

# Zinc oxide influences mitogen-activated protein kinase and TGF- $\beta$ 1 signaling pathways, and enhances intestinal barrier integrity in weaned pigs

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## Abstract

Weaning is the most significant event in the life of pigs and is always related with intestinal disruption. Although it is well known that zinc oxide (ZnO) exerts beneficial effects on the intestinal barrier, the mechanisms underlying these effects have not yet been fully elucidated. We examined whether ZnO protects the intestinal barrier via mitogen-activated protein kinases and TGF- $\beta$ 1 signaling pathways. Twelve barrows weaned at 21 d of age were randomly assigned to two treatments (0 versus 2200 mg Zn/kg from ZnO) for 1 wk. The results showed that supplementation with ZnO increased daily gain and feed intake, and decreased postweaning scour scores. ZnO improved intestinal morphology, as indicated by increased villus height and villus height:crypt depth ratio, and intestinal barrier function, indicated by increased transepithelial electrical resistance and decreased mucosal-to-serosal permeability to 4-ku FITC dextran. ZnO decreased the ratios of the phosphorylated to total JNK and p38 (p-JNK/JNK and p-p38/p38), while it increased the ratio of ERK (p-ERK/ERK). Supplementation with ZnO increased intestinal TGF- $\beta$ 1 expression. The results indicate that supplementation with ZnO activates ERK  $\frac{1}{2}$ , and inhibits JNK and p38 signaling pathways, and increases intestinal TGF- $\beta$ 1 expression in weaned pigs.

## Keywords

Zinc oxide, intestinal barrier, mitogen-activated protein kinases, transforming growth factor- $\beta$ 1, weanling pigs

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## Introduction

Because of their immature immune and digestive systems, young animals are susceptible to immunologic challenge.<sup>1</sup> Pigs subjected to early weaning stress exhibit more severe diarrhea and intestinal disruption compared with late-weaned pigs.<sup>2</sup> This results mainly from the activation of the mucosal immune system, initiated by bacteria, endotoxins and antigens.<sup>2</sup> The activated immune systems up-regulates the expression of pro-inflammatory cytokines in the intestine, such as TNF- $\alpha$ , IL-6, IL-1 $\beta$  and INF- $\gamma$ .<sup>3,4</sup> The inflammatory response and subsequent overproduction of pro-inflammatory cytokines induces a disrupted intestinal barrier function.<sup>1,4</sup>

It is well known that zinc exerts anti-inflammatory and anti-diarrheal effects, as well as intestinal barrier protection in animal models and clinical trials.<sup>5</sup> The beneficial role of zinc in the epithelial barrier has been

shown both *in vitro* and *in vivo*.<sup>6,7</sup> In particular, zinc oxide (ZnO) appears to exert a strong effect in resistance to intestinal diseases.<sup>6</sup> Early weaned piglets frequently experience intestinal barrier dysfunction, post-weaning diarrhea and growth retardation.<sup>4</sup> Supplementation with a high level of ZnO (between 2000 and 4000 mg/kg of zinc) in weanling piglets ameliorates diarrhea and improves piglet growth performance.<sup>8,9</sup> Although ZnO is reported to maintain mucosal epithelial integrity and the normal function

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of intestine in weanling piglets,<sup>8–10</sup> the molecular mechanisms have not yet been elucidated.

MAPK cascades transduce signals from a diverse array of extracellular stimuli and form a highly integrated network to regulate cell growth and cell differentiation, especially stress responses.<sup>11</sup> Our recent study showed that weaning stress activated MAPK signaling pathways in the intestine, which may be an important mechanism of weaning-associated enteric disorders of piglets.<sup>4</sup> Furthermore, TGF- $\beta$ 1 is also believed to play an important regulatory role in the post-weaning adaptation process in the intestine of the pig.<sup>12</sup> The canonical TGF- $\beta$  signaling effectors are the SMAD family proteins, which transduce signals from the cell surface directly to the nucleus to regulate target gene transcription.<sup>13</sup> However, research on zinc-regulated MAPK signaling and TGF- $\beta$ 1 expression has only so far been reported in *in vitro* cell models.<sup>13,14</sup> Few data are available regarding the effect of ZnO *in vivo*. This study was conducted to assess the impact of ZnO supplementation on intestinal barrier function and to explore whether ZnO supplementation could improve intestinal barrier function through regulation of the MAPK and TGF- $\beta$ 1 signaling pathways.

## Materials and methods

### Experimental design and sample collection

The experiment was approved by the Animal Care and Use Committee, Zhejiang University. Twelve crossbred (Duroc  $\times$  Landrace  $\times$  Yorkshire) barrows were weaned at 21 d of age (body mass  $6.1 \pm 0.4$  kg) and randomly assigned to two groups based on body weight and litter of origin. Weanling piglets were fed a diet containing 0 or 2200 mg Zn/kg from ZnO. Diets were formulated to meet or exceed National Research Council requirements for all nutrients (Table 1).<sup>15</sup> All piglets were individually housed in  $1.3 \times 1.2$  m<sup>2</sup> pens in an environmentally controlled nursery barn.<sup>16</sup> Each treatment had six replicate pens. The piglets were given access to feed and water *ad libitum*. Feed intake was recorded daily. The feeding trial lasted 1 wk. Average daily gain (ADG), average daily feed intake (ADFI) and feed conversion efficiency were calculated.

The animals' feces were observed every day, and the scour score, according to Hu et al.,<sup>8,9</sup> for each pig was recorded according. The fecal scoring scale was from 1 to 5, where 1 represented no diarrhea, hard feces; 2 represented no diarrhea, normal consistency of feces; 3 represented mild diarrhea, soft and partially formed feces; 4 represented moderate diarrhea, semi-liquid feces; and 5 represented severe diarrhea, watery feces.

After the feeding trial (d 8 post-weaning), all the piglets were killed and the gastrointestinal tract quickly removed. Intestinal segments measuring 10 cm in length

**Table 1.** Composition of the basal diet (on an as-fed basis).

Ingredients (g/kg)	
Maize	560.0
Soybean meal	294.0
Fish meal	55.0
Dried whey	45.0
Soybean oil	15.0
Limestone meal	5.0
Dicalcium phosphate	11.5
Sodium chloride	3.0
L-Lysine HCl	1.0
D,L-Methionine	0.5
Vitamin-mineral premix <sup>a</sup>	10.0
Analyzed composition (g/kg)	
Digestible energy <sup>b</sup> (MJ/kg)	14.4
Crude protein	215.3
Lysine	14.6
Methionine	3.8
Calcium	9.1
Total phosphorus	7.3
Zn (mg/kg)	127.9

<sup>a</sup>Provided per kg of diet: vitamin A, 6000.00 IU; vitamin D<sub>3</sub>, 500.00 IU; vitamin E, 40.00 IU; pantothenic acid, 20.00 mg; niacin, 30.00 mg; riboflavin, 5.00 mg; vitamin K<sub>3</sub>, 1.50 mg; thiamine, 2.00 mg; pyridoxine, 3.00 mg; biotin, 0.10 mg; folic acid, 0.60 mg; vitamin B<sub>12</sub>, 0.04 mg; choline, 800.00 mg; Cu (CuSO<sub>4</sub>·5H<sub>2</sub>O), 16.00 mg; Zn (ZnSO<sub>4</sub>), 100.00 mg; Fe (FeSO<sub>4</sub>), 125.00 mg; Mn (MnSO<sub>4</sub>·H<sub>2</sub>O), 15.00 mg; I (KI), 0.20 mg; Se (Na<sub>2</sub>SeO<sub>3</sub>), 0.30 mg.

<sup>b</sup>Digestible energy was calculated from data provided by the Feed Database in China.

were excised from the proximal jejunum, immediately placed in Ringer's solution and mounted in Ussing chambers, as described by Hu et al.<sup>9</sup> The specimens from the mid-jejunum were excised and fixed in 10% formalin. The mucosal samples from the mid-jejunum were harvested by scraping with a glass slide, rapidly frozen in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  for further analysis.

### Jejunal morphology and barrier function

The method for making hematoxylin and eosin slices of jejunal tissue, and measurements of villus height and crypt depth, was performed according to the procedures of Hu et al.<sup>8</sup> The transepithelial electrical resistance (TER) and mucosal-to-serosal permeability to 4-ku FITC dextran (FD4; Sigma-Aldrich, St. Louis, MO, USA) were determined *in vitro* in the Ussing chamber, according to the procedures outlined by Hu et al.<sup>4,9</sup> Briefly, the jejunum segments were mounted in the EasyMount Ussing chamber system connected to Acquire and Analyse software (VCC MC6;

Physiologic Instruments, San Diego, CA, USA). The TER ( $\Omega \cdot \text{cm}^2$ ) was recorded at 15-min intervals over a 2-h period and averaged to obtain the TER values for a given pig. Meanwhile, mucosal-to-serosal flux of FD4 ( $\text{ng}/\text{cm}^2/\text{h}$ ) was determined. The FD4 was added on the mucosal side, and the concentration of FD4 in the serosal side was analyzed using a fluorescence microplate reader (FLx800; Bio-Tek Instruments, Winooski, VT, USA).

### mRNA expression analysis by RT-PCR

The mRNA levels of TGF- $\beta$ 1 and TGF- $\beta$  receptors (T $\beta$ R), as well as their downstream signals, were determined by RT-PCR, as described by Liu et al.<sup>16</sup> and Chen et al.<sup>17</sup> Briefly, total RNA was extracted from jejunal mucosa using TRIzol reagent (Invitrogen, Carlsbad, CA, USA) following the manufacturer's guidelines. The purity and concentration of all RNA samples were measured using a Nano Drop spectrophotometer (ND-2000; NanoDrop Technologies, Wilmington, DE, USA). Reverse transcription using the PrimeScript RT reagent kit (TaKaRa Biotechnology, Dalian, China) was carried out following the manufacturer's instructions. Quantitative analysis of PCR was carried out on a StepOne Plus real-time PCR system (Applied Biosystems, Foster City, CA, USA) using SYBR Green Master mix (Promega, Madison, WI, USA) according to the manufacturer's instructions. The primers used are given in Table 2. Gene-specific amplification was determined by melting curve analysis and agarose gel

electrophoresis. The 2- $\Delta\Delta\text{Ct}$  method was used to analyze the relative changes in each target gene expression.<sup>18</sup> The change ( $\Delta$ ) in Ct values in each group was compared with the Ct value of GAPDH ( $\Delta\text{Ct}$ ). Subsequently,  $\Delta\Delta\text{CT}$  was computed for each target gene from the ZnO groups by subtracting the average  $\Delta\text{CT}$  for the control group. The final fold differences were computed as 2- $\Delta\Delta\text{Ct}$  for each target gene. All samples were run in triplicate. The results showed that GAPDH exhibited no difference between two groups.

### Western blot analysis

The Western blot analysis was performed according to the procedures outlined by Hu et al.<sup>4</sup> The following primary Abs were used: MAPK Abs, rabbit mAb; phospho-p38 (Thr180/Tyr182) rabbit mAb; phospho-ERK 1/2 (Thr202/Tyr204) rabbit mAb; phospho-JNK (Thr183/Tyr185) rabbit mAb; TGF- $\beta$ 1 rabbit mAb (all from Cell Signaling Technology, Danvers, MA, USA). The secondary Ab was HRP-conjugated anti-rabbit antibody (Cell Signaling Technology). Western blot was detected with an enhanced chemiluminescence detection kit (Amersham, Arlington Heights, IL, USA), photographed by a ChemiScope 3400 (Clinx Science Instruments, Shanghai, China) and analyzed using Quantity One software.  $\beta$ -Actin was used as an internal control, and exhibited no difference between the groups. The relative expression of TGF- $\beta$ 1 was expressed as the ratio of TGF- $\beta$ 1 and  $\beta$ -actin. The MAPK signal pathway activation was calculated as the ratios of the phosphorylation levels (p-JNK,

**Table 2.** Primer sequences used for RT-PCR.

Primer name	Primer sequence	Size (bp)	Accession numbers
TGF- $\beta$ 1	F:5'GGACCTTATCCTGAATGCCTT3' R:5'TAGGTTACCACTGAGCCACAAT3'	133	NM214015
T $\beta$ R I	F:5'CTGTGTCTGTCCACCATTCATTG3' R:5'CAACTTTGCTATGTCTGTCTCCCC3'	496	AF461808
T $\beta$ R II	F:5'CATCTCCTGCTAATGTTGTTGCC3' R:5'CGGTTCTAAATCCTGGGACACG3'	324	X70142
Smad2	F:5'GAAGAGAAGTGGTGTGAGAAAGCAG3' R:5'AATACTGGAGGCAAACTGGTGTC3'	428	BP437096
Smad3	F:5'TGGAGGAGGTGGAGAAATCAGAAC3' R:5'CACACTCGCTTGCTCACTGTAATC3'	541	AB052738
Smad4	F:5'CCTGAGTATTGGTGTTCATTGC3' R:5'TGATGCTCTGCCTTGGGTAATC3'	598	NM 214072
Smad7	F:5'TACTGGGAGGAGAAGACGAGAGTG3' R:5'TGGCTGACTTGATGAAGATGGG3'	241	AW359979
GAPDH	F:5'ATGGTGAAGGTCGGAGTGAAC3' R:5'CTCGCTCCTGGAAGATGGT3'	235	NM001206359

bp: base pair.

p-p38, p-ERK) and the total levels of MAPKs (JNK, p38, ERK 1/2).

### **Statistical analysis**

Data were analyzed using the SAS statistical package (SAS Institute, Cary, NC, USA),<sup>19</sup> with each animal considered an experimental unit. Results are expressed as mean  $\pm$  SD. The fecal score data were tested using the Mann–Whitney U-test. Differences between the other means were analyzed with an unpaired t-test. Differences were considered significant at  $P < 0.05$ .

## **Results**

### **Growth performance and diarrhea index**

The effects of ZnO on growth performance and the diarrhea index of piglets after weaning are shown in Table 3. Feeding high levels of ZnO (2200 mg Zn/kg in diet) to weanling piglets significantly improved ( $P < 0.05$ ) ADG and ADFI, and significantly reduced ( $P < 0.05$ ) the post-weaning scour scores compared with the control.

### **Jejunal morphology and barrier function**

The weanling piglets fed with ZnO had significantly higher ( $P < 0.05$ ) villus height and villus height:crypt depth ratio at the jejunal mucosa compared with the control pigs (Table 4). The TER and mucosal-to-serosal flux of FD4 in the Ussing chamber were used to assess the effects of ZnO on the intestinal barrier function of pigs. The TER values were significantly increased ( $P < 0.05$ ) and the mucosal-to-serosal fluxes of FD4 were significantly decreased ( $P < 0.05$ ) in the jejunum of weanling pigs supplemented with ZnO compared with the control pigs (Table 4).

### **MAPK signaling pathways**

Figure 1 shows the effects of ZnO on the three MAPK signaling pathways (JNK, p38, ERK). There was no difference in the total protein levels for the MAPKs between the two groups. However, compared with the

control, supplemental ZnO significantly decreased ( $P < 0.05$ ) the ratios of the phosphorylated forms and the total levels of JNK and p38 (p-JNK/JNK and p-p38/p38), while significantly increasing ( $P < 0.05$ ) the ratio of the phosphorylated forms and the total levels of ERK (p-ERK/ERK).

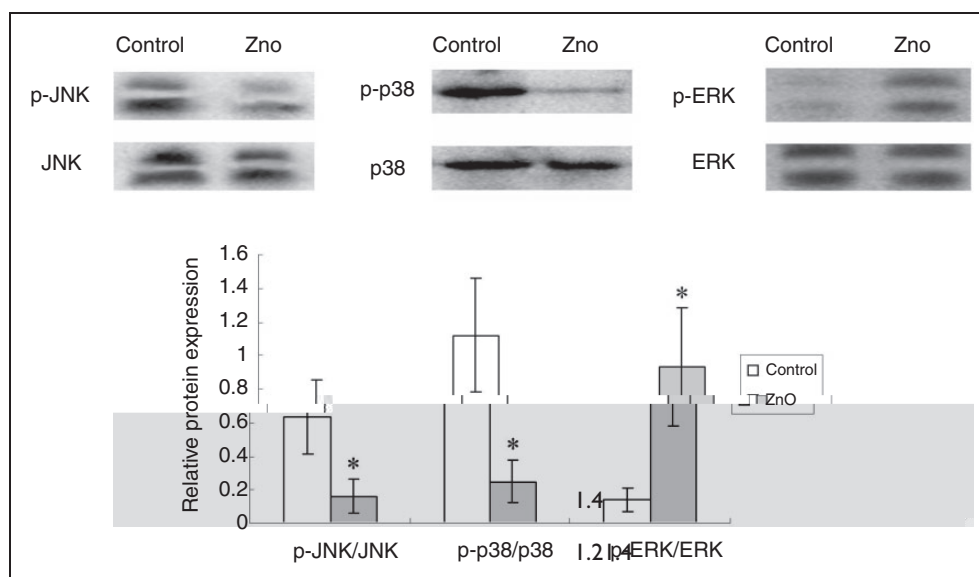
### **TGF- $\beta$ 1 expression and its canonical SMAD signaling pathway**

The mRNA level (Table 4) and expression (Figure 2) of TGF- $\beta$ 1 in jejunal mucosa was significantly increased ( $P < 0.01$ ) in piglets supplemented with ZnO compared with the control pigs. However, the mRNA levels of the T $\beta$ Rs (T $\beta$ R, T $\beta$ R II) and the downstream signals (SMAD 2, SMAD 3, SMAD7) showed no difference between the two groups (Table 5).

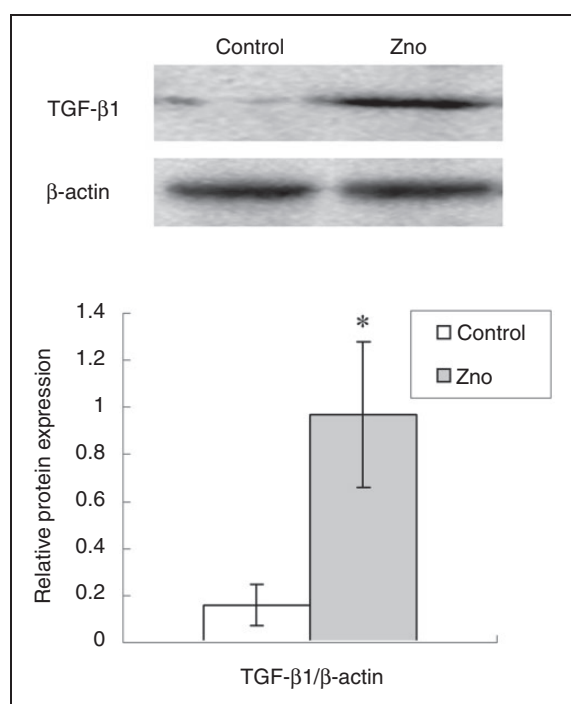
## **Discussion**

Weaning is one of the most stressful events in the life of piglets as they are abruptly forced to experience combined stressors, such as shifting from a milk-based diet to a cereal-based one, removal from the sow and litter-mates, fighting and establishment of the social hierarchy.<sup>4</sup> Post-weaning diarrhea and growth retardation are frequent occurrences in early-weaned piglets.<sup>4,10</sup> The duration and severity of diarrhea can be controlled by feeding pharmacological levels of ZnO (between 2000 and 4000 mg/kg of Zn) to the weanling piglets.<sup>8–10</sup> In this study, in line with previous research,<sup>8–10</sup> supplementing with ZnO to provide 2200 mg Zn/kg in the weanling pig diet improved growth performance and ameliorated weaning-associated diarrhea.

The gastrointestinal tract is fundamental to the uptake of nutrients, but it also acts as a physical barrier preventing harmful microorganisms and antigens from entry into deeper tissues. The intestinal epithelium barrier is formed from a single layer of epithelial cells and intercellular junctions. The tight junctions, the most apical of the intercellular junctions, work as the rate-limiting step in the paracellular pathway and form a



**Figure 1.** Effects of ZnO on MAPK signal pathways in the jejunal mucosa of piglets. The three MAPKs are JNK, p38, and ERKs 1 and 2. The bands are representative blots from one of six pigs. The values are calculated as the ratios of their phosphorylation levels (p-JNK, p-p38, p-ERK) and the total levels of MAPK. Data are means  $\pm$  SD ( $n = 6$ ). \*Differences were considered significant at  $P < 0.05$ .



**Figure 2.** Effects of ZnO on the expression of TGF- $\beta$ 1 in the jejunal mucosa of weanling piglets. The bands are representative blots from one of six pigs. The relative expression of TGF- $\beta$ 1 was expressed as the ratio of TGF- $\beta$ 1 and  $\beta$ -actin. Data are means  $\pm$  SD ( $n = 6$ ). \*Differences were considered significant at  $P < 0.05$ .

**Table 5.** Effects of ZnO on the mRNA expression of TGF- $\beta$ 1, T $\beta$ R and their downstream signals in weanling pigs.<sup>a</sup>

	Control	ZnO
TGF- $\beta$		

ulcerative colitis. Early weaning has been reported to impair the intestinal barrier function of piglets.<sup>4,20</sup> As reviewed by Wijtten et al.,<sup>21</sup> the intestinal barrier dysfunction induced by weaning is characterized by shortened villus height, increased mucosal permeability, disturbed active and passive absorption, and stimulated pro-inflammatory cytokine production. In the present study, the addition of ZnO to the diet increased villus height and the TER value, and decreased FD4 flux across the jejunal mucosa. These findings demonstrate that feeding high levels of ZnO to weanling piglets ameliorates weaning-associated intestinal injury and improves intestinal barrier function, which is consistent with recent research.<sup>9,10</sup> There is abundance of research on the modulation of zinc on intestinal mucosal repair. A high level of ZnO prevented the increase of paracellular permeability induced by enterotoxigenic *Escherichia*

selectively permeable barrier.<sup>4,9</sup> The disruption of tight junctions leads to increased intestinal permeability, which contributes to the initiation and development of many diseases, such as Crohn's disease and

coli in human Caco-2 enterocytes.<sup>6</sup> Rodriguez et al.<sup>22</sup> showed that malnutrition in guinea pigs was associated with a decreased number of tight junctional strands, and pharmacological doses of zinc prevented such an abnormality of tight junction morphology. High-dose zinc alleviated tight junction injury in the intestine of rats with acute colitis caused by dinitro-benzene-sulfonic acid.<sup>7</sup> However, the mechanisms of ZnO on mucosal integrity are still not well elucidated. Some researchers think that ZnO prevents diarrhea owing to its antibacterial activity. *In vitro* studies show that ZnO inhibits the growth of *Staphylococcus aureus* and *E. coli*.<sup>23</sup> In piglets, high doses of ZnO reduce bacterial translocation from the small intestine to the ileal mesenteric lymph node and decrease the viable *Clostridium* count in the intestine.<sup>24,25</sup> On the contrary, Jensen-Waern et al.<sup>26</sup> found that supplementation with 2500 mg/kg in the form of ZnO did not decrease *E. coli* or *Enterococci* in feces of weaned piglets.<sup>26</sup> Roselli et al.<sup>6</sup> reported that ZnO protected cells from enterotoxigenic *E. coli*-induced damage by inhibiting bacterial adhesion and internalization, preventing the disruption of barrier integrity and modulating cytokine expression, without a direct antibacterial effect.

To better elucidate the molecular mechanisms by which ZnO supplementation supports epithelial barrier integrity, we determined, for the first time, whether supplementation with ZnO influenced MAPK signaling pathways in weaned pigs. MAPKs are a family of serine/threonine protein kinases widely conserved among eukaryotes and convert extracellular stimuli into a wide range of cellular responses. The MAPK pathways consist of three major groupings and numerous related proteins that constitute interrelated signal transduction cascades.<sup>11</sup> The three major groupings are the ERK, JNK and p38 cascades. Using a porcine model of intestinal ischemia, Shifflett et al.<sup>11</sup> demonstrated that inhibition of JNK resulted in enhanced recovery of barrier function, whereas inhibition of ERK1/2 and p38 MAPK had a deleterious effect on mucosal recovery from ischemia. A recent study showed that weaning stress activated p38, JNK and ERK 1/2 signaling pathways in the intestine, which may be an important mechanism of weaning-associated enteric disorders of piglets.<sup>4</sup> So far, only a few studies in *in vitro* cell models have investigated the effects of zinc on MAPK signaling pathways. Liang et al.<sup>14</sup> found that zinc inhibited hydrogen peroxide-induced apoptosis in MC3T3-E1 cells via activation of ERK pathways and inhibition of p38 and JNK pathways. In the present study, we demonstrated, for the first time, that dietary ZnO supplementation decreases the relative protein levels for phosphorylated p38 and JNK, while increasing phosphorylated ERK 1/2, indicating that ZnO inhibits the JNK and p38 signaling pathways while activating ERK 1/2 signaling in weaned pigs. The JNK and p38 cascades are activated by inflammatory

cytokines and by a wide variety of cellular stresses.<sup>27</sup> Early weaning has been reported to up-regulate the expression of pro-inflammatory cytokines in the intestine.<sup>4</sup> A recent study showed that ZnO supplementation down-regulated the expression of pro-inflammatory cytokines in weaned pigs.<sup>9,28</sup> The down-regulation of inflammatory cytokines is in line with the beneficial effects of oral IFN- $\alpha$  in piglets at weaning.<sup>29,30</sup> Oral IFN- $\alpha$  modulated the inflammatory response to early weaning and increased the daily mean mass gain of pigs.<sup>30</sup> The inhibition of ZnO on p38 and JNK signaling pathways might be caused by the down-regulation of pro-inflammatory cytokine levels. The ERK 1/2 cascade is activated by growth factors, and preferentially regulates cell growth and differentiation.<sup>27</sup> Liang et al.<sup>31</sup> reported that zinc increased osteogenic function in mouse MC3T3-E1 osteoblasts through the stimulation of ERK signaling.<sup>31</sup> Wang et al.<sup>32</sup> reported that supplementation with 3000 mg Zn/kg from ZnO increased the A-Raf-1 expression in the jejunum of weaning piglets. Because Raf is a member of the Ras/Raf/MEK/ERK signaling pathway, the activation of ERK in ZnO-supplemented piglets may be expected. The present study demonstrated, for the first time, that dietary ZnO supplementation activated ERK 1/2 signaling in weaned pigs. In a follow-up study, the specific inhibitors of MAPKs may be used to investigate whether inhibition of MAPKs can affect the beneficial role of ZnO in intestinal barrier of weaned piglets.

TGF- $\beta$ 1 plays an important regulatory role in mucosal immune reactions and intestinal barrier restoration. During injury or disease, TGF- $\beta$  stimulates epithelial cell migration, increases extracellular matrix and integrin production, promotes intestinal epithelial restitution, and improves intestinal mucosa integrity.<sup>33,34</sup> IFN- $\gamma$  has been reported to be the main permeability increasing cytokine in the gut.<sup>35</sup> TGF- $\beta$ 1 preserves intestinal epithelial barrier function after exposure to IFN- $\gamma$ . TGF- $\beta$  plays an important role in the post-weaning adaptation process in the intestine of the pig. Mei and Xu<sup>12</sup> showed that the expression intensity of TGF- $\beta$ 1 at the intestinal villus epithelium decreased significantly 4 d after weaning. The transient decline in the TGF- $\beta$ 1 level of the intestinal villus following weaning contributed to post-weaning intestinal villus atrophy.<sup>12</sup> The research on ZnO regulation of TGF- $\beta$ 1 expression has so far only been reported *in vitro*. Zinc markedly increased the concentration of TGF- $\beta$ 1 in culture medium secreted by osteoblastic MC3T3-E1 cells.<sup>36</sup> In femoral tissues of newborn rats, and bone tissues with fracture healing, zinc has a stimulatory effect on the production of TGF- $\beta$ 1.<sup>37,38</sup> In human Caco-2 enterocytes, Roselli et al.<sup>6</sup> showed that the enterotoxigenic *E. coli* K<sub>88</sub> infection reduced TGF- $\beta$ 1 expression compared with uninfected cells, and ZnO up-regulated TGF- $\beta$  mRNA in *E. coli* K<sub>88</sub>-infected cells. However, owing to post-transcriptional

regulatory actions, the increased expression of TGF- $\beta$  mRNA did not indicate the increased expression of TGF- $\beta$  protein.<sup>35</sup> Therefore, in the current study, we analyzed the expression of both the mRNA and the protein. We showed, for the first time, that ZnO supplementation increased the intestinal expression of TGF- $\beta$ 1 in weaned pigs. The beneficial role of ZnO in intestinal barrier function might be related to the increase in intestinal TGF- $\beta$ 1 expression.

The canonical TGF- $\beta$  signaling pathway is mediated by SMAD family proteins. SMAD proteins—the only substrates of T $\beta$ R kinase—are critical mediators of the TGF- $\beta$  signaling transducer. SMAD 2 and SMAD3 are receptor-regulated SMADS (R-SMADS). Following stimulation by TGF- $\beta$ , SMAD 2 and SMAD 3 become phosphorylated. Phosphorylated SMAD 2/3 can complex with SMAD4 (the common-mediator SMAD), and then translocate to the nucleus and regulate gene expression.<sup>13</sup> To determine whether ZnO supplementation affected SMAD signaling pathways in weaned pigs, mucosa scraped from jejunum was analyzed for the expression of T $\beta$ R (T $\beta$ R, T $\beta$ R II) and the downstream signals (SMAD 2, SMAD 3, SMAD 7). The results showed that ZnO supplementation had no effect on the relative mRNA levels for T $\beta$ R levels (T $\beta$ R I, T $\beta$ R II) or the downstream signals (SMAD 2, SMAD 3, SMAD 7). We also analyzed phosphorylated SMAD 2/3, but we did not find a suitable Ab for pig. Using Ab for humans, we failed to detect the SMADs proteins by Western blotting. Therefore, we analyzed the changes in the mRNA level. However, several studies have also shown that the mRNA expression of SMADs can also be affected in some situations. In further studies, using other methods, we will try to detect the phosphorylation or other post-translational modifications of SMADs.

It is well-established that the beneficial role of TGF- $\beta$ 1 in the epithelial barrier is exerted mainly as a result of SMAD-independent signaling pathways.<sup>39</sup> Howe et al.<sup>33</sup> reported that TGF- $\beta$ 1-induced epithelial barrier enhancement was mediated by ERK 1/2 and SMAD 2/3 signaling pathways in human T84 colonic epithelial cells. TGF- $\beta$ 1 is known to activate the ERK 1/2 pathway in multiple cell types.<sup>33</sup> There is some evidence that the activation of the ERK 1/2 signaling is linked to the TGF- $\beta$ 1 induced modulation of tight junction permeability.<sup>40</sup> In cultured rat intestinal epithelial cells (IEC-6), TGF- $\beta$ 1 increases wound closures via ERK activation.<sup>41</sup> TGF- $\beta$ 1 stimulates re-epithelialization in human corneal epithelial cells through the Ras/MEK/ERK signaling pathway.<sup>42</sup> In mesenchymal cells, TGF- $\beta$ 1 induced ERK activation through the TGF- $\beta$ /phosphatidylinositol 3-kinase (PI3K)/p21-activated kinase2 (Pak2)/Raf/MEK/ ERK pathway.<sup>43</sup> TGF- $\beta$ 1 also activates ERK signaling through direct phosphorylation of ShcA.<sup>44</sup> ERK was reported to regulate the integrity of tight junction and the barrier

function of the epithelium.<sup>45</sup> In Caco-2 cells, phospho-ERK directly interacts with occludin and prevents hydrogen peroxide-induced disruption of tight junctions.<sup>45</sup> The activation of ERK 1/2 leads to increased expression of claudin-4 in MDCK cells.<sup>46</sup> In the present study, supplementation with ZnO increased the intestinal expression of TGF- $\beta$ 1 and activated the ERK 1/2 signaling pathway. The results indicated that the activation of ERK 1/2 signaling might be

8. Hu CH, Gu LY, Luan ZS, et al. Effects of montmorillonite-zinc oxide hybrid on performance, diarrhea, intestinal permeability and morphology of weanling pigs. *Anim Feed Sci Tech* 2012; 177: 108–115.
9. Hu C, Song J, Li Y, et al. Diosmectite-zinc oxide composite improves intestinal barrier function, modulates expression of pro-inflammatory cytokines and tight junction protein in early weaned pigs. *Br J Nutr* 2013; 110: 681–688.
10. Zhang B and Guo Y. Supplemental zinc reduced intestinal permeability by enhancing occludin and zonula occludens protein-1 (ZO-1) expression in weaning piglets. *Br J Nutr* 2009; 102: 687–693.
11. Shifflett DE, Jones SL, Moeser AJ, et al. Mitogen-activated protein kinases regulate COX-2 and mucosal recovery in ischemic-injured porcine ileum. *Am J Physiol Gastrointest Liver Physiol* 2004; 286: G906–G913.
12. Mei J and Xu RJ. Transient changes of transforming growth factor-beta expression in the small intestine of the pig in association with weaning. *Br J Nutr* 2005; 93: 37–45.
13. Mehra A and Wrana JL. TGF-beta and the Smad signal transduction pathway. *Biochem Cell Biol* 2002; 80: 605–622.
14. Liang D, Yang M, Guo B, et al. Zinc inhibits H(2)O(2)-induced MC3T3-E1 cells apoptosis via MAPK and PI3K/AKT pathways. *Biol Trace Elem Res* 2012; 148: 420–429.
15. National Research Council. *Nutrient requirements of swine*, 10th ed. Washington, DC: National Academy Press, 1998.
16. Liu Y, Chen F, Odle J, et al. Fish oil enhances intestinal integrity and inhibits TLR4 and NOD2 signaling pathways in weaned pigs after LPS challenge. *J Nutr* 2012; 142: 2017–2024.
17. Chen Y, Chen D, Tian G, et al. Dietary arginine supplementation alleviates immune challenge induced by *Salmonella enterica* serovar *Choleraesuis* bacterin potentially through the Toll-like receptor 4-myeloid differentiation factor 88 signalling pathway in weaned piglets. *Br J Nutr* 2012; 108: 1069–1076.
18. Livak KJ and Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001; 25: 402–408.
19. SAS Institute. *SAS/STAT User's guide*, version 8.1. Cary, NC: SAS Institute, 2000.
20. He Q, Tang H, Ren P, et al. Dietary supplementation with l-arginine partially counteracts serum metabonome induced by weaning stress in piglets. *J Proteome Res* 2011; 10: 5214–5221.
21. Wijtten PJ, van der Meulen J and Verstegen MW. Intestinal barrier function and absorption in pigs after weaning: a review. *Br J Nutr* 2011; 105: 967–981.
22. Rodriguez P, Darmon N, Chappuis P, et al. Intestinal paracellular permeability during malnutrition in guinea pigs: effect of high dietary zinc. *Gut* 1996; 39: 416–422.
23. Sawai J. Quantitative evaluation of antibacterial activities of metallic oxide powders (ZnO, MgO and CaO) by conductimetric assay. *J Microbiol Meth* 2003; 54: 177–182.
24. Huang SX, McFall M, Cegielski AC, et al. Effect of dietary zinc supplementation on *Escherichia coli* septicemia in weaned pigs. *Swine Health Prod* 1999; 7: 109–111.
25. Hu CH, Xiao K, Song J, et al. Effects of zinc oxide supported on zeolite on growth performance, intestinal microflora and permeability, and cytokines expression of weaned pigs. *Anim Feed Sci Tech* 2013; 181: 65–71.
26. Jensen-Waern M, Melin L, Lindberg R, et al. Dietary zinc oxide in weaned pigs—effects on performance, tissue concentrations, morphology, neutrophil functions and faecal microflora. *Res Vet Sci* 1998; 64: 225–231.
27. Haddad JJ and Harb HL. Cytokines and the regulation of hypoxia-inducible factor (HIF)-1alpha. *Int Immunopharmacol* 2005; 5: 461–483.
28. Hu CH, Song ZH, Xiao K, et al. Zinc oxide influences intestinal integrity, the expressions of genes associated with inflammation and TLR4-myeloid differentiation factor 88 signaling pathways in weanling pigs. *Innate Immun.* Epub ahead of print 16 August 2013. DOI: 10.1177/1753425913499947.
29. Razzuoli E, Villa R, Sossi E, et al. Characterization of the interferon- $\alpha$  response of pigs to the weaning stress. *J Interf Cytok Res* 2011; 31: 237–247.
30. Amadori M, Farinacci M, Begni B, et al. Effects of interferon- $\alpha$  on the inflammatory response of swine peripheral blood mononuclear cells. *J Interf Cytok Res* 2009; 29: 241–248.
31. Liang D, Yang M, Guo B, et al. Zinc upregulates the expression of osteoprotegerin in mouse osteoblasts MC3T3-E1 through PKC/MAPK pathways. *Biol Trace Elem Res* 2012; 146: 340–348.
32. Wang X, Ou D, Yin J, et al. Proteomic analysis reveals altered expression of proteins related to glutathione metabolism and apoptosis in the small intestine of zinc oxide-supplemented piglets. *Amino Acids* 2009; 37: 209–218.
33. Howe KL, Reardon C, Wang A, et al. Transforming growth factor-beta regulation of epithelial tight junction proteins enhances barrier function and blocks enterohemorrhagic *Escherichia coli* O157:H7-induced increased permeability. *Am J Pathol* 2005; 167: 1587–1597.
34. Hering NA, Andres S, Fromm A, et al. Transforming growth factor-beta, a whey protein component, strengthens the intestinal barrier by upregulating claudin-4 in HT-29/B6 cells. *J Nutr* 2011; 141: 783–789.
35. Beaupaire C, Smyth D and McKay DM. Interferon- $\gamma$  regulation of intestinal epithelial permeability. *J Interf Cytok Res* 2009; 29: 133–144.
36. Yamaguchi M and Hashizume M. Effect of beta-alanyl-L-histidinato zinc on protein components in osteoblastic MC3T3-E1 cells: increase in osteocalcin, insulin-like growth factor-I and transforming growth factor-beta. *Mol Cell Biochem* 1994; 136: 163–169.
37. Ma ZJ, Misawa H and Yamaguchi M. Stimulatory effect of zinc on insulin-like growth factor-I and transforming growth factor-beta1 production with bone growth of newborn rats. *Int J Mol Med* 2001; 8: 623–628.
38. Igarashi A and Yamaguchi M. Increase in bone growth factors with healing rat fractures: the enhancing effect of zinc. *Int J Mol Med* 2001; 8: 433–438.
39. Derynck R and Zhang YE. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. *Nature* 2003; 425: 577–584.
40. Martin-Martin N, Slattery C, Mc Morrow T, et al. TGF-beta1 mediates sirolimus and cyclosporine A-induced alteration of barrier function in renal epithelial cells via a noncanonical ERK1/2 signaling pathway. *Am J Physiol Renal Physiol* 2011; 301: F1281–F1292.
41. Suer S, Ampasala D, Walsh MF, et al. Role of ERK/mTOR signaling in TGFbeta-modulated focal adhesion kinase mRNA stability and protein synthesis in cultured rat IEC-6 intestinal epithelial cells. *Cell Tissue Res* 2009; 336: 213–223.
42. Secker GA, Shortt AJ, Sampson E, et al. TGFbeta stimulated re-epithelialisation is regulated by CTGF and Ras/MEK/ERK signalling. *Exp Cell Res* 2008; 314: 131–142.
43. Hough C, Radu M and Dore JJ. TGFbeta induced Erk phosphorylation of smad linker region regulates smad signaling. *PLoS One* 2012; 7: e42513.
44. Lee MK, Pardoux C, Hall MC, et al. TGF-beta activates Erk MAP kinase signalling through direct phosphorylation of ShcA. *EMBO J* 2007; 26: 3957–3967.
45. Basuroy S, Seth A, Elias B, et al. MAPK interacts with occludin and mediates EGF-induced prevention of tight junction disruption by hydrogen peroxide. *Biochem J* 2006; 393: 69–77.
46. Ikari A, Atomi K, Takiguchi A, et al. Epidermal growth factor increases claudin-4 expression mediated by Sp1 elevation in MDCK cells. *Biochem Biophys Res Commun* 2009; 384: 306–310.