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# Binding of the pathogen receptor HSP90AA1 to avibirnavirus VP2 induces autophagy by inactivating the AKT-MTOR pathway

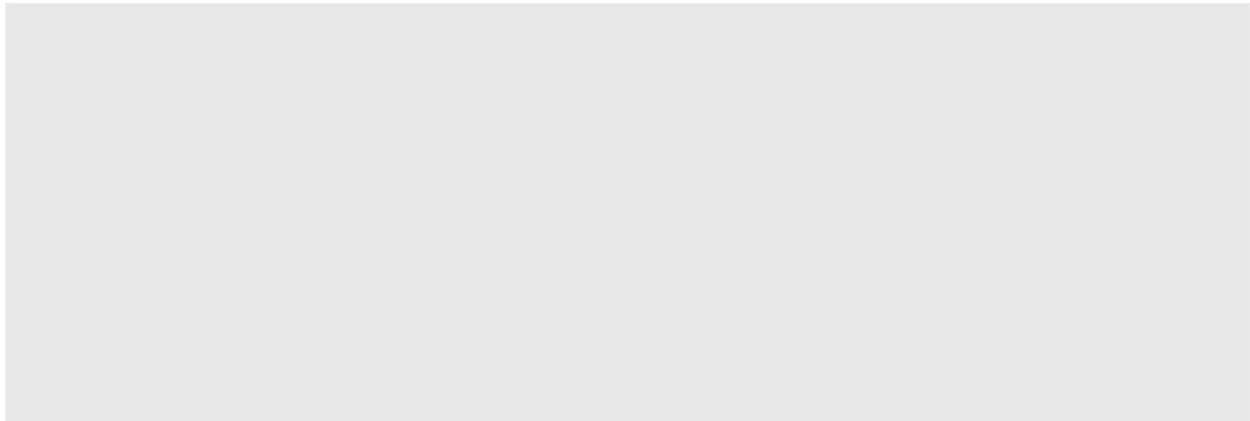
Boli Hu,<sup>1,2,†</sup> Yina Zhang,<sup>1,†</sup> Lu Jia,<sup>1</sup> Huansheng Wu,<sup>1</sup> Chengfei Fan,<sup>1</sup> Yanting Sun,<sup>1</sup> Chengjin Ye,<sup>1</sup> Min Liao,<sup>1</sup> and Jiyong Zhou<sup>1,2,\*</sup>

<sup>1</sup>Key Laboratory of Animal Virology of Ministry of Agriculture; Zhejiang University; Hangzhou, China; <sup>2</sup>College of Veterinary Medicine; Nanjing Agricultural University; Nanjing, China

<sup>†</sup>These authors contributed equally to this work.

: AKT-MTOR pathway, autophagy, avibirnavirus, HSP90AA1, influenza A virus, VP2

: ANOVA, analysis of variance; ATG5, autophagy-related 5; BCA, bicinchoninic acid; BECN1, Beclin-1, autophagy-related; cDNA, complementary DNA; CHIP, chromatin immunoprecipitation; DMEM, Dulbecco's modified Eagle's medium; dsRNA, double-stranded RNA; EBSS, Eagle's basal medium; EIF2AK2, eukaryotic translation initiation factor 2A; EIF2S1, eukaryotic translation initiation factor 2, beta 1; eGFP, enhanced green fluorescent protein; ER, endoplasmic reticulum; *G. gallus* (chicken); GAPDH, glyceraldehyde-3-phosphate dehydrogenase; GOPC, Golgi-associated PDZ domain-containing protein; GST, glutathione S-transferase; HE-IBDV, heterologous infection of IBDV; HE-IBDV, heterologous infection of IBDV; H, hemagglutination; HSP90AA1, heat shock protein 90 class A member 1; HSV-1, herpesvirus 1; IBDV, infectious bursal disease virus; IG, immunoglobulin G; LPS, lipopolysaccharide; Ab, antibody; MAP1LC3/LC3, microtubule-associated protein 1A class C domain family 3; MOI, multiplicity of infection; MTOR, mammalian target of rapamycin (serine/threonine kinase); N-NTA, N-terminal NTA; PAMP, pathogen-associated molecular pattern; PBS, phosphate buffered saline; PI3K, phosphatidylinositol 3-kinase; PRR, pattern recognition receptor; RNAi, RNA interference; SDS, sodium dodecyl sulfate; RNA, RNA



... (TLR),<sup>5</sup> ac... f... a... e... a...  
<sup>6</sup> e... f... a... e... e... a... d... f... a... f...  
 a... e... a... a... a... d... e... e... e... e...<sup>7-12</sup> V... e...  
 b... e... a... a... a... a... e... f... b... a...  
<sup>2,13-15</sup> H... e... e... e... a... a... f... e...  
<sup>16,17</sup> T... e... e... a... e... e... e... c... e... e... ac...  
 b... e... e... a... a... d... a... d... e... e...

A... a... a... c... e... ed... c... a... c... a... a...  
 e... a... e...; e... ed... f... b... a... d... e... e... a... d...  
 a... f... e... de... a...<sup>18</sup> A... a... c... de... ac... a...  
 a... c... a... a... a... d... c... e... e... ed... a... ed... a... a...  
 Mac... a... a... e... e... f... e... a... a... a... a...  
 ce... e... e... c... a... ce... ed... e... e... a... c... ce...<sup>19</sup>  
 A... a... e... ed... e... e... f... e... f... da... a... ed...  
 a... ed... ce... a... a... e... a... d... e... a... e... a... e... a... a...  
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 a... a...<sup>20</sup> Ac... a... f... e... e... a... de... 3... a... e...  
 (PI3K)-AKT-MTOR... a... d... AKT... b... e... ce... c... e...  
 (TSC)-MTOR... a... a... a... b... a... a... a... e... a... e... f...  
 a... a... a... a... ca... d... e... e... e... e... a... e... e... e... f...  
 MTOR.<sup>20</sup> T... e... ef... e... e... e... a... d... ec... b... e... e... a... a...  
 a... d... e... MTOR... a... a... a... a...

U... ace... a... a... e... a... a... a... a... ca... b... e... ac...  
 a... ed... a... a... e... e... e... c... a... f... c... a... fec...<sup>21</sup>  
 T... da... e... a... e... b... e... f... de... e... a... e... ec...  
 e... ea... e... effec... f... a... e... fec... a... a... Pa... e... ec...  
 e... e... (PRR) de... ec... a... e... b... ec... PAMP...  
 E... ace... a... a... e... a... e... ec... ed... a... TLR... e... ce... face...  
 e... d... a... c... a... e...; c... e... e... f... c... a...  
 a... a... d... ced...<sup>22-24</sup> C... a... c... a... e... ec... e... e...  
 ac... a... ace... a... a... e... de... ec... a... d... de... ade... a... d... b... e...  
 a... ded... RNA (dRNA) e... e... e... ca... e... e... ed... a... e...<sup>25</sup>  
 CD46... e... e... e... ce... e... b... a... e... a... e... PRR... e... e... e... a...  
 e... e... a... d... b... a... c... e... f... c... e... f... d... c... f... c... a...  
 a... a... a... a... e... e... (CD46-C... -1)-G... a... a... ca... ed... PDZ... a... d...  
 c... ed... c... f... c... a... e... (GOPC) a... a... a...<sup>26</sup> Hea...  
 e... 90 (HSP90AA1) a... a... a... c... e... ed... ec... a... c... a...  
 e... e... a... a... PRR... c... e... e... b... d... acc... de...  
 (LPS),<sup>27</sup> de... e... e... a... d... a... b... a...<sup>28,29</sup> H... e... e... e...  
 HSP90AA1... e... e... e... e... a... e... a... a... e... e... e... e...

I... e... e... e... d... e... add... e... e... HSP90AA1 ca... d... ce...  
 a... a... a... d... a... a... d... e... a... e... e... e... ca... e... e... a... ce... f...  
 a... ce... a... a... e... b... d... We... f... d... a... HSP90AA1 PRR...  
 a... e... VP2 f... f... a... b... a... fec... b... a... d... ea... e...  
 (IBDV) ac... a... e... MTOR... de... e... de... a... a... a... O... d...  
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 e... MTOR... a... a... da... a... a... a... a...

## Results

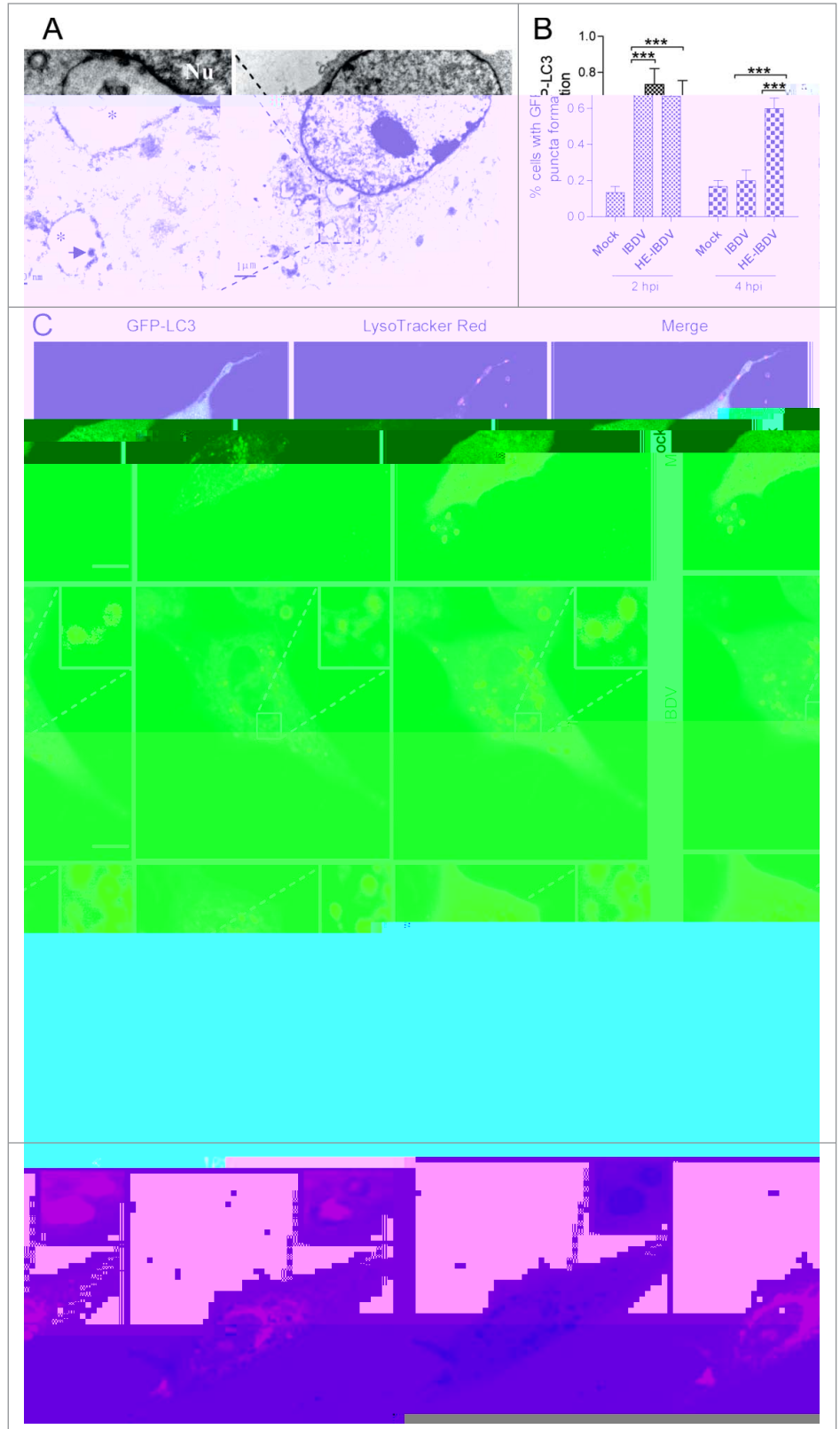
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 e... d... f... b... e... a... a... e... e... ce...<sup>30,31</sup> d... d...

ed... a... a... b... a... a... ce... a... d... a... e... e... e... ca... ed...  
 e... b... a... e... e... ce... fec... ed... DF-1 ce... ( . 1),  
 a... e... e... e... e... ed... a... a... e... c... a... ed... a...  
 b... a... a... d... ce... a... d... ac... a... e... f... fec... ed...  
*Gallus gallus*,<sup>32</sup> d... ca... a... a... a... a... ed...  
 de... ada... C... e... d... a... f... e... DF-1 ce... a... fec... ed...  
 e... a... ced... ee... e... ce... e... e... c... a... 3 (eGFP-LC3)  
 f... 24... e... e... fec... ed... IBDV... c... ba... ed... a... ac... f...  
 a... ed... IBDV (HE-IBDV), e... c... ca... e... e... c... e... f...  
 eGFP-LC3 a... d... L... T... ac... e... Red... a... b... e... d... a... a... e... e...  
 ce... e... e... ca... c... a... ed... c... f... ca... c... c... . 1

a... a... c... a... a... a... c... fec... ed... ce... e... e... e...  
 a... d... a... a... ca... eGFP-LC3... c... e... e... e... ca...  
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 e... e... e... ed... a... c... d... a... c... a... a... a...  
 IBDV... fec... ed... ce... b... HE-IBDV... ea... ed... ce... a... 4... T...  
 e... f... IBDV... e... e... e... a... e... f... a... a...  
 IBDV... fec... ed... eGFP-LC3... a... b... e... d... ce... e... a... ed...  
 c... c... a... a... b... d... ( Ab) a... a... e... VP2 f... IBDV... C...  
 f... ca... c... c... e... ea... ed... VP2... a... a... a... d... ed... b...  
 e... e... e... eGFP-LC3 ( . 1). C... ec... e... e... e... da... a...  
 d... ced... IBDV... fec... ed... a... d... HE-IBDV... ea... ed... ce... a... d...  
 IBDV... a... e... ca... ed... a... a... e... e...

T... e... e... e... a... f... IBDV... e... e... a... e... b... ed... a...  
 ab... 6...<sup>32</sup> T... e... ef... e... de... e... e... e... a... a... a...  
 e... ed... a... e... ca... a... e... f... fec... , e... e... a... ed... ce... fec... ed...  
 IBDV... a... a... c... f... fec... (MOI) f... 10 f... 0... 8...  
 e... e... a... e... c... a... e... e... d... e... LC3-II, a... a... a... e...  
 a... e...<sup>30</sup> . 2... a... a... a... c... a... c... fec... ed...  
 ce... e... a... f... ace... a... LC3-II... IBDV... fec... ed... ce... a...  
 c... ea... ed... a... 1... dec... ea... ed... e... a... ab... a... 4... a... d... ec...  
 e... ed... ca... a... 8... ( $P < 0.01$ ). H... e... e... ace... a... LC3-II...  
 e... e... HE-IBDV... ea... ed... ce... a... e... a... ed... e... e...  
 f... 1... 4... ( $P < 0.001$ ) a... d... b... e... a... dec... ea... ef... 4...  
 S... a... LC3-II... e... e... e... e... e... b... e... ed... IBDV... fec... ed...  
 HE-IBDV... ea... ed... 293T ce... ( . 1). I... add... LC3... e...  
 e... a... ed... e... a... a... c... b... de... ec... e... acc... a... f...  
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 IBDV... fec... ed... ce... dec... ea... ed... e... e... f... 1... 6... ( $P <$   
 $0.001$ ) a... d... b... e... a... c... ea... ea... 8... , d... ca... a... e... a...  
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 HE-IBDV... ea... ed... ce... dec... ea... ed... e... e... f... 1... 8...  
 ( $P < 0.001$ ), b... d... d... c... a... e... b... c... fec... ed... ce...  
 I... e... e... e... a... f... LC3-II... c... e... a... d... dec... ea... ef...  
 SQSTM1 acc... a... e... e... a... de... ec... ed... VP2... a... fec... ed...  
 293T ce... a... 48... af... e... a... fec... ( $P < 0.001$ , . 2). T...  
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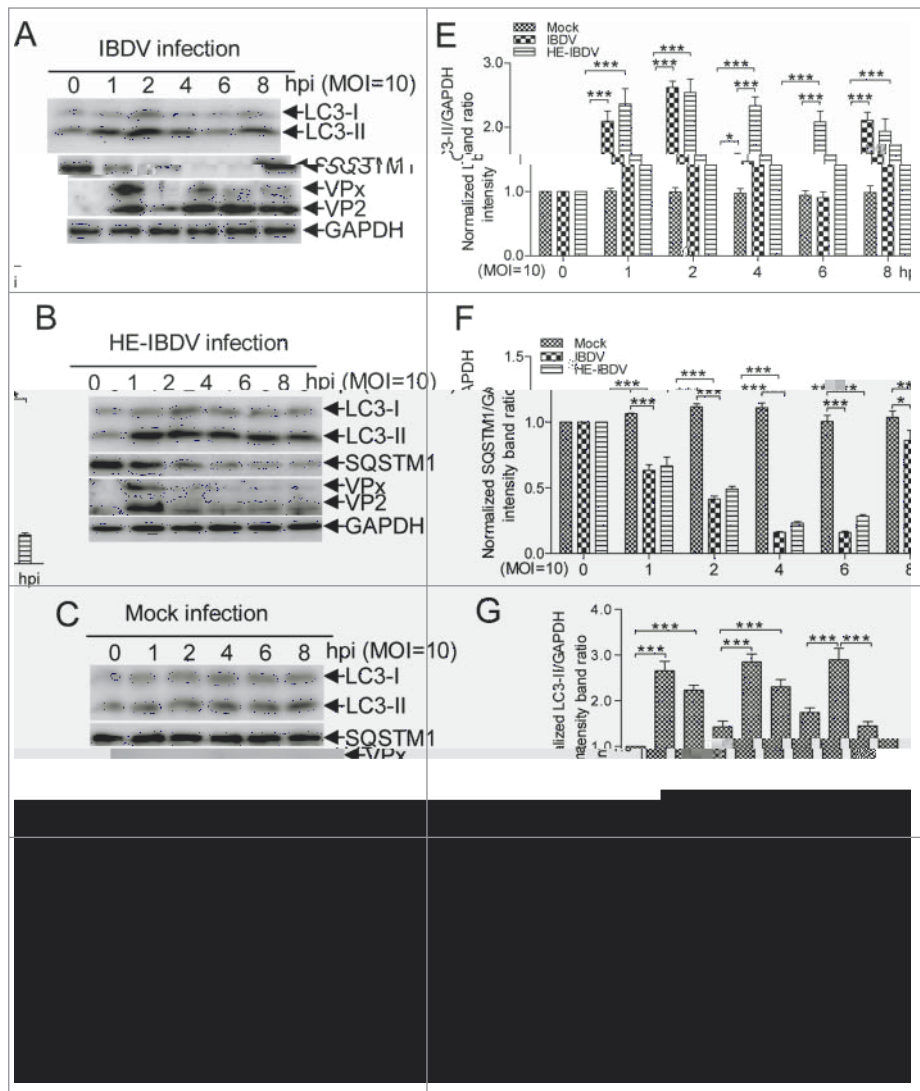
**Figure 1.** IBDV infection induces autophagosome accumulation in DF-1 cells. **(A)** Autophagic vacuoles in infected cells observed by transmission electron microscopy. Autophagic vacuole engulfs IBDV particle (black arrows) and organelle (asterisk) in cytoplasm of DF-1 cell infected with IBDV (MOI = 10) at 2 hpi. **(B)** DF-1 cells transfected with pEGFP-LC3 for 48 h, and then infected with IBDV (MOI = 10) or treated with HE-IBDV. At 2 hpi or 4 hpi, cells were incubated with LysoTracker Red (50 nM) for 30 min; intracellular autophagic vacuoles were observed under confocal microscopy. Statistical analysis of the number of cells with >3 autophagic vacuoles. At 2 hpi or 4 hpi, autophagic vacuoles were counted in IBDV-infected cells. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way analysis of variance (ANOVA); \*\*\* $P$  < 0.001 compared to control. **(C)** DF-1 cells transfected with pEGFP-LC3 for 48 h, and then infected with IBDV (MOI = 10) or treated with HE-IBDV. At 2 hpi or 4 hpi, cells were incubated with LysoTracker Red (50 nM) for 30 min; intracellular autophagic vacuoles were observed under confocal microscopy. Scale bars: 10  $\mu$ m. **(D)** DF-1 cells transfected with pEGFP-LC3 and infected with IBDV at 9 h post-transfection. The cells were fixed, immunostained with anti-VP2 mAb, and autophagosomes were observed under confocal microscopy. Scale bar: 10  $\mu$ m.



IBDV. The number of DF-1 cells infected with IBDV (MOI = 10) was significantly higher than that of HE-IBDV (MOI = 10) ( $P < 0.001$ ). Mock, HE-IBDV, and IBDV-infected cells were immunostained with anti-VP2 mAb. IBDV-infected cells were observed at 2 hpi and 4 hpi (MOI = 10) (Fig. 1D). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B).

The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B).

The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B). The number of cells with >3 autophagic vacuoles was significantly higher in IBDV-infected cells than in HE-IBDV-infected cells at 2 hpi and 4 hpi (Fig. 1B).



**Figure 2.** Characterization of IBDV-triggered autophagosome accumulation. (A) IBDV promotes increase of LC3-II and decrease of SQSTM1 within 1 hpi but inhibits it from 4 hpi. DF-1 cells were infected with IBDV (MOI = 10). (B) Increase of LC3-II and decrease of SQSTM1 was constantly promoted in HE-IBDV-treated cells. (C) LC3 amount and SQSTM1 accumulation in mock-infected cells were unaffected. Cells were harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. (D) DF-1 cells were infected with the purified infectious IBDV virions or treated with the purified heat-inactivated IBDV virions (MOI = 10), respectively. The purified infectious IBDV increases the amount of LC3-II within 1 hpi but inhibits it from 4 hpi. However, levels of LC3-II increase constantly in the purified HE-IBDV-treated cells. M, mock infected. HI, purified heat-inactivated IBDV virions. PI, purified infectious IBDV virions. (E, F) The ratio of LC3-II or SQSTM1 to GAPDH was normalized to control conditions in (A to C). (G) The ratio of LC3-II to GAPDH was normalized to control conditions in (D). (H) LC3-II increases and SQSTM1 decreases in VP2-transfected cells. 293T cells were transfected with the vector pFlag-VP2 for 48 h, harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way ANOVA, \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001 compared to control.

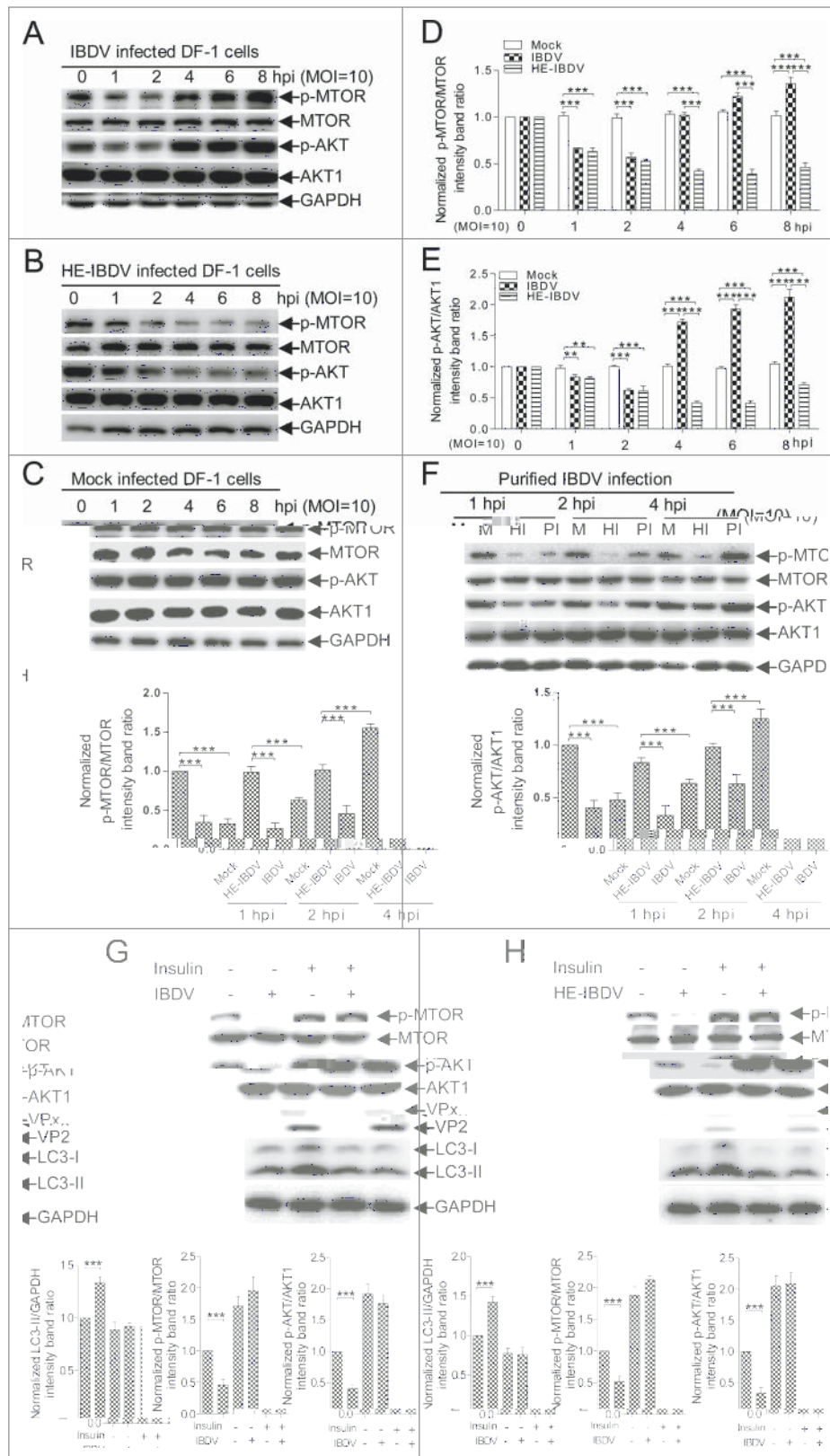
infected DF-1 cells ( $P < 0.001$ ,  $n = 3$ ) and HE-IBDV-infected 293T cells ( $P < 0.001$ ,  $n = 3$ ) and HE-IBDV-infected HE-293T cells ( $n = 3$ ),  $P < 0.001$ ), respectively. IBDV-infected DF-1 cells, HE-IBDV-infected HE-293T cells, and HE-IBDV-infected HE-293T cells were transfected with the vector pFlag-VP2 for 48 h, harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way ANOVA, \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001 compared to control.

infected DF-1 cells ( $P < 0.001$ ,  $n = 3$ ) and HE-IBDV-infected HE-293T cells ( $P < 0.001$ ,  $n = 3$ ) and HE-IBDV-infected HE-293T cells ( $n = 3$ ),  $P < 0.001$ ), respectively. IBDV-infected DF-1 cells, HE-IBDV-infected HE-293T cells, and HE-IBDV-infected HE-293T cells were transfected with the vector pFlag-VP2 for 48 h, harvested and analyzed by immunoblotting using anti-LC3, anti-SQSTM1, anti-VP2, or anti-GAPDH antibody. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way ANOVA, \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001 compared to control.

We evaluated AKT and mTOR activity in HE-293T cells infected with IBDV or HE-IBDV. IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to mock-infected cells ( $P < 0.001$ ). HE-IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to mock-infected cells ( $P < 0.001$ ). IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to HE-IBDV-infected HE-293T cells ( $P < 0.001$ ). IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to HE-IBDV-infected HE-293T cells ( $P < 0.001$ ).

The data suggest that IBDV and HE-IBDV promote autophagosome accumulation in HE-293T cells by increasing AKT activity and decreasing mTOR activity. This effect is not observed in HE-293T cells transfected with VP2 alone ( $P > 0.05$ ). IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to HE-IBDV-infected HE-293T cells ( $P < 0.001$ ). IBDV-infected HE-293T cells showed a significant increase in AKT activity and a significant decrease in mTOR activity compared to HE-IBDV-infected HE-293T cells ( $P < 0.001$ ).

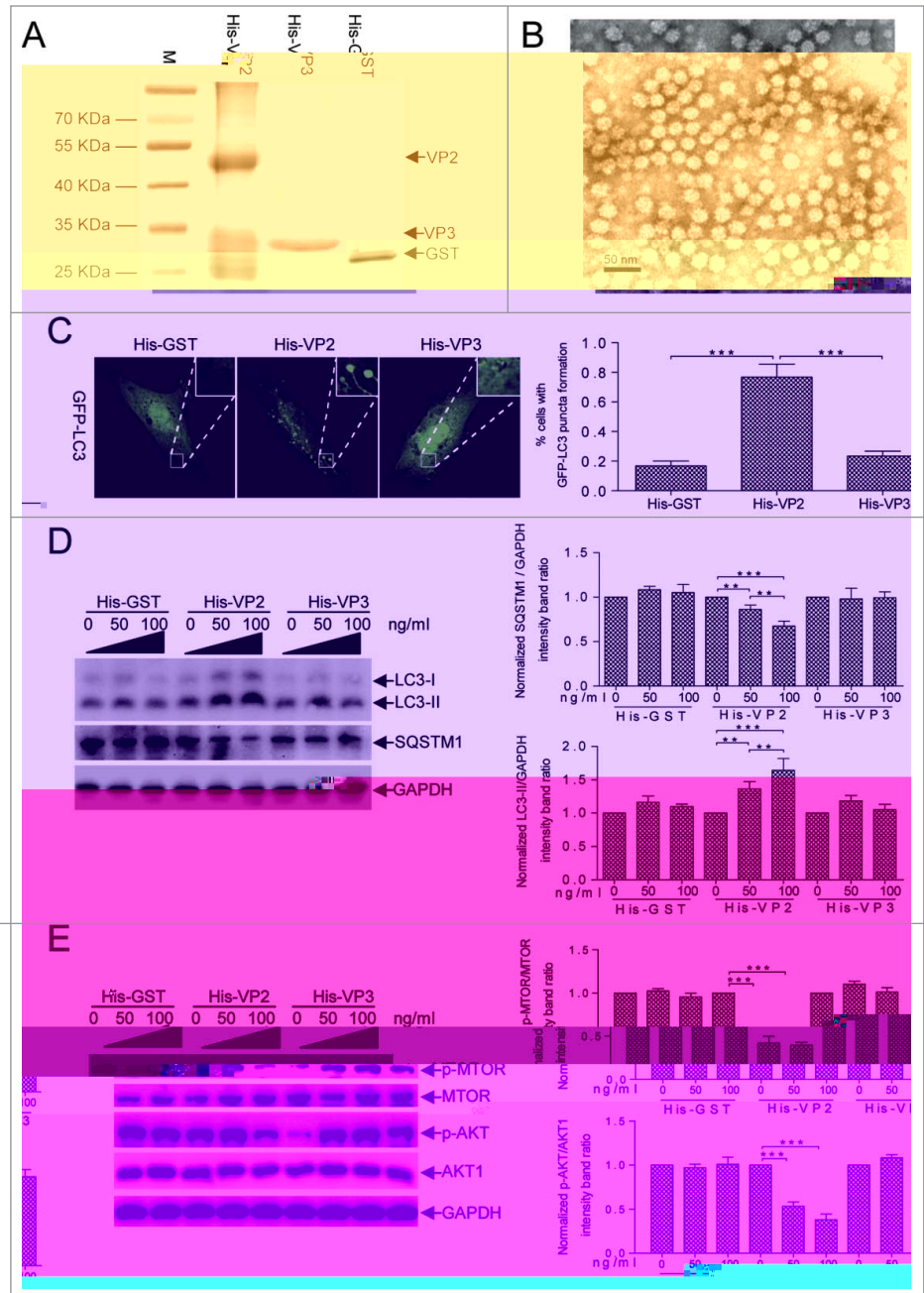




**Figure 4.** MTOR and AKT were inactivated at the early stage of IBDV infection. **(A)** DF-1 cells infected with IBDV (MOI = 10). Cells were harvested at 1, 2, 4, 6, and 8 h followed by immunoblotting with anti-p-MTOR, anti-MTOR (total protein), anti-p-AKT, anti-AKT1, or anti-GAPDH antibody. **(B)** Cells incubated with HE-IBDV for 1, 2, 4, 6, and 8 h followed by immunoblotting as in **(A)**. **(C)** Cells mock-infected and analyzed by immunoblotting as in **(A)**. **(D, E)** The ratio of p-MTOR to MTOR or p-AKT to AKT was normalized to mock infection and set at 1.0. **(F)** Cells were infected with purified IBDV or treated with purified HE-IBDV for 1, 2, and 4 h, and analyzed by immunoblotting using the antibody in **(A)**. **(G)** DF-1 cells pretreated with insulin (500 nM) for 1 h, infected with IBDV and cultured for 2 h, and processed by immunoblotting using the corresponding antibody in **(A)**. **(H)** DF-1 cells pretreated with insulin (500 nM) for 1 h, incubated with HE-IBDV for 2 h, and processed for immunoblotting using the antibody in **(A)**. The ratio of LC3 to GAPDH, p-MTOR to MTOR, or p-AKT to AKT was normalized to control conditions and set at 1.0. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way ANOVA; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$  compared to control.

H -VP2 SVP, H -VP3, and H -VP4 (GST) were used for immunoblotting. LC3-II and SQSTM1 were used as markers for autophagy. H -VP2, H -VP3, and H -VP4 were used as markers for IBDV infection. The ratio of LC3-II to GAPDH, p-MTOR to MTOR, or p-AKT to AKT was normalized to control conditions and set at 1.0. Error bars: Mean  $\pm$  SD of 3 independent tests. Two-way ANOVA; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$  compared to control.

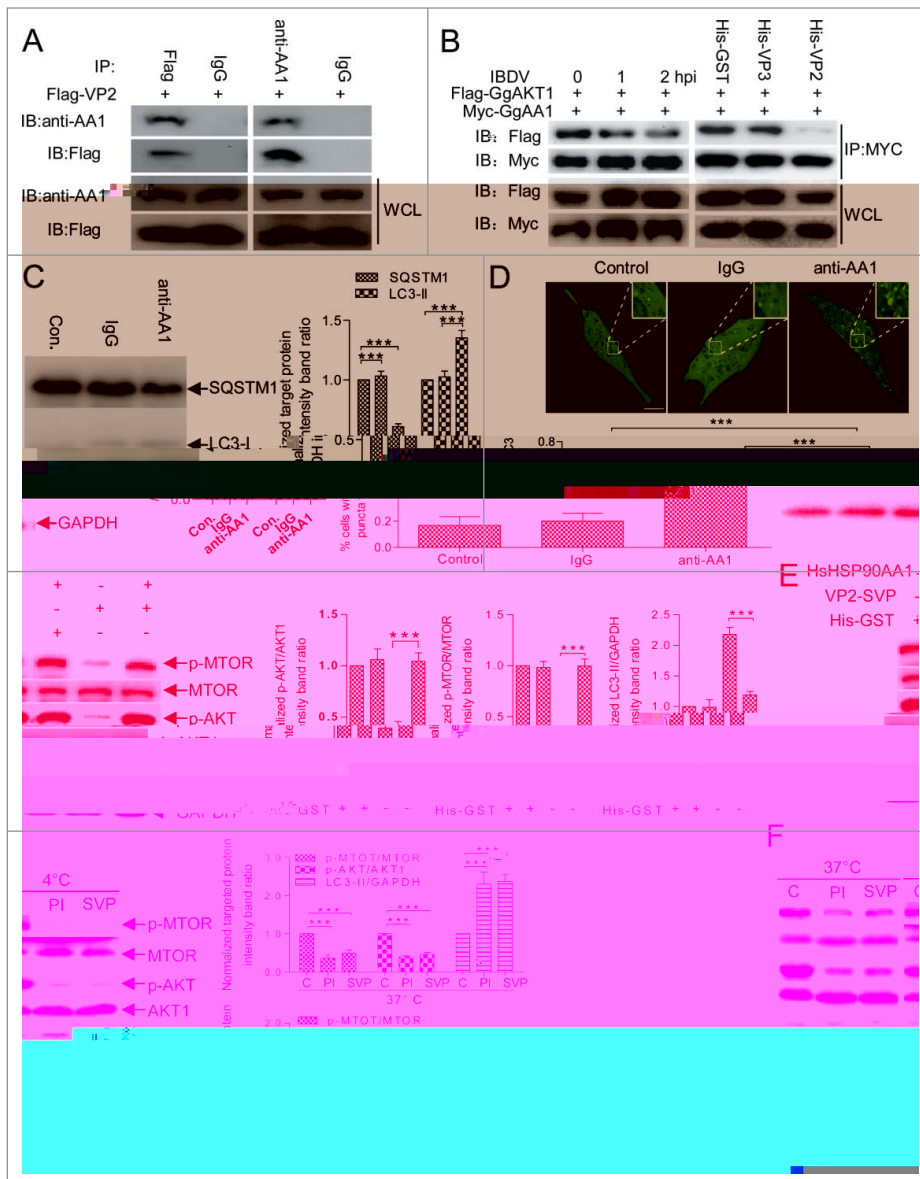
Ce e b a e face-d b ed  
 G HSP90AA1 a a e e ce  
 ec e IBDV;<sup>29</sup> c a c  
 HSP90 a f a a  
 AKT a e ac e ac  
 AKT.<sup>36</sup> We e a ed  
 ce a G HSP90AA1 ed  
 a ed VP2- d ced a a e  
 AKT-MTOR a a . We de ced  
 VP2 a d G HSP90AA1 e ac  
 a d AKT-MTOR a a a d  
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 a d c ce (.6). L e e  
 ce ce a e ea ed  
 ca ce a e ea ed a  
 ac e a -HSP90AA1- ea ed  
 ce a fec ed GFP-LC3 ( $P < 0.01$ , .6), b e e a  
 I G- ea ed ce . C e d ,  
 VP2 SVP- ea ed DF-1 ce ad  
 ca dec ea e f -MTOR  
 a d -AKT e e ( $P < 0.001$ )  
 a d a e a be ce a e f LC3-II  
 ( $P < 0.001$ ), e e , e a  
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 a ac a a e e a e de e de . T a da e d ced a a e a ed a a f IBDV  
 HSP90AA1-AKT-MTOR a a ed a f IBDV- fec a d VP2 ea e b c c DF-1 ce a



**Figure 5.** IBDV VP2 was sufficient for inducing autophagy via AKT and MTOR dephosphorylation. (A) His-tagged VP2 or VP3 expressed in *E. coli* BL21 and purified in Ni-NTA columns. The purified products were separated using SDS-PAGE and stained with Coomassie brilliant blue. (B) The subviral particles of the purified His-VP2 protein expressed in *E. coli*. Scale bar: 50 nm. (C) DF-1 cells transfected with eGFP-LC3 for 24 h and incubated with His-VP2 (100 ng/mL), His-VP3 (100 ng/mL), or His-GST (100 ng/mL) for 2 h and observed under confocal microscopy. The ratio of cells with >3 autophagic vacuoles was determined. Scale bars: 10 10 μm.m;μm. Error bars: Mean ± SD of 3 independent tests. (D, E) DF-1 cells incubated in DMEM containing His-GST, His-VP2, or His-VP3 for 4 h, were analyzed by western blotting with anti-LC3, anti-SQSTM1, anti-GAPDH, anti-p-MTOR, anti-MTOR, anti-p-AKT, and anti-AKT1 antibodies. The ratio of LC3 or SQSTM1 to GAPDH, p-MTOR to MTOR, and p-AKT to AKT were normalized to control conditions. Two-way ANOVA; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$  compared to the control.

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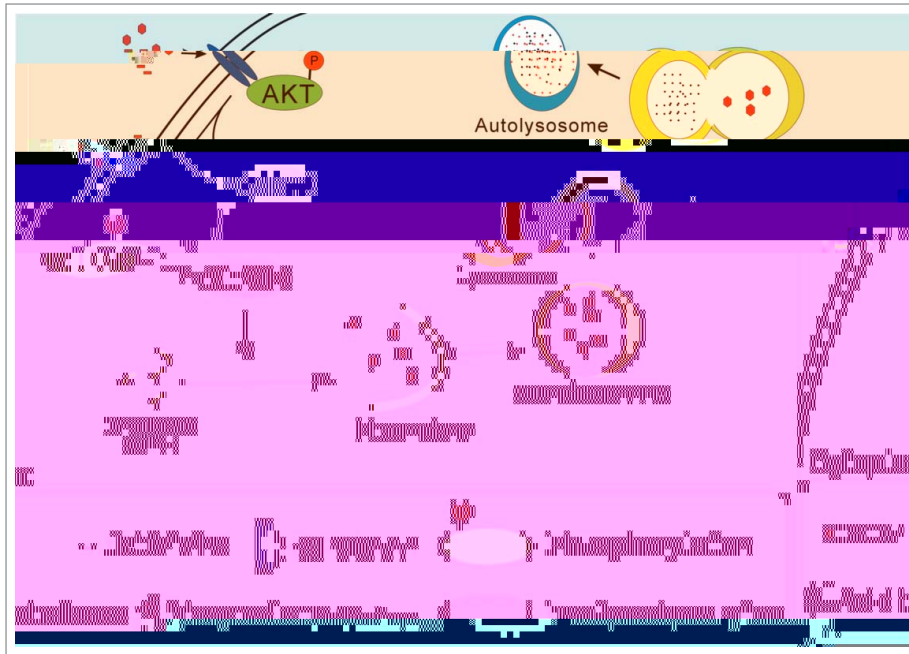
**Figure 6.** HSP90AA1 binding to VP2 triggers autophagy via AKT-MTOR dephosphorylation. **(A)** DF-1 cells transfected with pFlag-VP2 for 24 h. Whole cell lysates (WCL) were used for CoIP and western blotting with anti-Flag or anti-HSP90AA1 antibody (anti-AA1) and irrelevant IgG (Control). **(B)** DF-1 cells cotransfected with Myc-GgHSP90AA1 (Myc-GgAA1) and Flag-GgAKT1 for 48 h. Transfected cells were infected with IBDV for 1 or 2 h, or were incubated in DMEM containing His-VP2 (100 ng/ml), His-VP3 (100 ng/ml) or His-GST (100 ng/ml). Whole cell lysates of each sample were used for CoIP with anti-MYC antibody and western blotting with anti-Flag or anti-MYC antibody. **(C)** Western blotting performed using anti-LC3 antibody and anti-SQSTM1 mAb on lysates from DF-1 cells cultured in uncoated plates or in coated plates with anti-HSP90AA1 or irrelevant isotype control IgG for 4 h. The ratio of SQSTM1 or LC3-II to GAPDH was normalized to control conditions. **(D)** DF-1 cells transfected with pGFP-LC3 for 24 h and cultured in plates coated with negative control, IgG, or anti-HSP90AA1 for 4 h. Autophagic vacuoles were analyzed under confocal microscopy. The ratio of cells containing >3 ring-like GFP structures was determined. Scale bars: 10 10  $\mu$ m.mu;m. Error bars: Mean  $\pm$  SD of 3 independent tests. **(E)** DF-1 cells were incubated respectively with the His-GST, mixture of His-GST and HSP90AA1 (His-GST:HSP90AA1 = 1:2.5), mixture of SVP and HSP90AA1 (SVP:HSP90AA1 = 1:2.5) or SVP for 2 h, and analyzed by immunoblotting with anti-LC3, anti-p-MTOR, anti-MTOR, anti-p-AKT, anti-AKT1, or anti-GAPDH antibody. **(F)** DF-1 cells incubated with His-GST (100 ng/ml), SVP (100 ng/ml), or purified IBDV (MOI = 10) for 2 h at 37°C or 4°C. Cells were analyzed by immunoblotting using the antibody in **(E)**. The ratio of p-MTOR to MTOR, p-AKT to AKT or LC3-II to GAPDH was normalized to mock infection and set at 1.0. Two-way ANOVA; \*\*\* $P$  < 0.001 compared to control. C, His-GST; PtdIns, purified IBDV; SVP, His-VP2 subviral particle.

dd. e e G HSP90AA1, AKT, MTOR RNA (RNA) a ac ( . 3). .7 ca bed RNA a fec a, b IBDV fec a d VP2-SVP ea e dec ea ed SQSTM1 ca a d cea ed LC3-II e a ab ( $P$  < 0.001). C e e e, IBDV- fec ed a d VP2-SVP- ea ed DF-1 ce e c d f G HSP90AA1, G AKT, G MTOR, b IBDV fec a d VP2-SVP ea e e ed c ea e f SQSTM1 a d dec ea e f LC3-II ( $P$  < 0.001). S a e e e e ea ed DF-1 ce ea ed e e e ea ed ec c b f HSP90AA1 (17-AAG), AKT (LY294002) MTOR (a a c.) ( . 7). T e e da a de e e a d c f a a e a e a ed a e HSP90AA1-AKT-MTOR a a a a b VP2 b d HSP90AA1 e ea a e f IBDV fec ( . 8).

## Discussion

See a e e e de e a ed e e e e e e ed a e e f e a a a b e c d ce a e e, c a ICP34.5 f e e e e 1 (HSV-1), Nef f HIV, a d M2 f e a A .<sup>2,13,37</sup> O. e da e d c ea e e e S, db e fec d ce a a a a e a e e a ed 5 e e (ATG5).<sup>38</sup> I e d, a e e e c c e ea ed IBDV a ce e de a e e a da e e, e e e e fed e e e eGFP-LC3 c e a b e ed de c f ca c c . I d c f a b a a a a c dec ea ed VP2 acc a a d IBDV e ; c a , b f a e e e BECN1 c d ( . 3) ce a ed VP2 acc a a d a e . T e e e e e e ac a f ce a a a a defe e e e IBDV a .

T ca e , fec , , e  
de e , cec ac d  
ac ce e ba e ,  
a e ce c a . TLR3,  
TLR4, a d TLR7 , a a  
a d PGLYRP1 ( e d ca  
ec , e, 1)-LE , *Dro-*  
*sophila* d ce a ,  
a d b, d, 5,23,24,39 T  
ec a ce e e HSP90AA1  
fac ae a a , f a  
de a e f e, a d a a  
c ca e , a ce a  
f, c . Of e e fac  
HSP90AA1 a a a e,  
ece b, d LPS<sup>27</sup> a d  
de e ,<sup>28</sup> I a ,  
G HSP90AA1 a a ce ece -  
c e c e, f IBDV  
a ce.<sup>29</sup> H e e, ec -  
a c, ec , be ee, ce  
ece a d a e a,  
de d. I d ,  
a a d ced ed -  
a e 2 f , a b -  
a , fec ( . 1 2);  
HSP90AA1 e, ed f -  
a , fea a d a ea -  
e e ef ce,  
e e ed a d de aded IBDV  
a ce ed ae ( . 6).  
T e a c, ac e, c -  
a, IBDV, e e -  
be ed , IBDV- fec ed  
ac e a d ce.<sup>32</sup>  
T de a e  
G HSP90AA1 c ea a -  
b a , fec , b e .  
N ab , f a b a -  
e, c ded e, , VP2 e -  
ac ed G HSP90AA1 a d  
e ed a ( . 5



**Figure 8.** Proposed model of IBDV-induced autophagy via the HSP90AA1-AKT-MTOR pathway. IBDV-encoded VP2 binding to cell surface HSP90AA1 and leads to disassociation of phosphorylated AKT from HSP90AA1. The disassociated AKT then loses phosphorylation and results in dephosphorylation of MTOR. The dephosphorylated MTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

IBDV fecundity and HSP90AA1 activation are essential for the replication and transcription of IBDV. The IBDV VP2 protein binds to HSP90AA1 and activates the AKT-MTOR pathway. The activated AKT-MTOR pathway leads to the phosphorylation of Akt and the inhibition of mTOR. The phosphorylated Akt then dissociates from HSP90AA1 and the dephosphorylated mTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

The IBDV VP2 protein binds to HSP90AA1 and activates the AKT-MTOR pathway. The activated AKT-MTOR pathway leads to the phosphorylation of Akt and the inhibition of mTOR. The phosphorylated Akt then dissociates from HSP90AA1 and the dephosphorylated mTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

Receptor-mediated autophagy is a cellular process that involves the formation of autophagosomes. The formation of autophagosomes is regulated by several signaling pathways, including the AKT-MTOR pathway. The AKT-MTOR pathway is a central signaling pathway that regulates cell growth and survival. The activation of AKT-MTOR leads to the phosphorylation of Akt and the inhibition of mTOR. The phosphorylated Akt then dissociates from HSP90AA1 and the dephosphorylated mTOR then activates autophagosome formation.

IBDV VP2 protein binds to HSP90AA1 and activates the AKT-MTOR pathway. The activated AKT-MTOR pathway leads to the phosphorylation of Akt and the inhibition of mTOR. The phosphorylated Akt then dissociates from HSP90AA1 and the dephosphorylated mTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

The IBDV VP2 protein binds to HSP90AA1 and activates the AKT-MTOR pathway. The activated AKT-MTOR pathway leads to the phosphorylation of Akt and the inhibition of mTOR. The phosphorylated Akt then dissociates from HSP90AA1 and the dephosphorylated mTOR then activates autophagosome formation. The autophagosome engulfs IBDV virions, delivering them to lysosomes for final degradation.

**Materials and Methods**

DF-1 and 293T cells (ATCC) were cultured in DMEM (Life Technologies, 11995) supplemented with 10% fetal bovine serum (Gibco-BRL Life Technologies, 10099-141). IBDV (NB (1.0 × 10<sup>7</sup> TCID<sub>50</sub>/0.1 L) was prepared in ab. IBDV-infected DF-1 cells were harvested and centrifuged to obtain IBDV.

ac, a a e f ed a 90°C f 10 , a a e ba  
a d c a ed DF-1 a e de ec fec f  
ac, a ed

M e a -VP2 c a a b d e a d e a -VP3  
Ab e e a a ed ab. Rabb a -LC3B (2775),  
a - -AKT (Se 473) (4060), a d a - -MTOR (Se 2448)  
(5536) Ab e e c a ed f Ce S a Tec  
Rabb A -AKT1 (ab32505) a d a -MTOR (ab2732) a -  
b d e e c a ed f Abca Rabb a -HSP90AA1  
Ab (3670-1), abb a -SQSTM1 (3340-1), a -MYC

*His-VP2 and His-VP3 expression and purification*

GST and IBDV VP2 and VP3 were expressed in *E. coli* BL-21 using PET-28a vector. His-GST, His-VP2, and His-VP3 were purified using Ni-NTA resin.

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