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Solid-state fermentation of corn-soybean meal mixed feed with *Bacillus subtilis* and *Enterococcus faecium* for degrading antinutritional factors and enhancing nutritional value

Changyou Shi, Yu Zhang, Zeqing Lu and Yizhen Wang*

Abstract

Background: Corn and soybean meal (SBM) are two of the most common feed ingredients used in pig feeds. However, a variety of antinutritional factors (ANFs) present in corn and SBM can interfere with the bioavailability of nutrients and have negative health effects on the pigs. In the present study, two-stage fermentation using *Bacillus subtilis* followed by *Enterococcus faecium* was carried out to degrade ANFs and improve the nutritional quality of corn and SBM mixed feed. Furthermore, the microbial composition and in vitro nutrient digestibility of inoculated mixed feed were determined and compared those of the uninoculated controls.

Results: During the fermentation process, *B. subtilis* and lactic acid bacteria (LAB) were the main dominant bacteria in the solid-state fermented inoculated feed, and fermentation produced a large amount of lactic acid (170 mmol/kg), which resulted in a lower pH (5.0 vs. 6.4) than the fermented uninoculated feed. The amounts of soybean antigenic proteins (β -conglycinin and glycinin) in mixed feed were significantly decreased after first-stage fermentation with *B. subtilis*. Inoculated mixed feed following two-stage fermentation contained greater concentration of crude protein (CP), ash and total phosphorus (P) compared to uninoculated feed, whereas the concentrations of neutral detergent fiber (NDF), hemicellulose and phytate P in fermented inoculated feed declined ($P < 0.05$) by 38%, 53%, and 46%, respectively. Notably, the content of trichloroacetic acid soluble protein (TCA-SP), particularly that of small peptides and free amino acids (AA), increased 6.5 fold following two-stage fermentation. There was no difference in the total AA content between fermented inoculated and uninoculated feed. However, aromatic AAs (Phe and Tyr) and Lys in inoculated feed increased, and some polar AAs, including Arg, Asp, and Glu, decreased compared with the uninoculated feed. In vitro dry matter and CP digestibility of inoculated feed improved ($P < 0.05$) compared with the uninoculated feed.

Conclusion: Our results suggest that two-stage fermentation using *B. subtilis* followed by *E. faecium* is an effective approach to improve the quality of corn-soybean meal mixed feed.

Keywords: Antinutritional factors (ANFs), In vitro digestibility, Mixed feed, Nutritional value, Two-stage fermentation

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Background

C... (BM) ... C... H...
- BM
(ANF),
1.
2, 3. P
4. F
5.
P
ANF 6.
I C... (F),
ANF
7., 8.,
9.
F. H
F
F
(FLF)
A
FLF -FLF
H FLF
10. I BM
Bacillus subtilis
ANF, *Enterococcus faecium*
H

Methods

Micoo gani m and ba al \ b a e

B. subtilis J12-1
(). *B. subtilis* J12-1

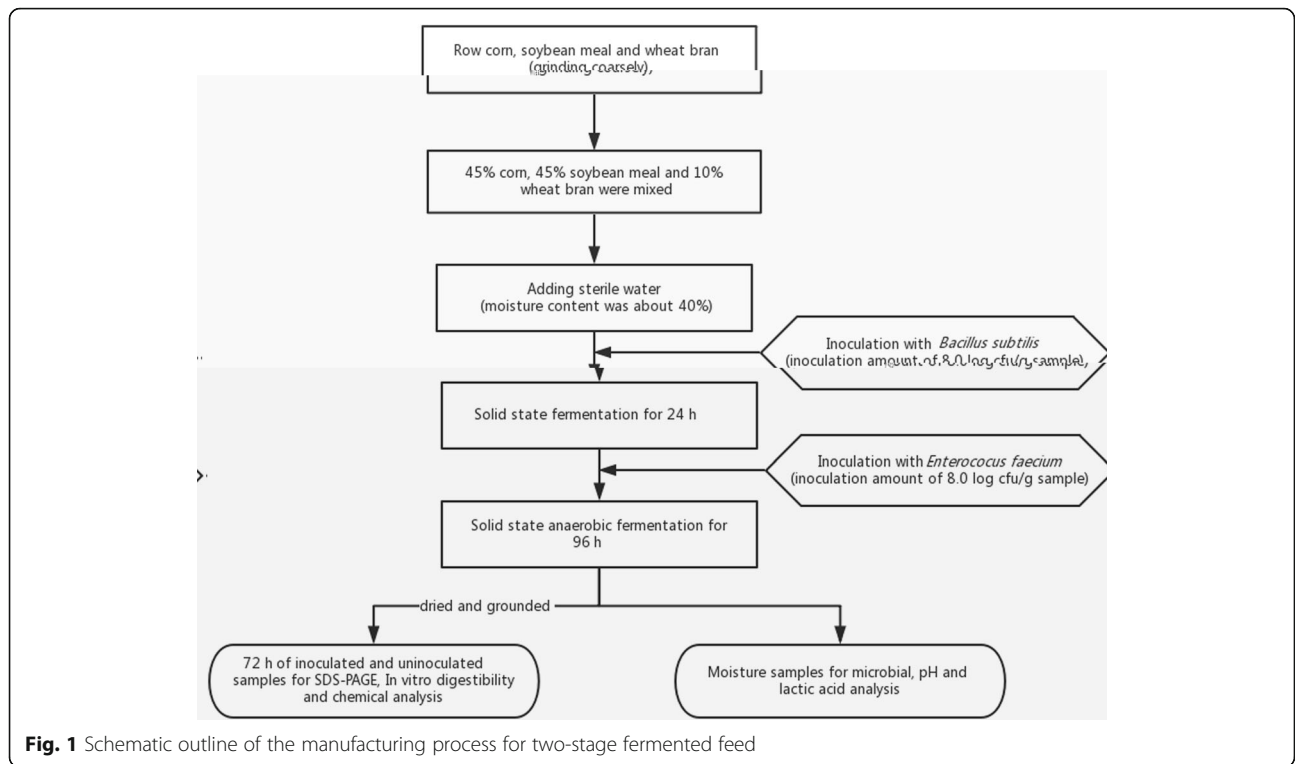


Fig. 1 Schematic outline of the manufacturing process for two-stage fermented feed

O L D, B , H , E)
30 C 2 . N . B
Bacillus
37 C
1 .

Chemical anal i

(24 *Bacillus subtilis*
48 *Enterococcus faecium*)
(DM), (CP),
(NDF), (ADF),
(C) (P)
AOAC I (2005).
(CA- P)
O
11.
N D. 12.
P P 28.2%
(L-8900; H , , J). B
6 /L HC
110 C 24 .M
M
H
() HI
99163 H (H , , I,
A) 2 . 18 L
20 μ

(N - J . B
C , N , C)
β-
(ELI A) (L B C , B , C).
Sodi m dodecyl sulfonate polyacrylamide gel
electrophoresis (SDS-PAGE)
E 13.
60-
1.5 L -HC (20 /L,
H 8.3), 0.1% D , 5 /L
5 μ / L 0.1
30
14,000, g 10
4 C (5804 , E , G),
E
B - P A K (B , A).
D -PAGE
14.
4 - 12%
0.1% D . A
65 120 . A

26616 (10–170 D) A
 C B B -250 (B
 A) 45 7%

In vitro digestion

A 15,
 I 150 L
 E 10,000 / L
 (: 3,000 / ,) (0.05 /L
 KC-HC H 2.0)
 39 C 150 () 4 H
 7.0 1 /L N OH, 150
 (: 250 / ,)
 39 C 150 4 A
 , 5 L 20%
 30
 3,000 15 ,
 105 C 4 CP
 AA I (%) = (
) /
 100%.

Statistical analysis

G L M A
 (A , 1999). A 0.05

Results

Microbial composition, pH and lactic acid concentration during SSF

F 2 H
 F.
 , *B. bacillus* LAB
 (F. 2). *B. subtilis*
 8.0 / . A 24
 9.6 / ,
 LAB
 (<3.0 /),
 LAB 24 8.1 /
E. faecium; 9.6 /
 48 D.
 LAB
B. subtilis. N ,
Enterobacteriaceae ()
 (8.3 /). 24 H ,

Enterobacteriaceae

(<3.0 /).
B. subtilis *E. faecium*
 H
 (F. 2).
 H
B. subtilis (6.8 24
 6.4 0
). A *E. faecium*, H
 6.8 5.0 A
 31 170 /

Biodegradation of soybean antigenic protein of mixed feed after fermentation

O 23–80 D
 (F. 3).
 β- α, α
 β,
 F *B. subtilis*
 α α β-
 F. I ,
 (<25 D)
 H ,
E. faecium

Chemical composition

1. B β-
 72
 2. C
 CP, P,
 NDE,
 P (P<0.05) 39%, 53%, 46%,
 N , CA- P (<10 D)
 1.18%, 6.5-
 I *B. subtilis*
E. faecium AA
 ; (A ,
 A , G.) (P
) L C

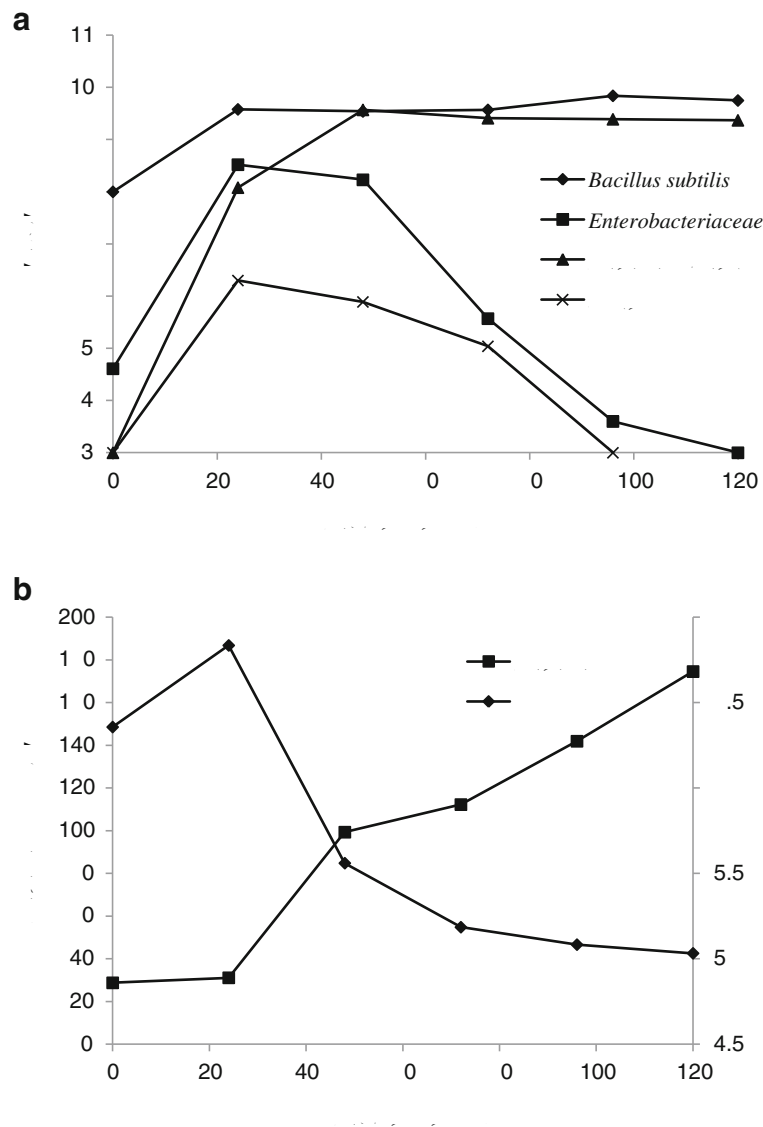


Fig. 2 Microbial composition (log cfu/g) **a**, pH, and lactic acid concentration (mmol/kg) **(b)** in inoculated feed during solid-state fermentation, on a DM basis

... AA ... 7%;
 ...
In vitro amino acid digestibility of the fermented sample
 ... AA ...
 ... 3. I ... CP ... DM ...
 ... (P < 0.05) ... 8% ... 11%,
 ... I ...
 ... 11 ...
 ... (H, I, L, M, ...
 P ...), ... (P < 0.05). N ...
 ... (H, P ...
 C) ... 10%.

Discussion

I ...
 ... E ...
 ... 16, 17.. FLF ...
 ... >9 ... / ... LAB ...
 ... (>150 ... /L), ...
 ... (GI) ...
 ... 18.. A ... FLF ...
 ... 19., ...
 ... 20., ...
 ... 21.. F ... FLF ...
 ... 22.,

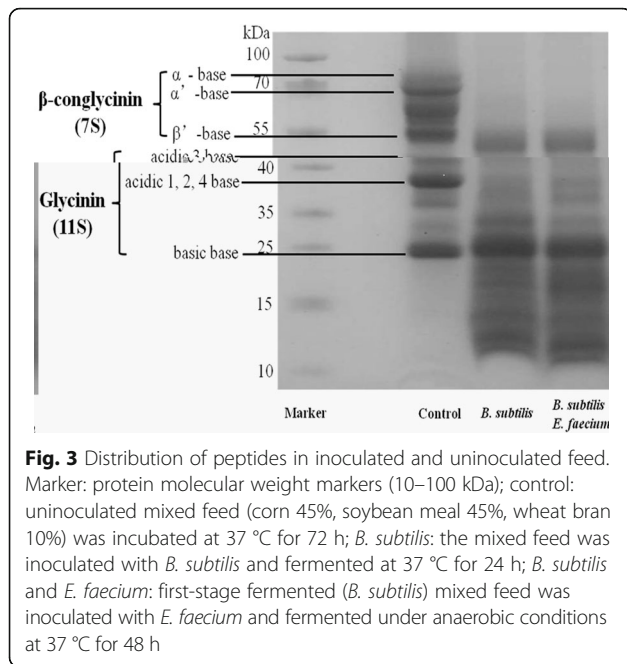


Fig. 3 Distribution of peptides in inoculated and uninoculated feed. Marker: protein molecular weight markers (10–100 kDa); control: uninoculated mixed feed (corn 45%, soybean meal 45%, wheat bran 10%) was incubated at 37 °C for 72 h; *B. subtilis*: the mixed feed was inoculated with *B. subtilis* and fermented at 37 °C for 24 h; *B. subtilis* and *E. faecium*: first-stage fermented (*B. subtilis*) mixed feed was inoculated with *E. faecium* and fermented under anaerobic conditions at 37 °C for 48 h

... I
 ... D.
 ... *B. subtilis* ... LAB
 ... LAB (9.4 ... /.)
 ... FLF. *B. subtilis*
 ... >9.0 ... /.
 ... H ... ,
 (...)
 ... N ... , *Enterobacteriaceae*,
 (...),
 ... FLF ... C ... J ... 26, 27.;
 10. A ... H ... 6.4 6.8 ...
 ... *B. subtilis* ... ,
 ... H ... , α ... α' ... β-

... D.
 ... H ... 6.8 5.0 ...
 ... FLF ... H
 3.8–4.5 23–25.,
 ... O ... H
 FLF ...
 ... H ...
 ... 10. I
 ... *Enterobacteriaceae* ...
 (<3.0 ... /). C ...
 H ...
 ... F ... H ...
 (... *Enterobacteriaceae*) ... GI
 18, 23..
 C - BM
 ... C ... C ...
 ... 60%
 β- ...
 1..
 ... 26, 27.;
 ... 28, 29. I
 ... *B. subtilis* ...
 ... - BM

Table 1 Effect of fermentation on the concentration of soybean antigenic protein, as air-dry basis

Item	Glycinin		β-conglycinin	
	Content, mg/g	Degradation ^a , %	Content, mg/g	Degradation, %
Raw mixed feed	63.74	-	31.76	-
Uninoculated feed ^b	61.02	-	32.15	-
<i>B. subtilis</i> ^c	7.97	86.94	6.98	78.28
<i>B. subtilis</i> and <i>E. faecium</i> ^d	8.47	86.12	7.12	77.53

^aDegradation rate = soybean antigenic protein content in uninoculated feed – soybean antigenic protein content in inoculated feed) / soybean antigenic protein content in uninoculated feed × 100%

^bUninoculated feed: sterile medium was added to mixed feed (45% corn, 45% soybean meal and 10% wheat bran) instead of inoculated bacteria, other experimental procedures were the same as those of inoculated mixed feed

^c*B. subtilis*: mixed feed was inoculated with *B. subtilis* and fermented at 37 °C for 24 h

^d*B. subtilis* and *E. faecium*: first-stage fermented (*B. subtilis*) mixed feed was inoculated with *E. faecium* and incubated under anaerobic conditions at 37 °C for 48 h

Table 2 Analyzed nutrient composition of fermented inoculated and uninoculated feed, as air-dry basis¹

Item	Inoculated feed	Uninoculated feed ²	AA composition	Inoculated feed	Uninoculated feed
DM,%	88.06 ± 1.02	89.09 ± 1.67	Indispensable AA, %		
CP,%	27.61 ± 2.73 ^a	24.03 ± 1.93 ^b	Arg	1.01 ± 0.15 ^b	1.17 ± 0.19 ^a
TCA-SP,%	8.85 ± 1.19 ^a	1.18 ± 0.12 ^b	His	0.58 ± 0.08	0.52 ± 0.11
Fat,%	3.37 ± 0.65	3.67 ± 0.73	Ile	0.78 ± 0.13	0.78 ± 0.16
NDF,%	8.33 ± 0.95 ^b	13.64 ± 0.99 ^a	Leu	1.50 ± 0.18	1.46 ± 0.23
ADF,%	3.58 ± 0.40	3.49 ± 0.76	Lys	1.17 ± 0.08 ^a	0.99 ± 0.10 ^b
Hemicellulose ³ ,%	4.75 ± 0.87 ^b	10.15 ± 0.56 ^a	Met	0.26 ± 0.05	0.23 ± 0.07
Ash,%	4.71 ± 0.51 ^a	3.77 ± 0.38 ^b	Phe	1.78 ± 0.26 ^a	0.86 ± 0.13 ^b
Ca,%	0.18 ± 0.03	0.17 ± 0.02	Thr	0.79 ± 0.12	0.75 ± 0.09
Total P,%	0.55 ± 0.05	0.49 ± 0.07	Val	1.06 ± 0.12	1.04 ± 0.17
Phytate P,%	0.21 ± 0.04 ^b	0.39 ± 0.04 ^a	Dispensable AA,%		
			Asp	1.68 ± 0.10 ^b	1.92 ± 0.17 ^a
			Ser	0.79 ± 0.14	0.75 ± 0.16
			Glu	3.23 ± 0.58	3.49 ± 0.44
			Gly	0.85 ± 0.18	0.80 ± 0.13
			Ala	0.98 ± 0.16	0.94 ± 0.10
			Cys	0.48 ± 0.05 ^a	0.38 ± 0.06 ^b
			Tyr	1.40 ± 0.21 ^a	0.67 ± 0.09 ^b
			Pro	1.09 ± 0.17	1.17 ± 0.21
			Total AA	19.56 ± 2.33	18.12 ± 2.47

¹Values are means of three replicates per treatment. Means in a row without common superscript differ significantly ($P < 0.05$)

²Uninoculated feed: sterile medium was added to mixed feed (45% corn, 45% soybean meal and 10% wheat bran) instead of inoculated bacteria, other experimental procedures were the same as those of inoculated feed

³Hemicellulose = NDF-ADF

14, 27.. ELI A
 β -
 78 88%,
 H
 CP
 AA
 H
 A G
 P L
B. subtilis
B. subtilis
 14..
B. bacillus 62.93%.
 AA L
 30.,
B. subtilis
 ().
 I
 CP, P
 31.,
 14, 32., 33, 34
 35.. ()
 F. H
 F.
 CA- P (8.8%)
 (1.2%). A
 CA- P

Table 3 In vitro CP and AA digestibility (%) of fermented inoculated feed and uninoculated¹

Item	Inoculated feed	Uninoculated feed ²
DM,%	70.60 ± 2.87 ^a	59.33 ± 2.32 ^b
CP,%	86.28 ± 2.23 ^a	78.36 ± 2.04 ^b
Indispensable AA,%		
Arg	82.50 ± 4.65	82.72 ± 3.87
His	84.91 ± 3.70 ^a	74.85 ± 3.46 ^b
Ile	80.49 ± 3.42 ^a	75.62 ± 2.44 ^b
Leu	77.30 ± 3.04 ^a	69.71 ± 2.81 ^b
Lys	84.59 ± 3.91	81.44 ± 3.60
Met	85.30 ± 3.96 ^a	70.31 ± 2.74 ^b
Phe	81.99 ± 4.25 ^a	65.64 ± 3.63 ^b
Thr	78.73 ± 4.12	75.03 ± 3.83
Val	80.74 ± 3.77 ^a	74.49 ± 3.48 ^b
Mean	81.29 ± 4.09 ^a	74.80 ± 3.21 ^c
Dispensable AA,%		
Asp	83.14 ± 5.32	78.54 ± 4.97
Ser	77.86 ± 3.13	74.29 ± 3.74
Glu	85.13 ± 2.47 ^a	80.25 ± 3.02 ^b
Gly	80.78 ± 4.21	76.70 ± 4.08
Ala	84.53 ± 3.38 ^a	75.51 ± 3.66 ^b
Cys	79.74 ± 3.64 ^a	67.90 ± 3.87 ^b
Tyr	81.86 ± 3.43 ^b	72.28 ± 3.10 ^c
Pro	79.31 ± 4.28	75.41 ± 3.94
Mean	82.72 ± 3.11 ^a	77.16 ± 3.04 ^b
Total AA,%	82.15 ± 3.43 ^a	76.07 ± 3.35 ^c

¹Values are means of three replicates per treatment. Means in a row without common superscript differ significantly ($P < 0.05$)

²Uninoculated feed: sterile medium was added to mixed feed (45% corn, 45% soybean meal and 10% wheat bran) instead of inoculated bacteria, other experimental procedures were the same as those of inoculated feed

CA- P (2-20 CA- P AA 38.. D- AA AA NDE, 39.. F, FMF (N P)- F 33., NDE, 14.45%, 43.72%, 86.08%, F. P. P

34. NDF. B. subtilis E. faecium - BM

Conclusions

B. subtilis E. faecium ANF (NDE, CA- P CP F H Enterobacteriaceae. DM CP O F

Additional file

Additional file 1: Strain identification information. (DOCX 1027 kb)

Abb e ia ion

AA: Amino acid; ANFs: Antinutritional factors; Ca: Calcium; CP: Crude protein; FLF: Fermented liquid feed; LAB: Lactic acid bacteria; NDF: Neutral detergent fiber; NSP: Non-starch polysaccharide; P: Phosphorus; SBM: Soybean meal; SDS-PAGE: Sodium dodecyl sulfate – polyacrylamide gel electrophoresis; SSF: Solid state fermentation; TCA-SP: Trichloroacetic acid soluble protein

Ackno ledgmen

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F nding

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A ailabili of da a and ma e ial

The datasets generated and analyzed during the current study are not publicly available. Please contact the authors for data requests.

A ho ' con ib ion

YZW and CYS conceived and designed the experiment. CYS and YZ carried out the experiment, including the solid-state fermentation, chemical analysis, and determination of in vitro digestibility. CYS analyzed the data and wrote the manuscript. ZQL verified the validity and checked the results. All authors read and approved the final version of this manuscript.

Compe ing in e e

The authors declare that they have no competing interests.

Con en fo p blica ion

Not applicable.

E hic app o al and con en o pa icipa e

Not applicable.

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Refe ence

- Li DF, Nelssen JL, Reddy PG, Blecha F, Hancock JD, Allee GL, et al. Transient hypersensitivity to soybean meal in the early-weaned pig. *J Anim Sci.* 1990;68:1790–9.
- Sun P, Li DF, Dong B, Qiao SY, Ma X. Effects of soybean glycinin on performance and immune function in early weaned pigs. *Arch Anim Nutr.* 2008;62:313–21.
- Sun P, Li DF, Li ZJ, Dong B, Wang FL. Effects of glycinin on IgE-mediated increase of mast cell numbers and histamine release in the small intestine. *J Nutr Biochem.* 2008;19:627–33.
- Guggenbuhl P, Simões-Nunes C. Effects of two phytases on the ileal apparent digestibility of minerals and amino acids in ileo-rectal anastomosed pigs fed on a maize rapeseed meal diet. *Livest Sci.* 2007;109:261–63.
- Zhang LY, Li DF, Qiao SY, Wang JT, Bai L, Wang ZY, et al. The effect of soybean galactooligosaccharides on nutrient and energy digestibility and digesta transit time in weaning piglets. *Asian-Aust J Anim Sci.* 2001;14:1598–604.
- Song YS, Pérez VG, Pettigrew JE, Martínez-Villaluenga C, de Mejía EG. Fermentation of soybean meal and its inclusion in diets for newly weaned pigs reduced diarrhea and measures of immunoreactivity in the plasm. *Anim Feed Sci Technol.* 2010;159:41–9.
- Wang Y, Liu XT, Wang HL, Li DF, Piao XS, Lu WQ. Optimization of processing conditions for solid-state fermented soybean meal and its effects on growth performance and nutrient digestibility of weaning pigs. *Livest Sci.* 2014;170:91–9.
- Zhang WJ, Xu ZR, Zhao SH, Sun JY, Yang X. Development of a microbial fermentation process for detoxification of gossypol in cottonseed meal. *Anim Feed Sci Technol.* 2007;135:176–86.
- Shi CY, He J, Yu J, Yu B, Huang Z, Mao XB, et al. Solid state fermentation of rapeseed cake with *Aspergillus niger* for degrading glucosinolates and upgrading nutritional value. *J Anim Sci Biotechnol.* 2015;6:13–20.
- Canibe N, Jensen BB. Fermented liquid feed – microbial and nutritional aspects and impact on enteric diseases in pigs. *Anim Feed Sci Technol.* 2012;173:17–40.
- Ovissipour M, Abedian A, Motamedzadegan A, Rasco B, Safari R, Shahiri H. The effect of enzymatic hydrolysis time and temperature on the properties of protein hydrolysates from Persian sturgeon (*Acipenser persicus*) viscera. *Food Chem.* 2009;115:238–42.
- Nair VC, Duvnjak Z. Reduction of phytic acid content in canola meal by *Aspergillus ficuum* in solid state fermentation process. *Appl Microbiol Biotechnol.* 1990;34:183–88.
- Faurobert M. Application of two-dimensional gel electrophoresis to *Prunus armeniaca* leaf and bark tissues. *Electrophoresis.* 1997;17:170–73.
- Hong KJ, Lee CH, Kim SW. *Aspergillus oryzae* GB-107 fermentation improves nutritional quality of food soybeans and feed soybean meals. *J Med Food.* 2004;7:430–35.
- Sakamoto K, Asano T, Furuya A, Takahashi S. Estimation of in vivo digestibility with the laying hen by an in vitro method using the intestinal fluid of the pig. *Brit J Nutr.* 1980;43:389–91.
- Plumed-Ferrer C, Von Wright A. Fermented pig liquid feed: nutritional, safety and regulatory aspects. *J Appl Microbiol.* 2009;106:351–68.
- Missotten JAM, Michiels J, Olyn A, De Smet S, Dierick NA. Fermented liquid feed for pigs. *Arch Anim Nutr.* 2010;64:437–66.
- van Winsen RL, Uurlings BAP, Lipman LJA, Sniijders JMA, Keuzenkamp D, Verheijden JHM. Effect of fermented feed on the microbial population of the gastrointestinal tracts of pigs. *Appl Environ Microb.* 2001;67:3071–76.
- Lyberg K, Lundh T, Pedersen C, Lindberg JE. Influence of soaking, fermentation and phytase supplementation on nutrient digestibility in pigs offered a grower diet based on wheat and barley. *Anim Sci.* 2006;82:853–58.
- Hong TTT, Thuy TT, Passoth V, Lindberg JE. Gut ecology, feed digestion and performance in weaned piglets fed liquid diets. *Livest Sci.* 2009;125:232–37.
- Missotten JAM, Michiels J, Degroote J, Smet SD. Fermented liquid feed for pigs: an ancient technique for the future. *J Anim Sci Biotechnol.* 2015;6:4.
- Jensen BB, Mikkelsen LL. Feeding liquid diets to pigs. In: Garnsworthy PC, Wiseman J, editors. *Recent Advances in Animal Nutrition*. Nottingham, UK: Nottingham University Press; 1998. p. 107–26.
- Canibe N, Jensen BB. Fermented and non-fermented liquid feed to growing pigs: Effect on aspects of gastrointestinal ecology and growth performance. *J Anim Sci.* 2003;81:2019–31.
- Canibe N, Jensen BB. Fermented liquid feed and fermented grain to piglets- effect on gastrointestinal ecology and growth performance. *Livest Sci.* 2007;108:232–35.
- Canibe N, Miettinen H, Jensen BB. Effect of adding *Lactobacillus plantarum* or a formic acid containing product to fermented liquid feed on gastrointestinal ecology and growth performance of piglets. *Livest Sci.* 2007;114:251–62.
- Aguirre L, Hebert EM, Garro MS, Giori GSD. Proteolytic activity of *Lactobacillus* strains on soybean proteins. *LWT-Food Sci Technol.* 2014;59:780–85.
- Chi CH, Cho SJ. Improvement of bioactivity of soybean meal by solid state fermentation with *Bacillus amyloliquefaciens* versus *Lactobacillus* spp. and *Saccharomyces cerevisiae*. *LWT-Food Sci Technol.* 2016;68:619–25.
- Frias J, Song YS, Martínez-Villaluenga C, González DME, Vidal-Valverde C. Immunoreactivity and amino acid content of fermented soybean products. *J Agric Food Chem.* 2008;56:99–105.
- Feng J, Liu X, Xu ZR, Lu YP, Liu YY. The effect of *Aspergillus oryzae* fermented soybean meal on growth performance, digestibility of dietary components and activities of intestinal enzymes in weaned piglets. *Anim Feed Sci Technol.* 2007;134:295–303.
- Simonen M, Palva I. Protein secretion in *Bacillus* species. *Microbiol Mol Biol Rev.* 1993;57:109–37.
- Hu JK, Lu WQ, Wang CL, Zhu RH, Qiao JY. Characteristics of solid-state fermented feed and its effects on performance and nutrient digestibility in growing-finishing pigs. *Asian-Aust J Anim Sci.* 2008;21:1635–41.
- Chen CC, Shih YC, Chiou PWS, Yu B. Evaluating nutritional quality of single stage- and two stage-fermented soybean meal. *Asian-Aust J Anim Sci.* 2010;23:598–606.
- Chiang G, Lu WQ, Piao XS, Hu JK, Gong LM, Thacker PA. Effects of feeding solid-state fermented rapeseed meal on performance, nutrient digestibility, intestinal ecology and intestinal morphology of broiler chickens. *Asian-Aust J Anim Sci.* 2010;23:263–71.
- Shi C, He J, Yu J, Yu B, Mao XB, Zheng P, et al. Amino acid, phosphorus, and energy digestibility of *Aspergillus niger* fermented rapeseed meal fed to growing pigs. *J Anim Sci.* 2015;93:2916–25.
- Sun H, Tang JW, Yao XH, Wu XF, Wang X, Feng J. Improvement of the Nutritional Quality of Cottonseed Meal by *Bacillus subtilis* and the Addition of Papain. *Int J Agric Biol.* 2012;14:563–68.
- Rozan P, Guillaume C, Bau HM, Schwertz A, Nicolas JP, Mejean L. Detoxication of rapeseed meal by *Rhizopus oligosporus* sp-T3: A first step towards rapeseed protein concentrate. *Int J Food Sci Technol.* 1996;31:85–90.
- Stokes JL, Gunness M. The amino acid composition of microorganisms. *J Bacteriol.* 1946;52:195–207.
- Kuchroo CN, Fox PF. Soluble nitrogen in cheddar cheese: Comparison of extraction procedures. *Milchwissenschaft.* 1982;37:331–35.
- Gilbert ER, Wong EA, Webb KE. Peptide absorption and utilization: Implications for animal nutrition and health. *J Anim Sci.* 2008;86:2135–55.