

# AMP-activated protein kinase is required for the anti-adipogenic effects of alpha-linolenic acid

Xihong Zhou, Weiche Wu, Jingqing Chen, Xinxia Wang and Yizhen Wang\*

## Abstract

**Background:** n-3 long chain polyunsaturated fatty acid (n-3 LC PUFA) increases  $\beta$ -oxidation and limits lipid accumulation in adipocytes. The current study was conducted to determine whether their precursor alpha-linolenic acid (ALA) could also exert the above effects and how AMP-activated protein kinase (AMPK) was involved.

**Methods:** AMPK $\alpha$ 1<sup>-/-</sup>, AMPK $\alpha$ 2<sup>-/-</sup> mice and wild-type (WT) mice were fed a high-fat diet (HFD) or HFD with ALA. Body weight was recorded weekly and serum was collected. Adipocytes size and expression of key players involved in mitochondrial biogenesis and lipid oxidation were also measured.

**Results:** Our results showed an elevated serum adiponectin level and a decreased leptin and insulin level in WT mice fed HFD with ALA when compared with WT mice fed HFD. In addition, dietary ALA decreased epididymal adiposity and adipocytes size in WT mice. At protein level, mitochondrial genes (peroxisome proliferator-activated receptor gamma coactivator 1 alpha [PGC1 $\alpha$ ] and nuclear respiratory factor-1 [nrf1]) and  $\beta$ -oxidation related genes (carnitine palmitoyltransferase 1A [CPT1a] and peroxisome proliferator-activated receptor alpha [PPAR $\alpha$ ]) were upregulated by dietary ALA in epididymal fat of WT mice. Consistently, dietary ALA also increased mitochondrial genomic DNA copy numbers. Moreover, lipogenesis was repressed by dietary ALA, indicated by that expression of fatty acid synthase (FAS), acetyl CoA carboxylase (ACC) and stearoyl-CoA desaturase 1 (SCD1) were decreased. However, these aforementioned effects were abolished in the AMPK $\alpha$ 1 and AMPK $\alpha$ 2 knockout mice.

**Conclusions:** Our results suggest that ALA could improve adipose tissue function and its anti-adipogenic effects are dependent on AMPK.

**Keywords:** Alpha-linolenic acid, AMP-activated protein kinase,  $\beta$ -oxidation, Mitochondrial biogenesis, Adipose tissue

## Introduction

Obesity is a global public health problem, and it is associated with a high risk of cardiovascular disease, type 2 diabetes, and certain cancers [1]. Adipose tissue is a specialized organ that stores energy in the form of triglycerides and secretes various hormones and cytokines that regulate metabolism and inflammation [2]. Adipogenesis is the process by which preadipocytes differentiate into mature adipocytes, and it is regulated by a complex network of transcription factors and signaling pathways [3].

One of the key transcription factors involved in adipogenesis is peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ), which is expressed in preadipocytes and upregulated during adipogenesis [4]. PPAR $\gamma$  activation promotes the expression of adipogenic genes, such as fatty acid synthase (FAS), acetyl CoA carboxylase (ACC), and stearoyl-CoA desaturase 1 (SCD1), and inhibits the expression of mitochondrial genes, such as peroxisome proliferator-activated receptor gamma coactivator 1 alpha (PGC1 $\alpha$ ) and nuclear respiratory factor-1 (NRF1) [5].

Recent studies have shown that n-3 long chain polyunsaturated fatty acids (n-3 LC PUFA) can improve adipose tissue function and reduce adiposity in animal models and humans [6]. n-3 LC PUFA increases  $\beta$ -oxidation and limits lipid accumulation in adipocytes [7]. The current study was conducted to determine whether their precursor alpha-linolenic acid (ALA) could also exert the above effects and how AMP-activated protein kinase (AMPK) was involved [8].

AMPK is a heterotrimeric complex composed of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits. It is activated by various stimuli, including low energy status, and it plays a central role in energy homeostasis and metabolic regulation [9]. AMPK activation promotes mitochondrial biogenesis and  $\beta$ -oxidation, and inhibits lipogenesis and lipolysis [10].

In this study, we investigated the effects of dietary ALA on adipose tissue function and metabolism in wild-type (WT) mice and AMPK $\alpha$ 1 and AMPK $\alpha$ 2 knockout mice. We found that dietary ALA improved adipose tissue function and reduced adiposity in WT mice, and these effects were abolished in the AMPK $\alpha$ 1 and AMPK $\alpha$ 2 knockout mice. Our results suggest that ALA could improve adipose tissue function and its anti-adipogenic effects are dependent on AMPK.

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Materials and methods

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Results

Effects of ALA on serum insulin, adiponectin and leptin concentration in WT, AMPK $\alpha$ 1<sup>-/-</sup> and AMPK $\alpha$ 2<sup>-/-</sup> mice

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AMPK $\alpha$ 2<sup>-/-</sup> AMPK $\alpha$ 1<sup>-/-</sup>

Effects of ALA on HFD-induced fat deposition in white adipose tissue required AMPK

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AMPK $\alpha$ 1<sup>-/-</sup> AMPK $\alpha$ 2<sup>-/-</sup> .

AMPK $\alpha$ 2<sup>-/-</sup> AMPK $\alpha$ 1<sup>-/-</sup> ,

Table 2 Effects of ALA on fat depots of WT, AMPK $\alpha 1^{-/-}$  and AMPK $\alpha 2^{-/-}$  mice fed HFD

	WT		AMPK $\alpha 1^{-/-}$		AMPK $\alpha 2^{-/-}$	
	HF	HF-A	HF	HF-A	HF	HF-A
Final BW (g)	42.6 ± 3.1	32.6 ± 1.8 <sup>ab</sup>	43.1 ± 3.0	42 ± 3.2	45.5 ± 3.4	44.2 ± 3.7
Epididymal						
weight (mg)	2270 ± 158	1224 ± 98 <sup>ab</sup>	2242 ± 201	2137 ± 198	2349 ± 178	2146 ± 184
DNA (μg/mg)	0.32 ± 0.04	0.67 ± 0.05 <sup>ab</sup>	0.29 ± 0.05	0.35 ± 0.06	0.29 ± 0.05	0.36 ± 0.07
Inguinal						
Weight (mg)	1090 ± 42	1037 ± 57	1097 ± 45	1058 ± 55	1102 ± 49	1072 ± 59
DNA (μg/mg)	0.42 ± 0.05	0.5 ± 0.06	0.36 ± 0.04	0.39 ± 0.06	0.38 ± 0.05	0.41 ± 0.07

Note: The values are means ± SE. <sup>a</sup>P < 0.05 for difference between different genotypes with HF-A diet; <sup>b</sup>P < 0.05 for difference between WT mice. BW, body weight. HF, mice fed high-fat diet; HF-A, mice fed high-fat diet with ALA.

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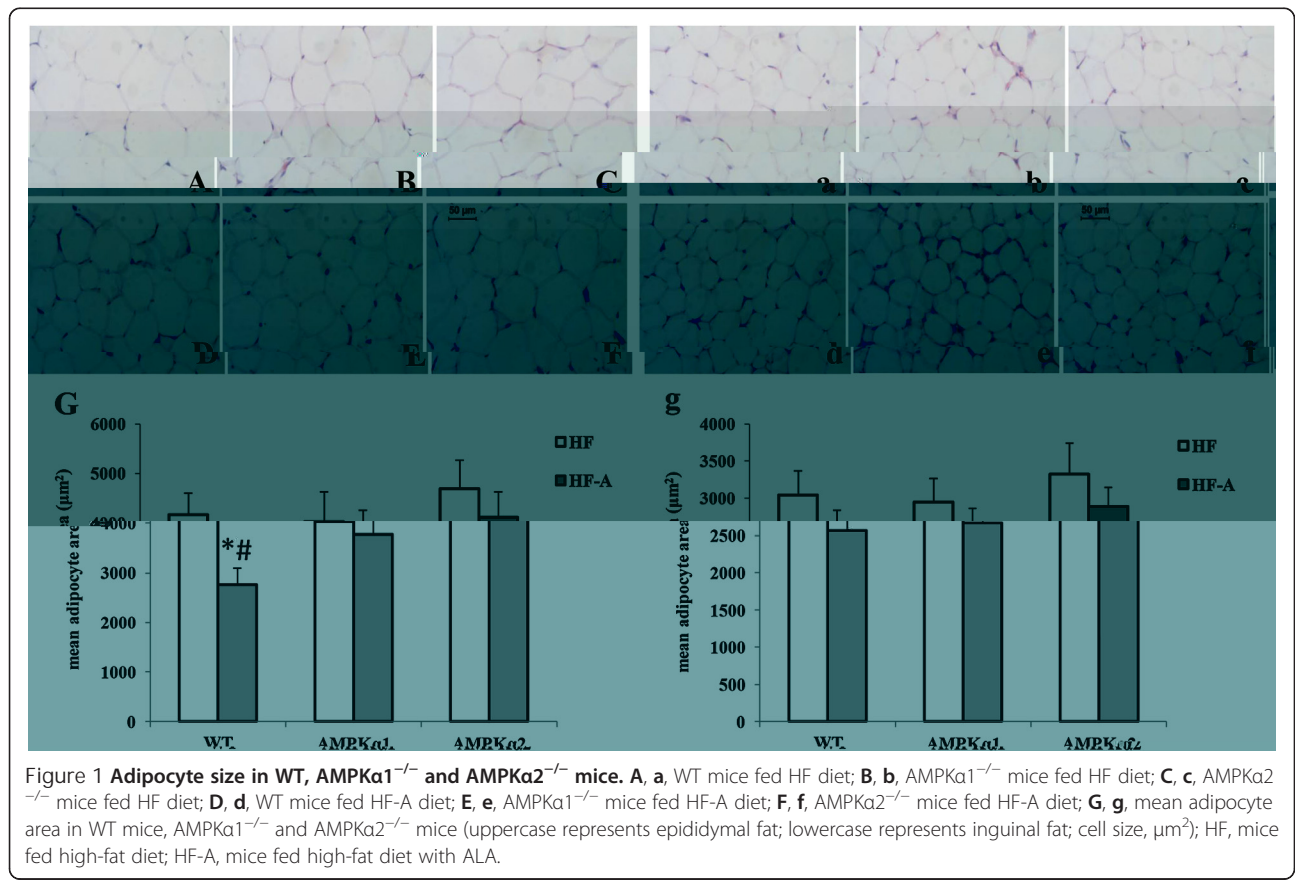


Figure 1 Adipocyte size in WT, AMPK $\alpha 1^{-/-}$  and AMPK $\alpha 2^{-/-}$  mice. **A, a**, WT mice fed HF diet; **B, b**, AMPK $\alpha 1^{-/-}$  mice fed HF diet; **C, c**, AMPK $\alpha 2^{-/-}$  mice fed HF diet; **D, d**, WT mice fed HF-A diet; **E, e**, AMPK $\alpha 1^{-/-}$  mice fed HF-A diet; **F, f**, AMPK $\alpha 2^{-/-}$  mice fed HF-A diet; **G, g**, mean adipocyte area in WT mice, AMPK $\alpha 1^{-/-}$  and AMPK $\alpha 2^{-/-}$  mice (uppercase represents epididymal fat; lowercase represents inguinal fat; cell size, μm<sup>2</sup>); HF, mice fed high-fat diet; HF-A, mice fed high-fat diet with ALA.

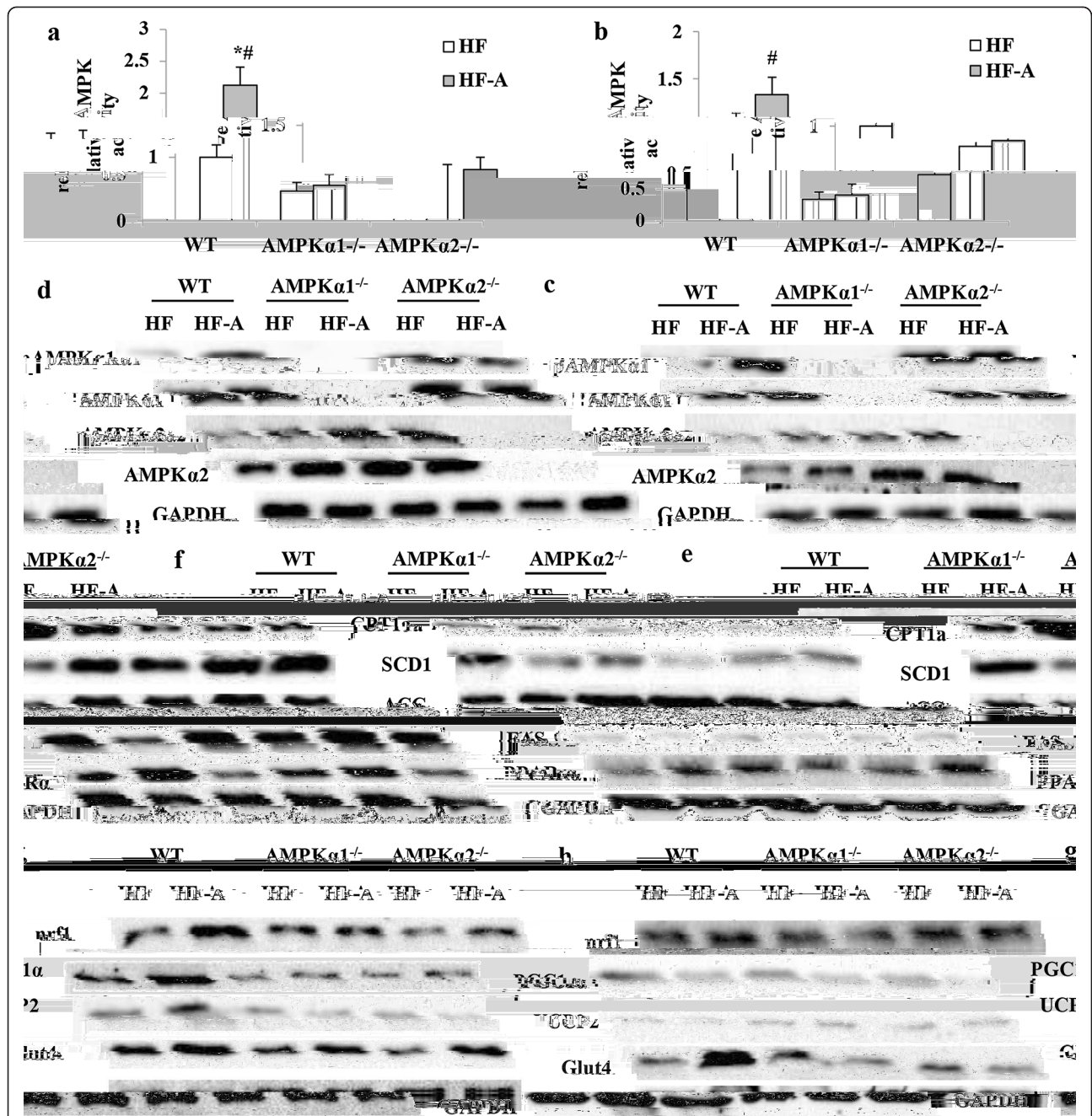


Figure 2 AMPK activity, protein expression of mitochondrial biogenesis and  $\beta$ -oxidation related genes in WT, AMPK $\alpha$ 1 $^{-/-}$  and AMPK $\alpha$ 2 $^{-/-}$  mice. HF, mice fed high-fat diet; HF-A, mice fed high-fat diet with ALA. (a, c, e, g, activity of AMPK and protein expressed in epididymal fat; b, d, f, h, activity of AMPK and protein expressed in inguinal fat). The values are means  $\pm$  SE. #P < 0.05 for difference between different genotypes with HF-A diet; \*P < 0.05 for difference between WT mice.

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carried out the experiments and collected the data; WYZ participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

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#### References

1. Azain MJ. Role of fatty acids in adipocyte growth and development. *J Anim Sci.* 2004;82:916–24.
2. Lapillonne A, Clarke SD, Heird WC. Polyunsaturated fatty acids and gene expression. *Curr Opin Clin Nutr Metab Care.* 2004;7:151–6.
3. Baillie RA, Takada R, Nakamura M, Clarke SD. Coordinate induction of peroxisomal acyl-CoA oxidase and UCP-3 by dietary fish oil: a mechanism for decreased body fat deposition. *Prostaglandins Leukot Essent Fatty Acids.* 1999;60:351–6.
4. Raclot T, Groscolas R, Langin D, Ferre P. Site-specific regulation of gene expression by n-3 polyunsaturated fatty acids in rat white adipose tissues. *J Lipid Res.* 1997;38:1963–72.
5. Raclot T, Oudart H. Selectivity of fatty acids on lipid metabolism and gene expression. *Proc Nutr Soc.* 1999;58:633–46.
6. Flachs P, Horakova O, Brauner P, Rossmeisl M, Pecina P, Franssen-van Hal N, et al. Polyunsaturated fatty acids of marine origin upregulate mitochondrial biogenesis and induce beta-oxidation in white fat. *Diabetologia.* 2005;48:2365–75.
7. Ruzickova J, Rossmeisl M, Prazak T, Flachs P, Sponarova J, Veck M, et al. Omega-3 PUFA of marine origin limit diet-induced obesity in mice by reducing cellularity of adipose tissue. *Lipids.* 2004;39:1177–85.
8. Lorente-Cebrian S, Costa AG, Navas-Carretero S, Zabala M, Martinez JA, Moreno-Aliaga MJ. Role of omega-3 fatty acids in obesity, metabolic syndrome, and cardiovascular diseases: a review of the evidence. *J Physiol Biochem.* 2013;69:633–51.
9. Gonzalez-Manan D, Tapia G, Gormaz JG, D'Espessailles A, Espinosa A, Masson L, et al. Bioconversion of alpha-linolenic acid to n-3 LCPUFA and expression of PPAR-alpha, acyl Coenzyme A oxidase 1 and carnitine acyl transferase I are incremented after feeding rats with alpha-linolenic acid-rich oils. *Food Funct.* 2012;3:765–72.
10. Oliva ME, Ferreira MR, Chicco A, Lombardo YB. Dietary Salvia (*Salvia hispanica* L) seed rich in alpha-linolenic acid improves adipose tissue dysfunction and the altered skeletal muscle glucose and lipid metabolism in dyslipidemic insulin-resistant rats. *Prostaglandins Leukot Essent Fatty Acids.* 2013;89:279–89.
11. Fukumitsu S, Villareal MO, Onaga S, Aida K, Han J. alpha-Linolenic acid suppresses cholesterol and triacylglycerol biosynthesis pathway by suppressing SREBP-2, SREBP-1a and -1c expression. *Cytotechnology.* 2013;65:899–907.
12. Monteiro J, Askarian F, Nakamura MT, Moghadasian MH, Ma DW. Oils rich in alpha-linolenic acid independently protect against characteristics of fatty liver disease in the Delta6-desaturase null mouse. *Can J Physiol Pharmacol.* 2013;91:469–79.
13. Krey G, Braissant O, L'Horsset F, Kalkhoven E, Perroud M, Parker MG, et al. Fatty acids, eicosanoids, and hypolipidemic agents identified as ligands of peroxisome proliferator-activated receptors by coactivator-dependent receptor ligand assay. *Mol Endocrinol.* 1997;11:779–91.
14. Louet JF, Chatelain F, Decaux JF, Park EA, Kohl C, Pineau T, et al. Long-chain fatty acids regulate liver carnitine palmitoyltransferase I gene (L-CPT I) expression through a peroxisome-proliferator-activated receptor alpha (PPARalpha)-independent pathway. *Biochem J.* 2001;354:189–97.
15. Le May C, Cauzac M, Diradourian C, Perdereau D, Girard J, Burnol AF, et al. Fatty acids induce L-CPT I gene expression through a PPARalpha-independent mechanism in rat hepatoma cells. *J Nutr.* 2005;135:2313–9.
16. Kopecky J, Rossmeisl M, Flachs P, Kuda O, Brauner P, Jilkova Z, et al. n-3 PUFA: bioavailability and modulation of adipose tissue function. *Proc Nutr Soc.* 2009;68:361–9.
17. Ceddia RB. The role of AMP-activated protein kinase in regulating white adipose tissue metabolism. *Mol Cell Endocrinol.* 2013;366:194–203.
18. Jager S, Handschin C, St-Pierre J, Spiegelman BM. AMP-activated protein kinase (AMPK) action in skeletal muscle via direct phosphorylation of PGC-1alpha. *Proc Natl Acad Sci U S A.* 2007;104:12017–22.
19. Puigserver P, Spiegelman BM. Peroxisome proliferator-activated receptor-gamma coactivator 1 alpha (PGC-1 alpha): transcriptional coactivator and metabolic regulator. *Endocr Rev.* 2003;24:78–90.
20. Ronnett GV, Klemm AM, Kim EK, Landree LE, Tu Y. Fatty acid metabolism, the central nervous system, and feeding. *Obesity (Silver Spring).* 2006;14 Suppl 5:2015–7.
21. Hun CS, Hasegawa K, Kawabata T, Kato M, Shimokawa T, Kagawa Y. Increased uncoupling protein2 mRNA in white adipose tissue, and decrease in leptin, visceral fat, blood glucose, and cholesterol in KK-Ay mice fed with eicosapentaenoic and docosahexaenoic acids in addition to linolenic acid. *Biochem Biophys Res Commun.* 1999;259:85–90.
22. Belzung F, Raclot T, Groscolas R. Fish oil n-3 fatty acids selectively limit the hypertrophy of abdominal fat depots in growing rats fed high-fat diets. *Am J Physiol.* 1993;264:R1111–8.
23. Ravussin ES, Smith SR. Increased fat intake, impaired fat oxidation, and failure of fat cell proliferation result in ectopic fat storage, insulin resistance, and type 2 diabetes mellitus. *Ann N Y Acad Sci.* 2002;967:363–78.
24. Hill JO, Peters JC, Lin D, Yakubu F, Greene H, Swift L. Lipid accumulation and body fat distribution is influenced by type of dietary fat fed to rats. *Int J Obesity Relat d Metabolic Disorders J Int Assoc Stud Obesity.* 1993;17:223–36.
25. Daval M, Diot-Dupuy F, Bazin R, Hainault I, Viollet B, Vaulont S, et al. Anti-lipolytic action of AMP-activated protein kinase in rodent adipocytes. *J Biol Chem.* 2005;280:25250–7.
26. Lihn AS, Jessen N, Pedersen SB, Lund S, Richelsen B. AICAR stimulates adiponectin and inhibits cytokines in adipose tissue. *Biochem Biophys Res Commun.* 2004;316:853–8.
27. Bauwens JD, Schmuck EG, Lindholm CR, Ertel RL, Mulligan JD, Hovis I, et al. Cold tolerance, cold-induced hyperphagia, and nonshivering thermogenesis are normal in alpha (1)-AMPK-/- mice. *Am J Physiol Regul Integr Comp Physiol.* 2011;301:R473–83.
28. Jelenik T, Rossmeisl M, Kuda O, Jilkova ZM, Medrikova D, Kus V, et al. AMP-activated protein kinase alpha2 subunit is required for the

39. Abel ED, Peroni O, Kim JK, Kim YB, Boss O, Hadro E, et al. Adipose-selective targeting of the GLUT4 gene impairs insulin action in muscle and liver. *Nature*. 2001;409:729–33.
40. Kurth-Kraczek EJ, Hirshman MF, Goodyear LJ, Winder WW. 5' AMP-activated protein kinase activation causes GLUT4 translocation in skeletal muscle. *Diabetes*. 1999;48:1667–71.
41. Daval M, Foufelle F, Ferre P. Functions of AMP-activated protein kinase in adipose tissue. *J Physiol*. 2006;574:55–62.

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