C5: B (F⁻², C²⁺ M²⁺ HBSS), 10⁵ / L.

S. *E. coli* O157:H7 (O157) -O157), EHEC (O157), (F⁻², C²⁺ M²⁺ HBSS), 10⁶ / L. S. *Li e i a m n a c . g e n e*, S. T. *V. a a h a e m d . i c .* (MA) N *S a l m o n e l l a* I M (C) LPS *E. coli* O157 (NH, USA) MARC 29F8. M 29F8 LPS *L. m n a c . g e n e* *V. a a h a e m d . i c .* G O- *N S a l m o n e l l a* *S. g a d e b e g* I () S. T. (F⁻², C²⁺ M²⁺ HBSS), 10⁶ / L. F⁻² *E. coli* O157:H7 ATCC 43888 ROC (ROC) 0.7319, 0.7690, 0.8484, 0.7817 0.7885, *E. coli* O157:H7 10¹, 10², 10³, 10⁴ 10⁵ CFU/ L. A *E. coli* O157:H7 0.7 AUR

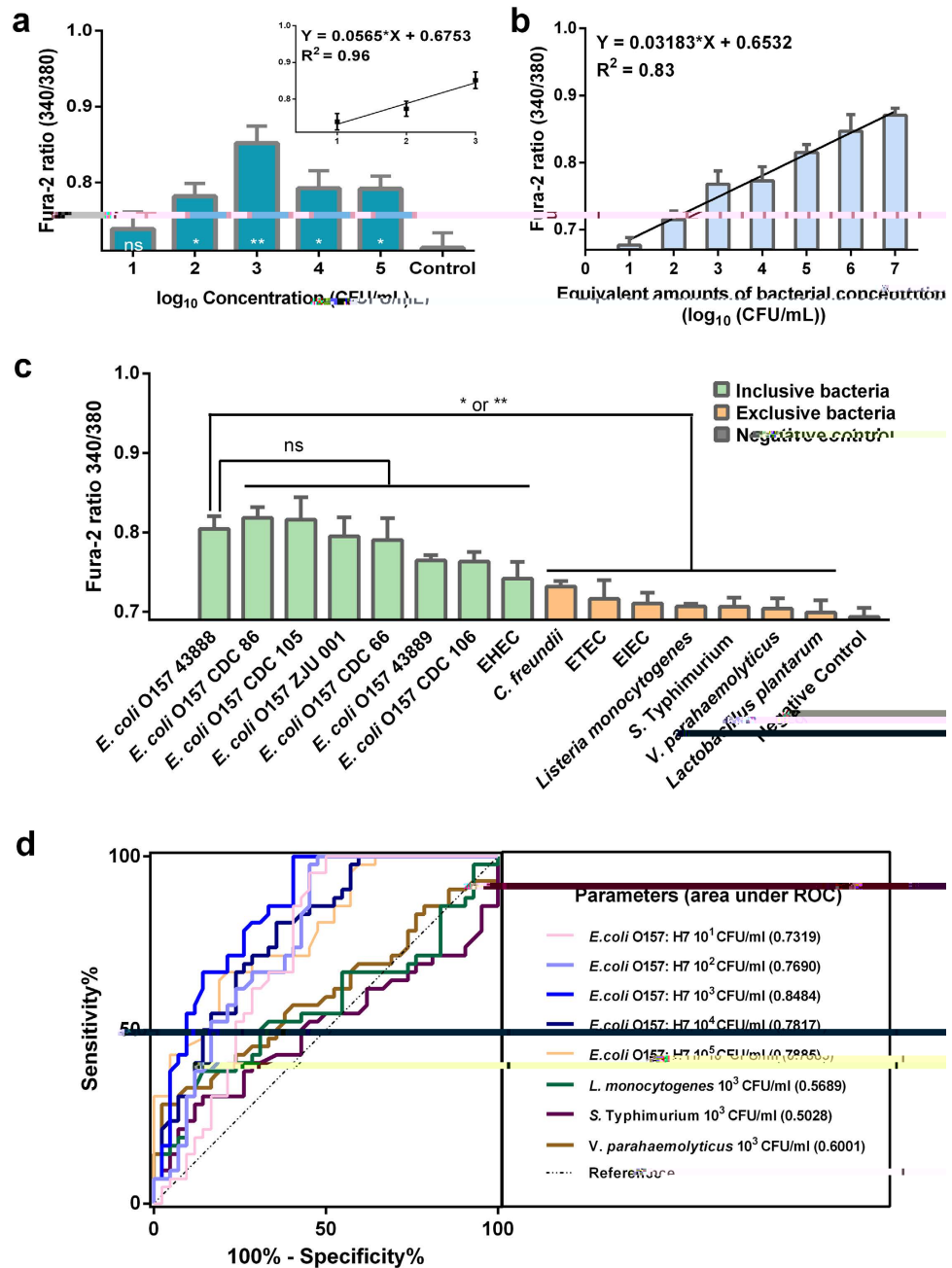


Figure 3. D. *E. coli* O157:H7. (a). P, F_{ura-2}, L. A. *E. coli* O157:H7, 10¹ 10⁷ CFU/ L. A. *E. coli* O157:H7, LPS, *E. coli* O157:H7, 10¹ 10⁷ CFU/ L. A. (b). P, F_{ura-2}, L. A. *E. coli* O157:H7, 10¹ 10⁷ CFU/ L. A. (c). I, F_{ura-2}, L. A. *E. coli* O157:H7, 10¹ 10⁷ CFU/ L. A. (d). ROC, Sensitivity, 100% - Specificity, AUR, 0.7319, 0.8484, 0.7817, 0.7885, 0.5689, 0.5028, 0.6001.

0.7319 0.8484, B, I, *E. coli* O157:H7, 10⁴ CFU/ L, 10⁵ CFU/ L, AUR, 0.5689, 0.5028, 0.6001,

Detection of

I *E. coli* O157:H7, B *E. coli* O157:H7 (10¹-10³ CFU/ L). *E. coli* O157:H7 10⁴ CFU/ L 10⁵ CFU/ L, BCR- C²⁺ *E. coli* O157:H7, T, B, A, F .3, LPS (10¹-10⁷ CFU/ L). I T 51.

I *E. coli* O157:H7, B LPS O-157) G (S) (T S5), *E. coli* O157:H7 L. monocytogene *Lac bacill. lan a m*, EHEC (I F .3, B ROC F .3, B 10 10¹ CFU/ L, 10 30 *E. coli* O157:H7 10²-10⁵ CFU/ L, F CFU/ L. A 5.9 × 10² (SPR)⁵², (QCM)⁵⁵. *E. coli* O157:H7 (SPCE) 70 56, 10 CFU/ L (LRSP-FS) 40 57, 67 CFU/ L (MNP) 8 58, 500 CFU/) 5 11. I CANAR (30 μL 18 *E. coli* O157:H7. I B L. F CFU/ L (26 CFU 30 μL) B *E. coli* O157:H7 8.6 × 10² A C²⁺ - B C²⁺ F -2 *E. coli* O157:H7 B F -2. T B, B I B MARC 29 8 C²⁺ - F -2 *E. coli* O157:H7 C²⁺ BCR C²⁺ F -2 B 10² CFU/ L B *E. coli* O157:H7 10

Methods

Reagents. D₁₀ (DMEM), D₁₉ (DMEM PR), D₂₀ (FBS), H₁ (HBSS), C²⁺ M²⁺-HBSS, MEM (MEM NAA), 0.4% TBS, F₁-2/AM 0.5 M EDTA (BD (S₁, MD, USA). G₁ I M-HRP (A D (A D S₁, USA), I B (LPS) (G₁ S₁, K₁) S₁ B (S₁, C₁), A₁ S₁ (S₁ L₁, MO, USA). A₁ M D -8 (B₁, MA, USA).

B cell lines and culture conditions. B₁ MARC 29F8 ATCC (A₁ T₁ C₁ C₁, M₁, A₁; ATCC CRL-2508). B₁ DMEM, 4 M L₁ I M. I₁ 1.5 /L₁, 4.5 /L₁, 1% MEM NAA 10% FBS. 37°C 5 10 7% CO₂ L₁ - - MARC 29F8 (72 . MARC 29F8 1:10 10% - FBS T-25 T-75 (F₁, O₁, USA) (72) C (B -R₁, H₁, CA, USA) TC10 A₁ C

Bacterial strains and culture conditions. B₁ A₁ T₁ C₁ (ATCC), C₁ C₁ I₁ C₁ C₁ (CICC), C₁ N₁ C₁ M₁ C₁ C₁ (CMCC), C₁ C₁ C₁ D₁ C₁ P₁ S₁ (P₁ CDC) U₁ (JU). *Lac bacill lan a m* MRS BHI 37°C, *Li e ia m n a c . gene* 48 24 , S₁ S₁ A₁ (TSA).

Scanning electron microscopy (SEM) imaging SEM I () H₁ TM1000 (S₁, J₁).

Preparation of a B cell biosensor. B₁ C²⁺ B₁ C²⁺ - F₁ -2, C²⁺ MARC 29F8 F₁ -2³³. I₁ , C²⁺ HBSS DMEM (PR) 37°C 5 . B₁ 4.5 F₁ -2/AM 10⁶ PR) 30 37°C 15 30 L HBSS . A₁ F₁ -2/AM, C²⁺ M²⁺- HBSS

Detection of O157:H7 in pure culture. B₁ 30 μ F₁ -2 MARC 29F8 96- , 30 μ (340 380) F₁ -2 0.1% T₁ -100 4.5 M₁ (F340 (EDTA), F₁ 340- 380- (B T , F380) 510 S₁ H₁ H₁ M₁ -M₁ M₁) 120 . D₁ E. coli O157:H7 ATCC 43888 HBSS 6.2 × 10¹ 6.2 × 10⁹ 5.9 × 10¹ 5.9 × 10⁵ LPS E. coli O157:H7 ATCC 43888 B₁ T₁ 8 7 S₁ I₁ 10³ CFU/ L 43888 HBSS (S₁ T₁ S₁ T₁ S₂). E. coli O157:H7 ATCC F₁ -2 B

Detection of *E. coli* O157:H7 in ground beef. T B

E. coli O157:H7
225 HBSS F (L
I., Q., C.) S 400 (S, N, UK) 1
E. coli O157:H7 10³, 10⁴, 10⁵
CFU/ L 9
E. coli O157:H7 10², 10³, 10⁴ CFU/ L.
E. coli O157:H7 HBSS
F -2 B

Statistical analysis. D ± . . . ()
G P P (G P, S D, CA). P
Optimization of the B cell biosensor
E. coli O157: H7 U
< 0.05

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