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Binding of the pathogen receptor HSP90AA1 to avibirnavirus VP2 induces autophagy by inactivating the AKT-MTOR pathway

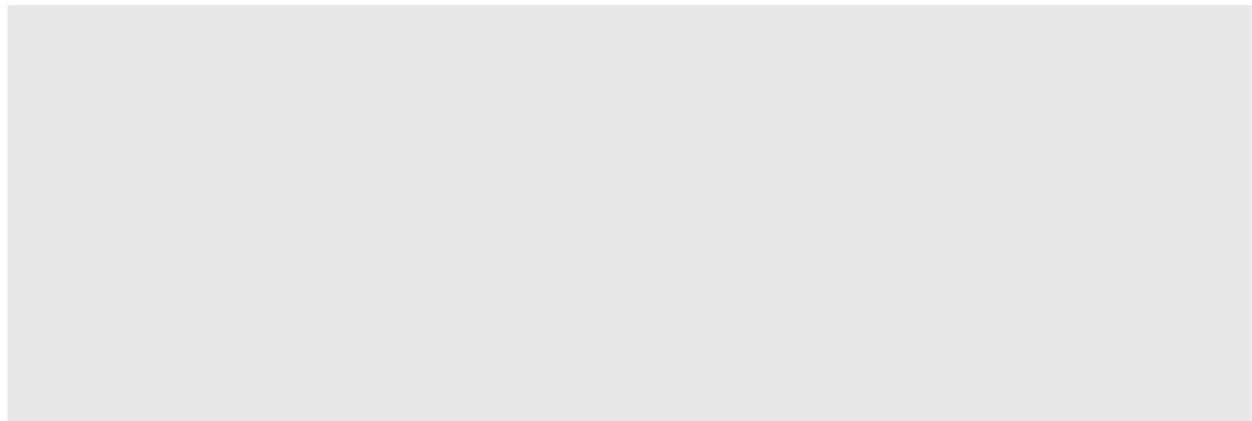
Boli Hu,^{1,2,†} Yina Zhang,^{1,†} Lu Jia,¹ Huansheng Wu,¹ Chengfei Fan,¹ Yanting Sun,¹ Chengjin Ye,¹ Min Liao,¹ and Jiyong Zhou^{1,2,*}

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[†]These authors contributed equally to this work.

: AKT-MTOR pathway, autophagy, HSP90AA1, avibirnavirus VP2

bb : ANOVA, analysis of variance; ATG5, autophagy-related 5; BCA, bicinchoninic acid; BECN1, Beclin-1, autophagy-related; cDNA, complementary DNA; CHIP, co-immunoprecipitation; DMEM, Dulbecco's modified Eagle's medium; dsRNA, double-stranded RNA; EBSS, Earle's balanced salt solution; EIF2AK2, eukaryotic translation initiation factor 2-kinase 2; EIF2S1, eukaryotic translation initiation factor 2, subunit 1; eGFP, enhanced green fluorescent protein; ER, endoplasmic reticulum; G, *Gallus gallus* (chicken); GAPDH, glyceraldehyde-3-phosphate dehydrogenase; GOPC, Golgi-associated PDZ domain-containing protein; GST, glutathione S-transferase; HE-IBDV, heterologous infection of IBDV; H, H₂O₂ (hydrogen peroxide); HSP90AA1, heat shock protein 90 α class A (class A) chaperone; HSV-1, herpesvirus 1; IBDV, infectious bursal disease virus; IG, immunoglobulin G; LPS, lipopolysaccharide; Ab, antibody; MAP1LC3/LC3, microtubule-associated protein 1 light chain 3; MOI, multiplicity of infection; MTOR, mechanistic target of rapamycin (serine/threonine kinase); N-NTA, N-terminal; PAMP, pathogen-associated molecular pattern; PBS, phosphate-buffered saline; PI3K, phosphatidylinositol (3-)-OH kinase; PRR, pattern recognition receptor; RNA, ribonucleic acid; SDS, sodium dodecyl sulfate; RNA, ribonucleic acid



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Results

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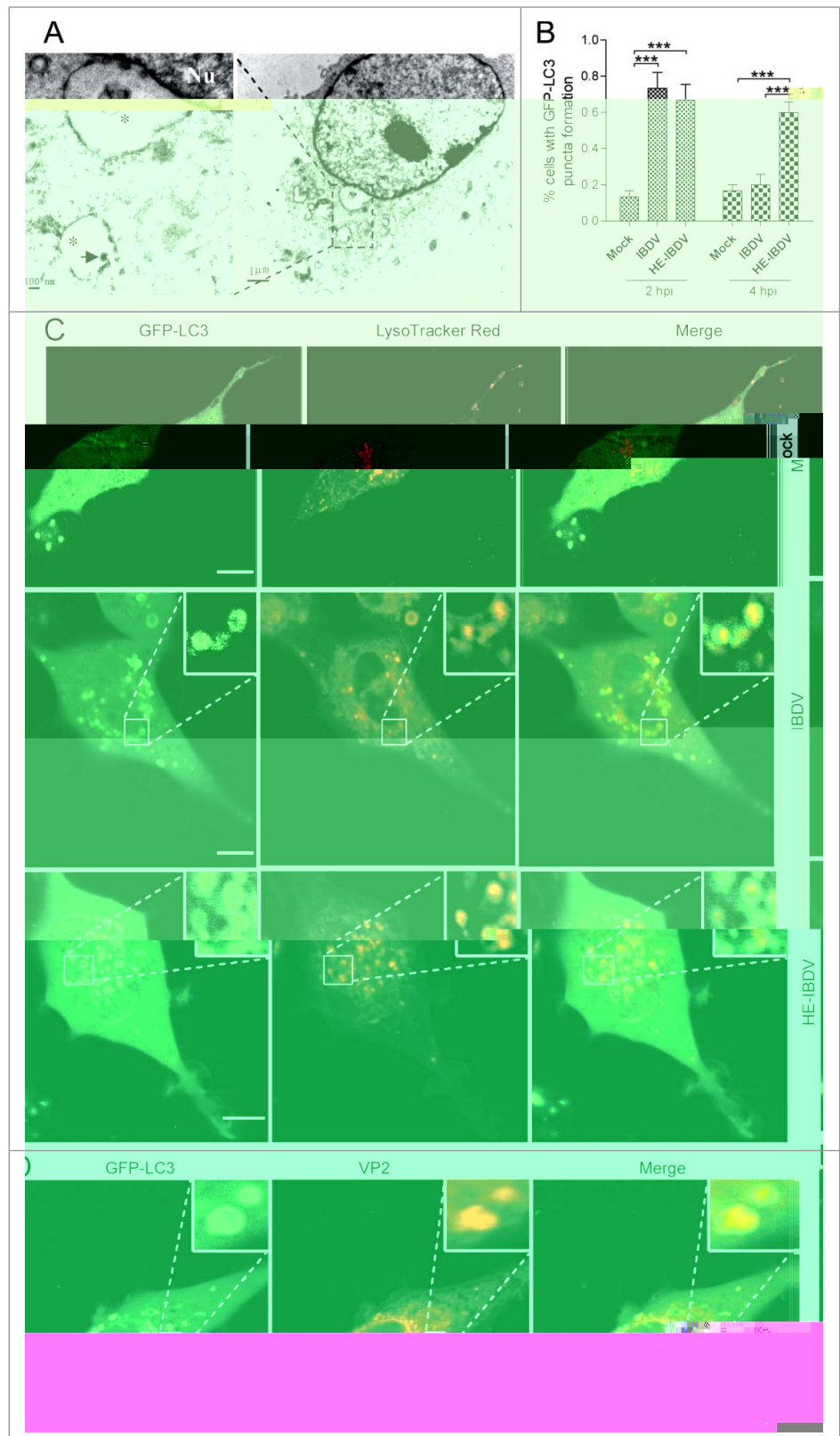
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Figure 1. IBDV infection induces autophagosome accumulation in DF-1 cells. **(A)** Autophagic vacuoles in infected cells observed by transmission electron microscopy. Autophagic vacuole engulfs IBDV particle (black arrows) and organelle (asterisk) in cytoplasm of DF-1 cell infected with IBDV (MOI = 10) at 2 hpi. **(B)** DF-1 cells transfected with peGFP-LC3 for 48 h, and then infected with IBDV (MOI = 10) or treated with HE-IBDV. At 2 hpi or 4 hpi, cells were incubated with LysoTracker Red (50 nM) for 30 min; intracellular autophagic vacuoles were observed under confocal microscopy. Statistical analysis of the number of cells with >3 autophagic vacuoles. At 2 hpi or 4 hpi, autophagic vacuoles were counted in IBDV-infected cells. Error bars: Mean \pm SD of 3 independent tests. Two-way analysis of variance (ANOVA); *** P < 0.001 compared to control. **(C)** DF-1 cells transfected with peGFP-LC3 for 48 h, and then infected with IBDV (MOI = 10) or treated with HE-IBDV. At 2 hpi or 4 hpi, cells were incubated with LysoTracker Red (50 nM) for 30 min; intracellular autophagic vacuoles were observed under confocal microscopy. Scale bars: 10 μ m. **(D)** DF-1 cells transfected with peGFP-LC3 and infected with IBDV at 9 h post-transfection. The cells were fixed, immunostained with anti-VP2 mAb, and autophagosomes were observed under confocal microscopy. Scale bar: 10 μ m.



IBDV, a double-stranded RNA virus, is a member of the Herpesviridae family. IBDV infection in DF-1 cells leads to the formation of autophagosomes, which are characterized by the presence of IBDV particles (black arrows) and organelles (asterisk) in the cytoplasm. At 2 hpi or 4 hpi, cells were incubated with LysoTracker Red (50 nM) for 30 min; intracellular autophagic vacuoles were observed under confocal microscopy. Scale bars: 10 μ m. **(D)** DF-1 cells transfected with peGFP-LC3 and infected with IBDV at 9 h post-transfection. The cells were fixed, immunostained with anti-VP2 mAb, and autophagosomes were observed under confocal microscopy. Scale bar: 10 μ m.

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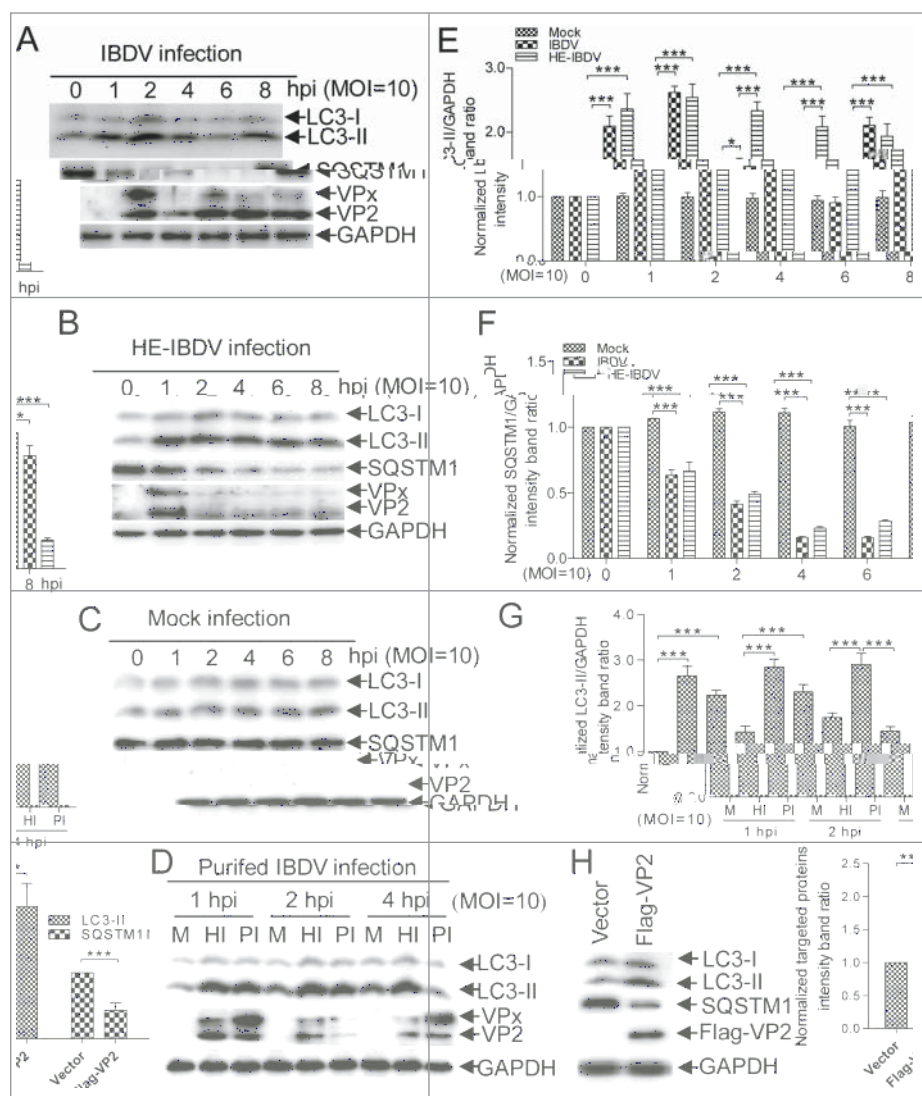


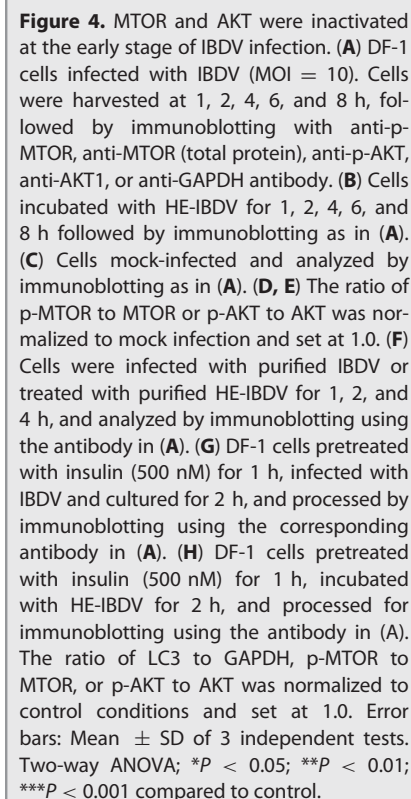
Figure 2. Characterization of IBDV-triggered autophagosome accumulation. (A) IBDV promotes increase of LC3-II and decrease of SQSTM1 within 1 hpi but inhibits it from 4 hpi. DF-1 cells were infected with IBDV (MOI = 10). (B) Increase of LC3-II and decrease of SQSTM1 was constantly promoted in HE-IBDV-treated cells. (C) LC3 amount and SQSTM1 accumulation in mock-infected cells were unaffected. Cells were harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. (D) DF-1 cells were infected with the purified infectious IBDV virions or treated with the purified heat-inactivated IBDV virions (MOI = 10), respectively. The purified infectious IBDV increases the amount of LC3-II within 1 hpi but inhibits it from 4 hpi. However, levels of LC3-II increase constantly in the purified HE-IBDV-treated cells. M, mock infected; HI, purified heat-inactivated IBDV virions; PI, purified infectious IBDV virions. (E, F) The ratio of LC3-II or SQSTM1 to GAPDH was normalized to control conditions in (A to C). (G) The ratio of LC3-II to GAPDH was normalized to control conditions in (D). (H) LC3-II increases and SQSTM1 decreases in VP2-transfected cells. 293T cells were transfected with the vector pFlag-VP2 for 48 h, harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. Error bars: Mean \pm SD of 3 independent tests. Two-way ANOVA, * P < 0.05; ** P < 0.01; *** P < 0.001 compared to control.

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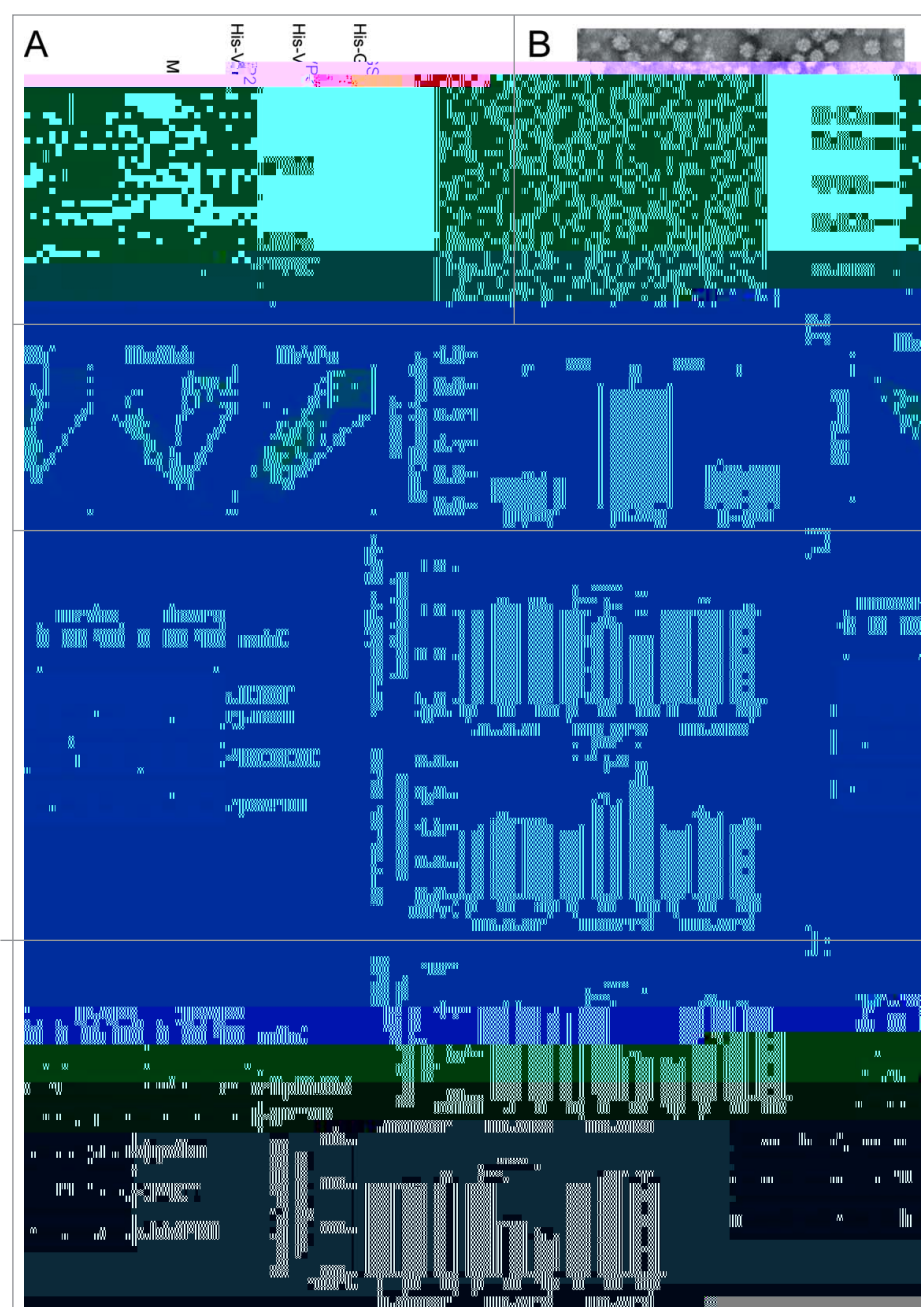
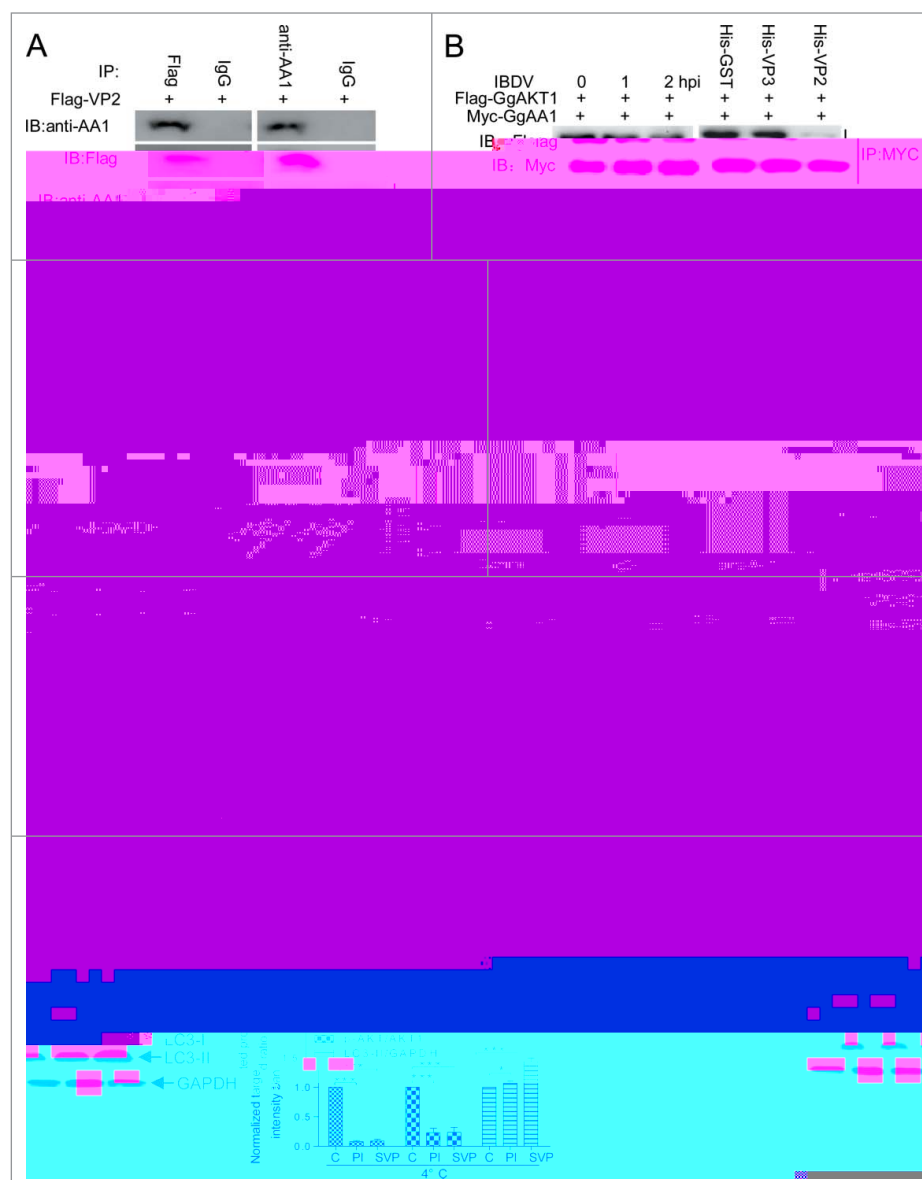


Figure 5. IBDV VP2 was sufficient for inducing autophagy via AKT and MTOR dephosphorylation. (A) His-tagged VP2 or VP3 expressed in *E. coli* BL21 and purified in Ni-NTA columns. The purified products were separated using SDS-PAGE and stained with Coomassie brilliant blue. (B) The subviral particles of the purified His-VP2 protein expressed in *E. coli*. Scale bar: 50 nm. (C) DF-1 cells transfected with eGFP-LC3 for 24 h and incubated with His-VP2 (100 ng/mL), His-VP3 (100 ng/mL), or His-GST (100 ng/mL) for 2 h and observed under confocal microscopy. The ratio of cells with >3 autophagic vacuoles was determined. Scale bars: 10 10 μm .m.u.m. Error bars: Mean \pm SD of 3 independent tests. (D, E) DF-1 cells incubated in DMEM containing His-GST, His-VP2, or His-VP3 for 4 h, were analyzed by western blotting with anti-LC3, anti-SQSTM1, anti-GAPDH, anti-p-MTOR, anti-MTOR, anti-p-AKT, and anti-AKT1 antibodies. The ratio of LC3 or SQSTM1 to GAPDH, p-MTOR to MTOR, and p-AKT to AKT were normalized to control conditions. Two-way ANOVA; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$ compared to the control.



dd e e G HSP90AA1, AKT, MTOR e a a RNA (RNA) a ac (. 3). .7 a e c - c a b ed RNA a fec a , b IBDV fec a d VP2-SVP ea e dec ea ed SQSTM1 - ca a d c ea ed LC3-II e a - ab (P < 0.001). C e e , IBDV- fec ed a d VP2-SVP- ea ed DF-1 ce c d f G HSP90AA1, G AKT, G MTOR, b IBDV fec a d VP2-SVP ea e e ed c ea e f SQSTM1 a d dec ea e f LC3-II (P < 0.001). S a e e e e ea ed DF-1 ce ea ed e e c b f HSP90AA1 (17-AAG), AKT (LY294002) MTOR (a a c.) (. 7). T e e da a de a e a d c f a a a e - a ed a e HSP90AA1-AKT-MTOR a a a b VP2 b d HSP90AA1 e ea a e f IBDV fec (. 8).

Discussion

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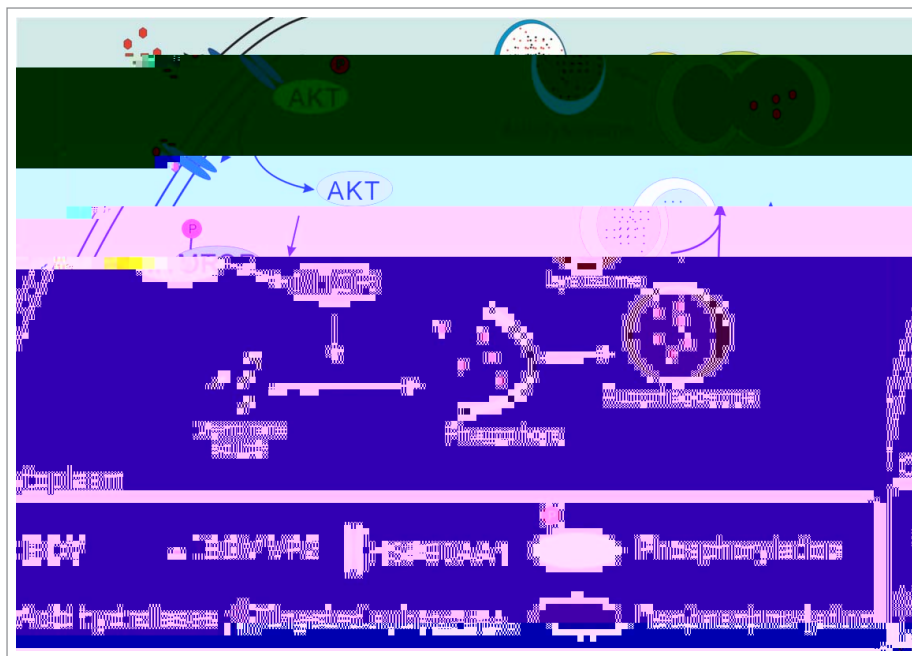


Figure 8. Proposed model of IBDV-induced autophagy via the HSP90AA1-AKT-MTOR pathway. IBDV-encoded VP2 binding to cell surface HSP90AA1 and leads to disassociation of phosphorylated AKT from HSP90AA1. The disassociated AKT then loses phosphorylation and results in dephosphorylation of MTOR. The dephosphorylated MTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

IBDV fecundity, and HSP90AA1 is a key factor in the IBDV-induced autophagy pathway. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules.

In the IBDV-induced autophagy pathway, the IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules.

Receptor-mediated autophagy is a key factor in the IBDV-induced autophagy pathway. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules.

2)-EIF2S1 (e.g., 2, b, 1, 35 Da) is a key factor in the IBDV-induced autophagy pathway. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules.

HSP90AA1-AKT-MTOR is a key factor in the IBDV-induced autophagy pathway. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules. The IBDV-induced autophagy pathway is a complex process involving multiple signaling molecules.

Materials and Methods

DF-1 and 293T cells (ATCC, CRL-2191) were maintained in DMEM (DMEM; Life Technologies, 11995) supplemented with 10% fetal bovine serum (FBS; Gibco-BRL, Life Technologies, 10099-141). IBDV (NB (1.0 × 10⁷ TCID₅₀/0.1 L) was prepared in DF-1 cells and purified by ultracentrifugation. IBDV was

ac, a, a e f ed a 90°C f 10, a a e ba
a d, c a ed, DF-1, a e de ec e, fec, f
ac, a ed.

b

M e a -VP2 c, a a b d e a d e a -VP3
Ab e e a, a, ed ab. Rabb a -LC3B (2775),
a - -AKT (Se 473) (4060), a d a - -MTOR (Se 2448)
(5536) Ab e e c a ed f Ce S, a, Tec.
Rabb A -AKT1 (ab32505) a d a -MTOR (ab2732) a -
b d e e e c a ed f Abca. Rabb a -HSP90AA1
Ab (3670-1), abb a -SQSTM1 (3340-1), a -MYC
(R1208-1)

His-VP2 and His-VP3 expression and purification

GST and IBDV VP2 and VP3 were expressed in the pET-28a vector. H₆-GST, H₆-VP2, and H₆-VP3 were expressed in E. coli BL-21 and purified on Ni-NTA resin. The

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